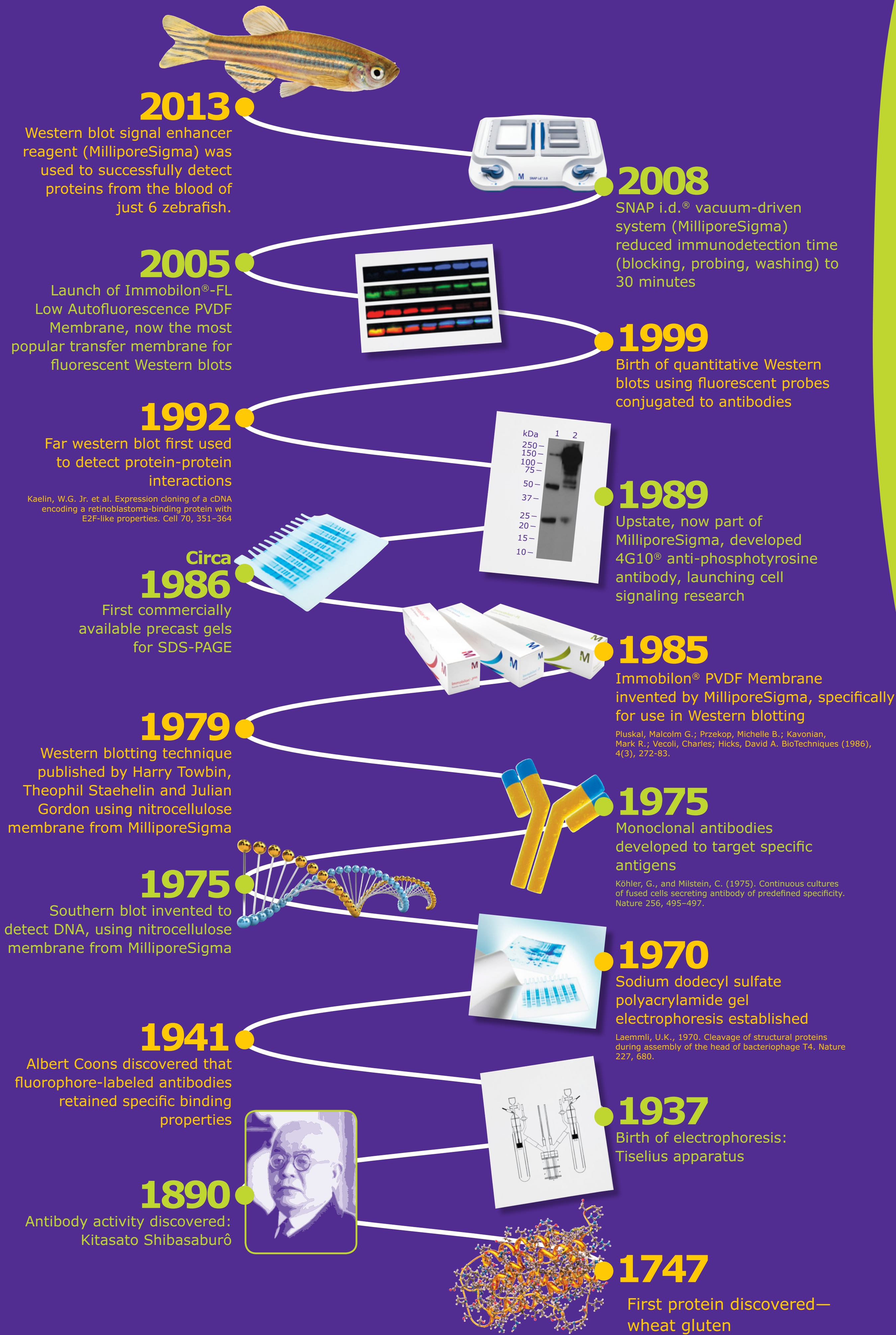


# HISTORY OF WESTERN BLOTTING

**MILLIPORE SIGMA**



## TIMELESS Recipes and Solutions for Blotting Success

### Standard recipes for Western blotting

**TBS 10x (concentrated Tris-buffered saline)**  
200 mM Tris  
1500 mM NaCl

**For 1 L:**  
24 g Tris base (formula weight: 121.1 g)  
88 g NaCl (formula weight: 58.4 g)  
Dissolve in 900 mL Milli-Q<sup>®</sup> water  
pH to 7.6 with 12 N HCl  
Add Milli-Q<sup>®</sup> water to a final volume of 1 L

**1X solution:** mix 1 part of the 10X solution with 9 parts Milli-Q<sup>®</sup> water and adjust pH to 7.6.

**TBST (Tris-buffered saline, 0.1% Tween<sup>®</sup> 20)**

**For 1 L:**  
100 mL of TBS 10X  
900 mL of Milli-Q<sup>®</sup> water  
1 mL Tween<sup>®</sup> 20

**4X SDS-PAGE sample loading buffer**  
1.5 mL of 1 M Tris-HCl pH 6.8  
3 mL of 1 M DTT (dithiothreitol)  
0.6 g of SDS (sodium dodecyl sulfate)  
0.03 g of bromophenol blue  
2.4 mL of glycerol  
Bring final volume to 7.5 mL with Milli-Q<sup>®</sup> water  
If solution is orange/yellow in color, add 1 drop of 5 M NaOH to adjust pH  
Make 500 µL aliquots and store at -20°C

**SDS-PAGE gel making buffer 1.5 M Tris-HCl (for separating gel)**

118.2 g of Tris-HCl,  
Add 450 mL Milli-Q<sup>®</sup> water and adjust pH to 8.8  
Add Milli-Q<sup>®</sup> water to final volume 500 mL  
Filter and degas

**SDS-PAGE gel making buffer 1 M Tris-HCl (for stacking gel)**

78.8 g of Tris-HCl  
Add 450 mL Milli-Q<sup>®</sup> water and adjust pH to 6.8  
Add Milli-Q<sup>®</sup> water to final volume 500 mL  
Filter and degas

**SDS-PAGE 10X gel running buffer**

60 g Tris base (248 mM)  
288 g glycine (1.92 M)  
20 g SDS (1% w/v)  
Milli-Q<sup>®</sup> water to final volume 2 L  
No need to pH, filter, or degas  
Dilute to 1X for running SDS-PAGE gels

**10X PBS (1 L)**  
81.8 g NaCl (1.4 M)  
20.1 g KCl (270 mM)  
14.2 g Na<sub>2</sub>HPO<sub>4</sub> (100 mM)  
2.45 g KH<sub>2</sub>PO<sub>4</sub> (18 mM)  
pH 7.4  
Add Milli-Q<sup>®</sup> water to final volume of 1 L

**Transfer buffer 10X**  
15.2 g Tris base  
72.1 g glycine  
5.0 g SDS  
Milli-Q<sup>®</sup> water to final volume 500 mL

**Transfer buffer 1X (make fresh each time)**

50 mL 10X transfer buffer  
100 mL methanol  
Milli-Q<sup>®</sup> water to final volume 500 mL

**SDS-PAGE Coomassie<sup>®</sup> staining solution**

1.25 g Coomassie<sup>®</sup> blue R-250  
225 mL methanol  
225 mL Milli-Q<sup>®</sup> water  
50 mL glacial acetic acid

**SDS-PAGE destaining solution**

300 mL methanol (30%)  
100 mL acetic acid (10%)  
600 mL Milli-Q<sup>®</sup> water

### Recipes for TruPAGE<sup>™</sup> Precast Gel System

**20X TruPAGE<sup>™</sup> TEA-Tricine SDS Running Buffer**

179.0 g triethanolamine  
143.3 g tricine  
20 g SDS  
Milli-Q<sup>®</sup> water to final volume 1.0 L  
The pH should be between 8.2-8.3 at 25 °C.

**20X TruPAGE<sup>™</sup> Tris-MOPS SDS Express Running Buffer**

145.2 g Tris base  
143.3 g MOPS  
20 g SDS  
Milli-Q<sup>®</sup> water to final volume 1.0 L  
The pH should be between 8.2-8.3 at 25 °C.  
*NOTE: Antioxidant (5 mM sodium bisulfite) is required in the inner chamber if using this buffer.*

**4X TruPAGE<sup>™</sup> LDS Sample Buffer**

4.0 g glycerol  
0.40 g lithium dodecyl sulfate  
1.2 g triethanolamine  
0.40 g Ficol<sup>®</sup> 400  
2.5 mg Phenol Red  
2.5 mg Brilliant Blue G250  
7.0 mg EDTA  
Milli-Q<sup>®</sup> water to final volume 10 mL  
The pH should be between 7.7-7.8 at 25 °C.  
Dissolve triethanolamine (0.8 M) and pH to 7.6 before adding other reagents.

**800X TruPAGE<sup>™</sup> Running Antioxidant**

4.2 g Sodium Bisulfite  
Milli-Q<sup>®</sup> water to final volume 10 mL

**20X TruPAGE<sup>™</sup> Transfer Buffer**

30.3 g Tris base  
144.1 g glycine  
Milli-Q<sup>®</sup> water to final volume 1.0 L.

**10X Dithiothreitol sample reducer**

38.6 mg DTT  
Milli-Q<sup>®</sup> water to final volume 1.0 mL

#### Prepare and Grow

- Cells, media, sera, cultureware
- Stericup<sup>®</sup> media filtration devices



Precise Westerns start with low-binding, high recovery Stericup<sup>®</sup> vacuum filters – for media volumes of 150 mL – 1,000 mL

#### Collect and Extract

- ProteoExtract<sup>®</sup> kits for fractionation
- CytoBuster<sup>™</sup>, BugBuster<sup>™</sup> and CellLytic<sup>™</sup> protein extraction reagents
- Benzonase<sup>®</sup> nuclease
- cOmplete<sup>™</sup> and SIGMAFAST<sup>™</sup> protease inhibitors

Maintain and preserve protein functionality following cell lysis using individual protease inhibitors and protease inhibitor cocktails.



#### Enrich and Concentrate

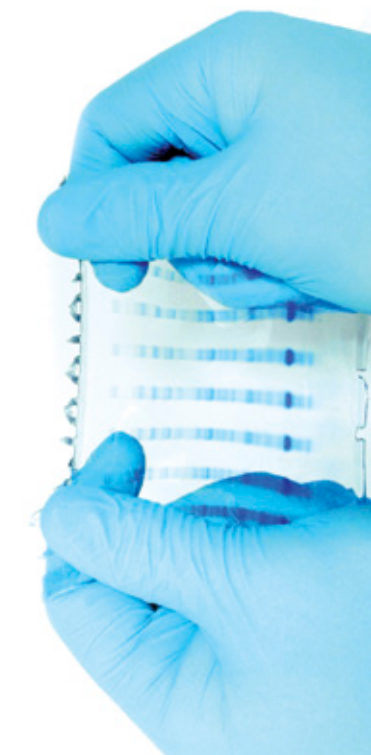
- Amicon<sup>®</sup> filters for concentration and buffer exchange
- PureProteome<sup>®</sup> magnetic beads for enrichment and purification
- Recombinant protein purification affinity gels



Unlike methods that require careful aspiration to avoid sample loss, PureProteome<sup>®</sup> magnetic beads are isolated using a magnetic rack, for total removal of buffers and complete recovery with no sample dilution.

#### Qualify and Electrophorese

- TruPAGE<sup>™</sup> precast gels
- Buffers (including Trizma<sup>®</sup> Tris base), solvents, prestained MW markers
- Direct Detect<sup>®</sup> IR spectrometer for protein sample qualification



TruPAGE<sup>®</sup> precast gels are tear-resistant to reduce handling failures that commonly occur during gel electrophoresis and downstream processing.

#### Blot and Analyze

- Range of Immobilon<sup>®</sup> PVDF transfer membranes
- Western-validated antibodies
- SNAP i.d.<sup>®</sup> 2.0 protein detection system
- Immunodetection reagents



Blot efficiently using our Western blotting reagents, including Blok<sup>®</sup> noise-cancelling reagents, Luminata<sup>™</sup> Western HRP substrates, Immunoreaction Enhancer Reagents, and ReBlot<sup>™</sup> Plus antibody stripping kits, all carefully optimized for your success.