

Technical Bulletin

## Fructose Assay Kit

**Catalogue number MAK519**

### Product Description

Fructose (C<sub>6</sub>H<sub>12</sub>O<sub>6</sub>, also called levulose or laevulose), is a monosaccharide found in honey, tree fruits, berries, melons, and some root vegetables along with glucose and galactose. The human body can use fructose for energy, however, too much consumption may lead to high triglycerides. Simple, direct and high-throughput assays for fructose determination find wide applications. The reagent system reacts directly and specifically with fructose to form a colored product. Glucose and galactose do not interfere. The color intensity at 565 nm is directly proportional to the fructose concentration in the sample.

The linear detection range of the kit is 12 to 1000 µM fructose. The kit is suitable for determination of fructose in food, juice, beverage, other agricultural products, and biological samples such as serum, plasma, urine, saliva, milk, culture medium. It is also used to study the effects of drugs on fructose metabolism.

### Components

The kit is sufficient for 100 colorimetric assays in 96-well plates.

• Assay Buffer	10 mL
• Enzyme	1 vial
• Enzyme Buffer	150 µL
• PMS Solution	1.5 mL
• MTT Solution	1.5 mL
• Standard (20 mM D-Fructose)	400 µL

### Reagents and Equipment Required but Not Provided

- Pipetting devices and accessories (For example - multichannel pipettor)
- Spectrophotometric multi-well plate reader.
- Clear bottom 96-well plates (For example - Corning Costar). Cell culture or tissue culture treated plates are not recommended.
- 1.5 mL centrifuge tubes
- 6 N Hydrochloric acid
- 6 N Sodium hydroxide

### Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices. Storage/Stability

The kit is shipped on wet ice. Store all components at -20 °C.

## Preparation Instructions

Briefly centrifuge small vials prior to opening. Equilibrate all components to room temperature.

Enzyme: Reconstitute Enzyme by adding 120  $\mu\text{L}$  Enzyme Buffer to the Enzyme tube. Make sure Enzyme is fully dissolved by pipetting up and down. Store the reconstituted Enzyme at  $-20\text{ }^{\circ}\text{C}$  and use within 2 months. Keep thawed tubes on ice during assay.

Note:

(1) The following substances interfere and should be avoided in sample preparation: ascorbic acid, SDS ( $> 0.2\%$ ), sodium azide, NP-40 ( $> 1\%$ ) and Polysorbate-20 ( $> 1\%$ ).

(2) This assay is based on a kinetic reaction. To ensure identical incubation time, addition of Working Reagent to standard and samples should be quick, and mixing should be brief but thorough. Use of a multi-channel pipettor is recommended.

## Procedure

All Samples and Standards should be run in duplicates.

### Sample Preparation

- Liquid Samples

Assay can be performed directly on Samples such as serum, plasma and fruit juices can be assayed directly. Fruit juices may contain high concentrations of fructose, it is recommended to dilute juice Sample 50-fold (Dilution Factor (DF)= 50) in purified water prior to assay.

- Milk Samples

1. Clear the milk Samples by mixing 600  $\mu\text{L}$  milk with 100  $\mu\text{L}$  6 N HCl.
2. Centrifuge at 14,000 rpm for 5 min.
3. Transfer 300  $\mu\text{L}$  supernatant into a clean tube and neutralize with 50  $\mu\text{L}$  6N NaOH. The neutralized supernatant is ready for assay (DF = 1.36).

Transfer 20  $\mu\text{L}$  of the Sample into two separate wells.

Note:

1. It is prudent to test several dilutions to determine an optimal dilution factor (DF).

### Standard curve preparation:

Prepare 1000  $\mu\text{M}$  Standard by mixing 12  $\mu\text{L}$  20 mM Standard and 228  $\mu\text{L}$  purified water. Dilute Standards as described in the Table 1.

**Table 1.** Dilution of Fructose Standards

Well No.	1000 $\mu\text{M}$ Fructose Standard	Purified Water ( $\mu\text{L}$ )	Fructose ( $\mu\text{M}$ )
1	100 $\mu\text{L}$	0	1000
2	60 $\mu\text{L}$	40	600
3	30 $\mu\text{L}$	70	300
4	0 $\mu\text{L}$	100	0

Transfer 20  $\mu\text{L}$  of the diluted Standards into separate wells of a clear, flat-bottom 96-well plate.

### Working Reagents

Keep Working Reagent protected from light. Do not expose Working Reagent to light for more than 5 minutes.

1. Mix enough reagents for the number of assays to be performed. Prepare Working Reagents according to Table 2.

**Table 2.** Preparation of Working Reagents for 96-well plate assay.

Reagent	Working Reagent
Assay Buffer	56 $\mu\text{L}$
Reconstituted Enzyme	1 $\mu\text{L}$
PMS Solution	14 $\mu\text{L}$
MTT Solution	14 $\mu\text{L}$

2. Add 80  $\mu\text{L}$  of Working Reagent to the Standard and Sample wells.
3. Tap plate to mix briefly and thoroughly.

### Measurement

1. Incubate for 60 minutes at room temperature, protected from light.
2. Read optical density at 565 nm (520-600 nm).

## Results

1. Subtract the Blank value (Well #4) from the standard values.
2. Plot  $\Delta OD$  of the Standard against Standard concentrations.
3. Determine slope and calculate the concentration of fructose in the Sample using the following equation:

$$(\mu\text{M}) = \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{Slope } (\mu\text{M}^{-1})} \times \text{DF}$$

where:

$\text{OD}_{\text{Sample}}$  = Sample Optical Density reading

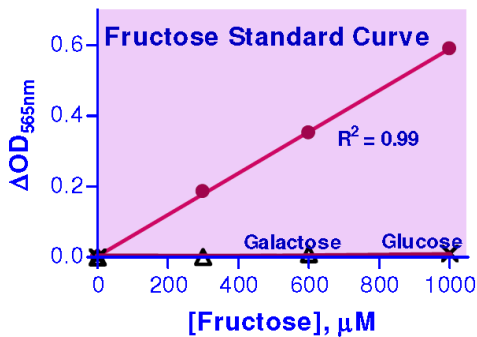
$\text{OD}_{\text{Blank}}$  = Optical Density reading of blank (well #4)

DF = Sample Dilution Factor (DF = 1 for undiluted Samples)

Note: If the calculated fructose concentration of a sample is higher than 1000  $\mu\text{M}$ , dilute sample in water and repeat the assay. Multiply result by the Dilution Factor, DF.

### Figure 1.

Typical Colorimetric Fructose Standard Curve



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