

## Product Information

### ADME/Tox Cell Lines Canine MDCKII Wild Type Cells 1 Vial 2-6 Million Cells

Catalog Number **MTOX1300**

Store at  $-130\text{ }^{\circ}\text{C}$  or below in liquid nitrogen vapor phase

## TECHNICAL BULLETIN

### Product Description

MDCK II - Madin-Darby canine kidney- is a subclone derived from the heterogenous parent line - MDCK (ECACC catalogue no. 85011435). MDCK II cells, which predominate in later passages from MDCK, are reported to display electrical resistance of  $100\text{ ohm/cm}^2$ . This strain is thought to be derived from the distal tubule or collecting duct of the nephron. The cell line can be used as an experimental model to study the generation and maintenance of cell surface polarity in epithelial cells

ATP-binding cassette (ABC) transporters are a family of transmembrane proteins that utilize ATP hydrolysis for translocation of substrates across membranes. ABC transporters are known to play a critical role in the development of multidrug resistance. Evaluation of membrane transporter pharmacology in drug disposition and drug-drug interactions (DDI) is critical to the pharmaceutical safety evaluations of new drug entities. Selection of the targeted gene(s) was based on the considerable body of evidence supporting its crucial role in the development of multidrug resistance.<sup>8</sup>

### Components

This product contains 1 cryovial of Canine MDCKII Wild Type Cells which have been frozen at a density of 2-6 million cells. The vial contains sufficient cells to seed one 24 or 96 well Millicell® plate.

### Cell Line Description

Parental Cell Line: ECACC Cat. No. 00062107.

Note: Please see 00062107 product datasheet from ECACC for additional information about the origin of this cell line. Cytogenetic information is based on initial seed stock at Sigma Life Science. Cytogenetic instability has been reported in the literature for some cell lines

### Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

### Storage/Stability

Upon receiving a shipment of frozen cells it is important the end user gives the shipment attention without delay. To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at  $-70\text{ }^{\circ}\text{C}$ . Storage at  $-70\text{ }^{\circ}\text{C}$  will result in loss of viability.

Precaution: It is recommended that protective gloves and clothing always be used, and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to the gas phase may result in the rapid expansion of the vessel, potentially blowing off its cap with dangerous force creating flying debris. At the time a cell line is ordered, end users should also consider the culture conditions for the new cell line and make sure the appropriate measures are taken.

## Protocol for Thawing and Seeding 24 Well or 96 well plates

**Note:** One cryovial contains enough cells to seed one plate.

### Reagents and Equipment Required but Not Provided

**Note:** Neither media nor supplements are supplied with the vials. These must be obtained prior to receiving the vials.

- Fetal Bovine Serum (FBS), Catalog No. F4135
- Minimum Essential Medium Eagle (MEM), Catalog No. M2279
- L-Glutamine solution, Catalog No. G7513
- Penicillin-Streptomycin, Catalog No. P4333
- Millicell® 24-well plate assembly, Catalog Number PSRP010R5
- Millicell® 24-well receiver tray, Catalog Number SPMW010R
- Millicell® 96-well plate assembly, Catalog Number PSRP004
- Millicell® 96-well receiver tray, Catalog Number SPMW004R

### MDCKII Medium Preparation

Prepare Medium by adding FBS, Catalog No. F4135, at a final concentration of 10% (v/v) in MEM, Catalog No. M2279.

MEM is supplemented with L-Glutamine, Catalog No. G7513, to a final concentration of 2 mM.

Penicillin-Streptomycin, Catalog No. P4333, is added to MEM at a final concentration of 1% (v/v). This medium is formulated for use with a 5% CO<sub>2</sub> in air atmosphere.

### Protocol for Thawing and Seeding Millicell® Plates

1. Pre-warm MDCKII Medium in 37 °C water bath.
2. Pipette 10 mL of pre-warmed, medium into a sterile centrifuge tube.
3. Prepare an absorbent paper with 70% ethyl alcohol.
4. Remove cryovial from liquid nitrogen. Under a laminar flow hood, briefly twist the cap a quarter turn to relieve the internal pressure and then close again.
5. Quickly transfer the cryovial to a 37 °C water bath. While holding the tip of the vial, gently agitate for 1–2 minutes, being careful not to allow water to penetrate the cap. **Note: Do not submerge cryovial completely.**
6. Watch the cryovial closely. When just a small crystal of ice remains, remove it from the water bath.
7. Wipe the outside of the vial with 70% ethyl alcohol absorbent paper and place it under laminar flow hood.

8. Aseptically transfer the cell suspension to the centrifuge tube with pre-warmed medium and pellet cells by centrifugation at 800 RPM for three minutes.
9. Aspirate the media from the tube, and re-suspend the cell pellet in the appropriate amount of pre-warmed medium (13 mL for 24 well plates or 11 mL for 96 well plates).
10. Mix cells thoroughly and aliquot 500 µL per well (Millicell® 24-well plate assembly, Catalog Number PSRP010R5 ) or 100 µL per well (Millicell® 96-well plate assembly, Catalog Number PSRP004).
11. Add 25 ml media (Millicell® 24-well receiver tray, Catalog Number SPMW010R) or 5 ml media (Millicell® 96-well receiver tray, Catalog Number SPMW004R) to the bottom receiver tray of plate assembly.
12. Place the plate assembly in a 37 °C, 5% CO<sub>2</sub> incubator. Incubate the plate assembly until the cells form a confluent monolayer, replacing the medium in both the wells and receiver tray every 3-4 days. Assays may be performed post-confluent cells from days 2-5.

### References

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2. Cong, L., et al., Multiplex genome engineering using CRISPR/Cas systems. Science 2013; 339(6121):819-23.
3. Friedland, A. E., et al, Heritable genome editing in C. elegans via a CRISPR-Cas9 system. Nat Methods. 2013 Jun 30. doi: 10.1038/nmeth.2532.
4. Hwang, W.Y., et al., Efficient genome editing in zebrafish using a CRISPR-Cas system. Nat Biotechnol. 2013; 31(3): 227-9.
5. Jinek, M., et al., A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. Science 2012; 337(6096): 816-21.
6. Jinek, M., et al., RNA-programmed genome editing in human cells. Elife 2013; 2:e00471. doi: 10.7554/eLife.00471.
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8. The International Transporter Consortium (2010 White Paper), Membrane transporters in drug development. Nature Reviews Drug Discovery, 9,215-236 (2010).
9. Chen, W., et al. in Cell Culture Models of Biological Barriers In-Vitro Test Systems for Drug Absorption and Delivery. (Lehr, C-M., ed.), Taylor & Francis, (New York, NY: 2002) pp. 143-163.

Additional product and technical information at [www.sigma.com](http://www.sigma.com).  
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