

Restriction Endonuclease Bsm I

From Bacillus stearothermophilus NUB36

Cat. No. 11 292 307 001

200 units (10 U/μl)



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Store at -15 to -25°

Stability/Storage

The undiluted enzyme solution is stable when stored -15 to -25°C until the control date printed on the label. Do not store below −25°C to avoid freezing. Note: Product is shipped on dry ice

Sequence specificity

GAATGCN/N Bsm I recognizes the sequence CTTAC/GNN

and generates fragments with 3'-cohesive termini (1).

Compatible ends

The enzyme is not known to generate compatible ends.

Isoschizomers

The enzyme is not known to have isoschizomers.

Methylation sensitivity

Bsm I is inhibited by the presence of 6-methyladenine, as indicated (*). 5-Methylcytosine does not inhibit (°).

Storage buffer

10 mM Tris-HCl, 50 mM KCl, 0.1 mM EDTA, 1 mM Dithiothreitol, 200 µg/ml bovine serum albumin, 50% Glycerol (v/v), pH approx. 7.4 (at +37°C).

Incubation buffer (10x, included)

500 mM Tris-HCl, 1 M NaCl, 100 mM MgCl₂, 10 mM Dithioerythritol, pH 7.5 (at +37°C) (= SuRE/Cut Buffer H)

Activity in SuRE/Cut Buffer System

Bold face printed buffer indicates the recommended buffer for optimal activity:

Α	В	L	М	Н
0-10%	50-75%	0-10%	25-50%	100%

Incubation temperature +65°C

Unit definition

One unit is the enzyme activity that completely cleaves 1 μg λDNA in 1 h at +65° C in a total volume of 25 μl incubation buffer H.

Typical experiment

Component	Final concentration
DNA	1 μg
10 × SuRE/Cut Buffer H	2.5 μl
Repurified water	Up to a total volume of 25 μl
Restriction enzyme	1 unit

Incubate at +65°C for 1 h

Heat inactivation

There is no information about Bsm I and heat inactivation available

Number of cleavage sites on different DNAs (2):

λ	Ad2	SV40	Φ X174	M13mp7	pBR322	pBR328
46	10	4	3	1	1	3

Activity in PCR buf-Relative activity in PCR mix (Taq DNA Polymerase buffer) is 0%. The PCR mix contained λ target DNA, primers, 10 mM Tris-HCl (pH 8.3, +20°C), 50 mM KCl, 1.5 mM MgCl_2 , 200 $\mu\mathrm{M}$ dNTPs, 2.5 U Taq DNA polymerase. The mix was subjected to 25 amplification cvcles.

Troubleshooting

A critical component is the DNA substrate. Many compounds used in the isolation of DNA such as phenol, chloroform, ethanol, SDS, high levels of NaCl, metal ions (e.g., Hg²⁺, Mn²⁺) inhibit or alter recognition specificity of many restriction enzymes. Such compounds should be removed by ethanol precipitation followed by drying, before the DNA is added to the restriction digest reaction. Appropriate mixing of the enzyme is recommended.

Quality control

Lot-specific certificates of analysis are available at www.lifescience.roche.com/certificates

Absence of unspecific endonuclease activities

1 μg λDNA is incubated for 16 h in 50 μl incubation buffer with excess of Bsm I. The number of enzyme units which do not change the enzyme-specific pattern is stated in the certificate of analysis.

Absence of exonuclease activity

Approx. 5 µg [3H] labeled calf thymus DNA are incubated with 3 µl Bsm I for 4 h at +37°C in a total volume of 100 μl 50 mM Tris-HCl, 10 mM MgCl₂, 1 mM Dithioerythritol, pH approx. 7.5. The release of radioactivity is calculated as a percentage value of liberated to input radioactivity per unit of enzyme (stated in the certificate of analysis).

Ligation and recutting assay Bsm I fragments obtained by complete digestion of 1 μg λDNA are ligated with 1 U T4-DNA ligase in a volume of 10 µl by incubation for 16 h at +4°C in 66 mM Tris-HCl, 5 mM MgCl₂, 5 mM Dithiothreitol, 1 mM ATP, pH 7.5 (at 20°C).

The percentage of ligation and subsequent recutting with Bsm I yielding the typical pattern of $\lambda DNA \times Bsm$ I fragments is determined and stated stated in the certificate of analysis.

References

- Kessler, C. & Manta, V. (1990) Gene 92, 1-250.
- Inagaki, K. et al. (1990) "Isolation and identification of restiction endonuclease Asp35HI from Acidiphilium species 35H" Nucl. Acids Res. **18**, 6155; Rebase The Restriction Enzyme Database:
- Benchmate: http://roche-applied-science.com/benchmate

Ordering Information

Product	Application	Packsize	Cat. No.
Rapid DNA Liga- tion Kit	Ligation of sticky- or blunt-ended DNA fragments in just 5 min at 15 - 25 °C.	Kit (40 DNA ligations)	11 635 379 001
T4 DNA Ligase	Ligation of sticky- and blunt- ended DNA fragments.	100 U 500 units (1 U/μl)	10 481 220 001 10 716 359 001
rAPid Phosphatase	Dephosphorylation of 5'-phosphate residues from nucleic acids	1000 U 5000 U	04 898 133 001 04 898 141 001
rAPid Dephos and Ligation Kit	Dephosphorylation of nucleic acids.	40 reactions 160 reactions	04 898 117 001 04 898 125 001
Alkaline Phospha- tase (AP), special quality for molecu- lar biology	Dephosphorylation of 5'-phosphate residues from nucleic acids.	1000 U (20 U/µl)	11 097 075 001
Agarose MP	Multipurpose agarose for analytical and prepara- tive electrophoresis of nucleic acids	100 g 500 g	11 388 983 001 11 388 991 001
Agarose LE	Separation of nucleic acids in the range 0.2 - 1.5 kbp	100 g 500 g	11 685 660 001 11 685 678 001
Agarose Gel DNA Extraction Kit	For the elution of DNA fragments from agarose gels.	1 Kit (max. 100 reactions)	11 696 505 001
High Pure PCR Product Purifica- tion Kit	Purification of PCR or enzymatic modification reaction (e.g. restriction digest)	50 purifications 250 purifications	11 732 668 001 11 732 676 001
SuRE/Cut Buffer Set for Restriction Enzymes	Incubation buffers A, B, L, M and H for restriction enzymes	1 ml each (10× conc. solutions)	11 082 035 001
SuRE/Cut Buffer A	Restriction enzyme incubation	5×1 ml ($10 \times$ conc. solution)	11 417 959 001
SuRE/Cut Buffer B	Restriction enzyme incubation	5×1 ml ($10 \times$ conc. solution)	11 417 967 001
SuRE/Cut Buffer H	Restriction enzyme incubation	5×1 ml ($10 \times$ conc. solution)	11 417 991 001
SuRE/Cut Buffer L	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 975 001
SuRE/Cut Buffer M	Restriction enzyme incubation	5×1 ml ($10 \times$ conc. solution)	11 417 983 001
Water, PCR Grade	Specially purified, double-distilled, deionized, and autoclaved	100 ml (4 vials of 25 ml) 25 ml (25 vials of 1 ml) 25 ml	03 315 843 001 03 315 932 001 03 315 959 001
BSA, special qual- ity for molecular biology	Maintaining enzyme stability	(1 vial of 25 ml) 20 mg (1 ml)	10 711 454 001

Changes to previous version

Editorial changes

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Commonly used bacterial strains

Strain	Genotype
BL21	<i>E. coli B F</i> $^-$ <i>dcm ompT hsdS(r_B- m_B-) gal</i> (Studier, F.W. <i>et al</i> (1986) <i>J. Mol. Biol.</i> , 189 , 113.)
C600 ^e	supE44 hsdR2 thi-1 thr-1 leuB6 lacY1 tonA21; (Hanahan, D. (1983) J. Mol. Biol. 166 , 557.)
DH5α	supE44 Δ(lacU169 (φ80d/acZΔM15) hsdR17 recA1 endA1 gyrA96 thi-1 relA1; (Hanahan, D. (1983) J. Mol. Biol. 166 , 557.)
HB101	supE44 hsdS20 recA13 ara-14 proA2 lacY1 galK2 rpsL20 xyl-5 mtl-1; (Hanahan, D., (1983) J. Mol. Biol. 166 , 557.)
JM108	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi Δ (lac-proAB); (Yanisch- Perron, C. et al., (1985) Gene 33 , 103.)
JM109	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi Δ (lac-proAB) F[traD36proAB ⁺ , lacl ^q lacZ Δ M15]; (Yanisch- Perron, C. et al., (1985) Gene 33 , 103.)
JM110	rpsL (Str ^f) thr leu thi-I lacY galK galT ara tonA tsx dam dcm supE44 Δ (lac-proAB) F[traD36proAB ⁺ , lacf ^f lacZ Δ M15]; (Yanisch- Perron, C. et al., (1985) Gene 33 , 103.)
K802	supE hsdR gal metB; (Raleigh, E. et al., (1986) Proc.Natl. Acad.Sci USA, 83, 9070.; Wood, W.B. (1966) J. Mol. Biol., 16 , 118.)
SURE ^r	recB recJ sbc C201 uvrC umuC::Tn5(kan') lac , Δ(hsdRMS) endA1 gyrA96 thi relA1 supE44 F'[proAB ⁺ lacI ^q lacZΔM15 Tn10 (tet'); (Greener, A. (1990) Stratagies, 3 , 5.)
TG1	supE hsd Δ5 thi Δ(lac-proAB) F[traD36proAB ⁺ , lacl ^q lacZΔM15]; (Gibson, T.J. (1984) PhD Theses. Cambridge University, U.K.)
XL1-Blue ^r	supE44 hsdR17 recA1 endA1 gyrA46 thi relA1 lac F'[proAB ⁺ , lacl ^q lacZ∆M15 Tn10 (tet ^{f)}]; (Bullock et al., (1987) BioTechniques, 5, 376.)

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