



93349 93352 Trizma[®] Base ('TRIS Buffer', 2-Amino-2-hydroxymethyl-1,3-propanediol, Tris(hydroxymethyl)aminomethane, THAM, Trometamol)

CAS number: 77-86-1

Product Description:

Appearance: Clear colorless to very faint yellow

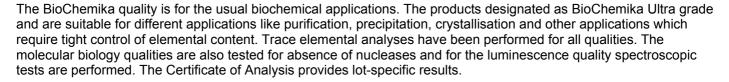
Molecular formula: $NH_2C(CH_2OH)_3$ Formula weight: 121.14 g/molMp: $168 - 172^{\circ}C^1$

Solubility: 4 M in H₂O, 20°C, complete, colorless

pH: 10.5-12.0 (4 M in H₂O, 25°C)

pK_a(20 °C): 8.3¹

93352 BioChemika 93349 BioChemika Ultra



Trizma is a registered trademark of Sigma-Aldrich for tris(hydroxymethyl)aminomethane, commonly called Tris.

Additional information on temperature and concentration effects, and on the use of pH electrodes with Trizma buffers is available from the technical service. Tris is an established basimetric standard and buffer used in biochemistry and molecular biology. It may be used by itself as a buffer or as a component of mixed buffer formulations. These different buffer formulations include:

- Tris-EDTA (TE) buffer
- Tris magnesium buffer
- Tris-acetate-EDTA (TAE) buffer
- Tris-borate-EDTA (TBE) buffer
- Tris-buffered saline (TBS)
- Tris-buffered saline with dextrose (TBS-D)
- Tris-glycine buffer
- Tris-phosphate EDTA buffer
- · Tris-SDS buffer
- Tris-sucrose
- Tris-Tricine-SDS buffer

Tris salts are used in protein crystallization at various pH values (see crystallization kits 82009, 70437, 75403, 86684, 73513). 3,4,5,6 The use of low-ionic strength Tris buffers in the formation of intermediate filaments of lamin from *Caenorhabditis elegans* has been described. Tris has been utilized in studies of double stranded complexes of peptide nucleic acids (PNA) and their complementary DNA sequences, by use of anion exchange HPLC. The use of Tris in capillary electrochromatography and UV analysis of tocopherols and tocotrienols has been reported.



Preparation Instructions:

This product is soluble in water (4 mol/l), yielding a clear, colorless solution. When preparing Tris solutions at a given pH and temperature, it is necessary to choose the proper

mixture to give the desired final pH at the desired temperature.

For precise applications, use a carefully calibrated pH meter with a glass/calomel combination electrode.

Storage/Stability:

Trizma solutions can be autoclaved. Tris has a significant temperature coefficient:

- From 5 °C to 25 °C, the pH decreases an average of 0.03 pH units per °C.
- From 25 °C to 37 °C, the pH decreases an average of 0.025 pH units per °C.

Precautions and Disclaimer:

For Laboratory Use Only. Not for drug, household or other uses.

References

- Gomori, G., Preparation of Buffers for Use in Enzyme Studies. Methods Enzymol., 1, 138-146 (1955).
- 2. Molecular Cloning: A Laboratory Manual, 3rd ed., Sambrook, J. and Russell, D.W., CSHL Press (Cold Spring Harbor, NY: 2001), pp. 5.8, 5.30, 5.43, 5.60, 5.76, 10.25-10.26, 12.75, 12.84, 12.87, 13.52, 16.29-16.31, A1.2-1.3, A1.7-A1.8, A1.17-A1.18, A1.22, A8.42-A8.43.
- 3. Brzozowski, A. M., et al., Structural analysis of a chimeric bacterial α-amylase. High-resolution analysis of native and ligand complexes. Biochemistry, 39(31), 9099-9107 (2000).
- 4. Knapp, S., et al., Crystallization and preliminary crystallographic analysis of an amylopullulanase from the hyperthermophilic archaeon *Pyrococcus woesei*. Proteins, 23(4), 595-597 (1995).
- 5. Andrykovitch, M., et al., Crystallization and preliminary X-ray diffraction studies of NusG, a protein shared by the transcription and translation machines. Acta Crystallogr. D Biol. Crystallogr., 58 (Pt 12), 2157-2158 (2002).
- 6. Campos, A., et al., Crystallization and preliminary X-ray analysis of FlhD from *Escherichia coli*. J. Struct. Biol., 123(3), 269-271 (1998).
- 7. Karabinos, A., et al., The single nuclear lamin of *Caenorhabditis elegans* forms *in vitro* stable intermediate filaments and paracrystals with a reduced axial periodicity. J. Mol. Biol., 325(2), 241-247 (2003).
- 8. Lesignoli, E., et al., Recognition and strand displacement of DNA oligonucleotides by peptide nucleic acids (PNAs). High-performance ionexchange chromatographic analysis. J. Chromatogr. A., 922(1-2), 177-185 (2001).
- 9. Abidi, S. L., and Rennick, K. A., Capillary electrochromatographic evaluation of vitamin Eactive oil constituents: tocopherols and tocotrienols. J. Chromatogr. A, 913(1-2), 379-386 (2001).
- 10. S.P. Fling, D.S. Gregerson, In a high molar concentration for molecular weight determination of peptides and proteins by gel electrophoresis, Anal. Biochem. 155, 83 (1986)
- 11. J. Ambler, M. Rodgers, Electrophoresis of proteins on cellulose acetate membranes, Clin. Chem. 26, 1221 (1980)
- 12. A. Paulus et al., Calibration of polyacrylamide gel columns for the separation of oligonucleotides by capillary electrophoresis, Electrophoresis 11, 702 (1990)
- 13. A. Hofstetter et al., Determination of trace amounts of fluorine from a single sodium carbonate fusion of small geological sample masses, Analyst 116, 65 (1991)