



User Guide

# Amicon® Ultra-4 10K Centrifugal Filter Devices

for volumes up to 4 mL

UFC801008

UFC801024

UFC801096

 For in vitro diagnostic use



English

## Introduction

Amicon® Ultra-4 10K centrifugal filter devices provide fast ultrafiltration, with the capability for high concentration factors and easy concentrate recovery from dilute and complex sample matrices. The vertical design and available membrane surface area provide fast sample processing, high sample recovery (typically greater than 90% of dilute starting solution), and the capability for 80-fold concentration. Typical processing time is 10 to 20 minutes. Solute polarization and subsequent fouling of the membrane are minimized by the vertical design, and a physical deadstop in the filter device prevents spinning to dryness and potential sample loss. The concentrate is collected from the filter device sample reservoir using a pipettor, while the ultrafiltrate is collected in the provided centrifuge tube. The device can be spun in a swinging-bucket (for optimal performance) or fixed-angle rotor. Amicon® Ultra-4 10K devices are supplied non-sterile and are for single use only.

## Intended Use

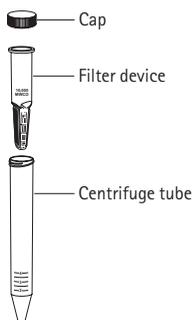
The Amicon® Ultra-4 product line includes 5 different cutoffs (Molecular Weight Cutoff, MWCO), however, the Amicon® Ultra-4 10K device (10,000 MWCO) is the only device intended for in vitro diagnostic use. It can be used to concentrate serum, urine, cerebrospinal fluid, and other body fluids prior to analysis. For information on other Amicon® Ultra-4 cutoffs, go to [www.millipore.com](http://www.millipore.com) and enter *Amicon Ultra-4* in the search box.

## Applications

- Concentration of biological samples containing antigens, antibodies, enzymes, nucleic acids (DNA/RNA samples, either single- or double-stranded), microorganisms, column eluates, and purified samples
- Purification of macromolecular components found in tissue culture extracts and cell lysates, removal of primer, linkers, or molecular labels from a reaction mix, and protein removal prior to HPLC
- Desalting, buffer exchange, or diafiltration

## Materials Supplied

The Amicon® Ultra-4 10K device is supplied with a cap, a filter device, and a centrifuge tube.



## Required Equipment

- Centrifuge with swinging-bucket (preferred) or fixed-angle rotor with wells/carriers that can accommodate 17 mm × 124 mm 15 mL conical-bottomed tubes  
**CAUTION:** To avoid damage to the device during centrifugation, check clearance before spinning.
- Pipettor with 200 microliter ( $\mu\text{L}$ ) tip for concentrate recovery

## Suitability

Preliminary recovery and retention studies are suggested to ensure suitability for intended use. See the "How to Quantify Recoveries" section.

## Shelf Life

Shelf life is 3 years from date of manufacture. Refer to expiration date on package label.

## Rinsing Before Use

The ultrafiltration membranes in Amicon® Ultra-4 10K devices contain trace amounts of glycerine. If this material interferes with analysis, rinse the device with buffer or Milli-Q® water before use. If interference continues, rinse with 0.1 N NaOH followed by a second spin of buffer or Milli-Q® water.

**CAUTION:** Do not allow the membrane in Amicon® Ultra filter devices to dry out once wet. If you are not using the device immediately after rinsing, leave fluid on the membrane until the device is used.

## How to Use Amicon® Ultra-4 Centrifugal Filter Devices

1. Add up to 4 mL of sample (3.5 mL if using a 23° fixed-angle rotor) to the Amicon® Ultra filter device.
2. Place capped filter device into centrifuge rotor (swinging-bucket preferred); counterbalance with a similar device.
3. **When using a swinging-bucket rotor**, spin the device at  $4,000 \times g$  maximum for approximately 10–20 minutes.

**When using a fixed-angle rotor**, orient the device with the membrane panel facing up and spin at  $7,500 \times g$  maximum for approximately 10–20 minutes.

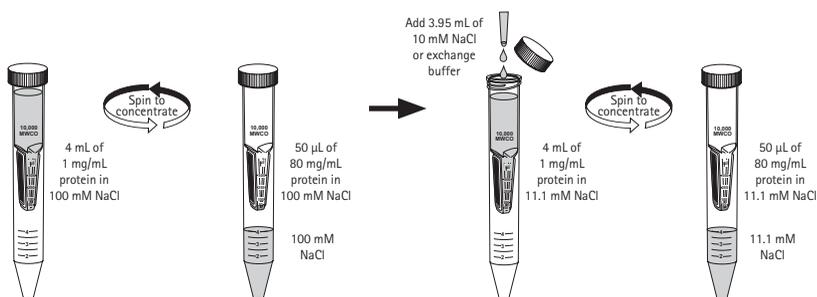
**NOTE:** Refer to Figure 1 and Table 1 for typical spin times.

4. To recover the concentrated solute, insert a pipettor into the bottom of the filter device and withdraw the sample using a side-to-side sweeping motion to ensure total recovery. The ultrafiltrate can be stored in the centrifuge tube.

**NOTE:** For optimal recovery, remove concentrated sample immediately after centrifugation.

## Desalting or Diafiltration

Desalting, buffer exchange, or diafiltration are important methods for removing salts or solvents in solutions containing biomolecules. The removal of salts or the exchange of buffers can be accomplished in the Amicon® Ultra-4 device by concentrating the sample, then reconstituting the concentrate to the original sample volume with any desired solvent. The process of “washing out” can be repeated until the concentration of the contaminating microsolute has been sufficiently reduced. See example below.

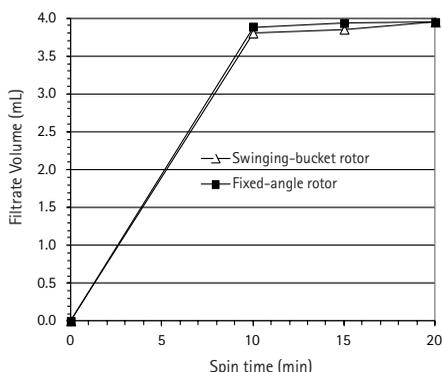


## Performance

### Flow Rate

Factors affecting flow rate include sample concentration, starting volume, chemical nature of solute, relative centrifugal force, centrifuge rotor angle, membrane type, and temperature. Figure 1 and Table 1 can be used to estimate the time required to achieve a given volume of filtrate or concentrate. A typical spin time for a 4 mL sample is approximately 10 to 20 minutes. While most of the sample is filtered in the first 10 to 15 minutes of centrifugation, the lowest concentrate volume (30–75  $\mu\text{L}$ ) is reached after spinning for 15 to 20 minutes.

Figure 1. Typical Filtrate Volume vs. Spin Time



Spin conditions: Swinging-bucket rotor,  $4,000 \times g$ , or fixed-angle rotor,  $7,500 \times g$ , room temperature, 4 mL starting volume. Protein marker used: Cytochrome c,  $n=6$ .

Table 1. Typical Concentrate Volume vs. Spin Time

Spin time (min)	Concentrate volume ( $\mu\text{L}$ )	
	Swinging-bucket rotor $4,000 \times g$	$35^\circ$ Fixed-angle rotor $7,500 \times g$
10	176	97
15	76	54
20	58	35

Spin conditions: Room temperature, 4 mL starting volume.  
Protein marker used: Cytochrome c,  $n=6$  (mean value of 3 membrane lots).  
Shaded volumes were used for the calculation of protein recovery in Table 3.

## Protein Retention and Concentrate Recovery

The membranes used in Amicon® Ultra devices are characterized by a molecular weight cutoff (MWCO); that is, their ability to retain molecules above a specified molecular weight. Solutes with molecular weights close to the MWCO may be only partially retained. Membrane retention depends on the solute's molecular size and shape. For most applications, molecular weight is a convenient parameter to use in assessing retention characteristics. For best results, use a membrane with a MWCO at least two times smaller than the molecular weight of the protein solute that one intends to concentrate. Refer to Table 2.

Table 2. Typical Retention of Protein Markers

Marker/Concentration	Molecular Weight	Device MWCO	% Retention Swinging-bucket	% Retention Fixed-angle	Spin Time (min)
α-Chymotrypsinogen (1 mg/mL)	25,000	10K	>95	>95	15
Cytochrome c (0.25 mg/mL)	12,400		>95	>95	15
Vitamin B-12 (0.2 mg/mL)	1,350		<15	<15	15

Spin Conditions: Swinging-bucket rotor, 4,000 × g, or 35° fixed-angle rotor, 7,500 × g, 4 mL starting volume, room temperature, n=6 (mean value of 3 membrane lots).

Factors that determine sample recovery include the nature of the protein solute relative to the device MWCO chosen, starting concentration, and concentration factor. Table 3 provides typical recoveries for the Amicon® Ultra-4 10K device.

Table 3. Typical Concentrate Recovery

Marker/ Concentration	Device MWCO	Spin Time (min)	Concentrate Volume (µL)		Concentration Factor (x)		Concentrate Recovery (%)	
			Swinging- bucket	Fixed- angle	Swinging- bucket	Fixed- angle	Swinging- bucket	Fixed- angle
Cytochrome c (0.25 mg/mL)	10K	15	76	54	52.3	76.6	97.3	98.5

Spin Conditions: Swinging-bucket rotor, 4,000 × g, or 35° fixed-angle rotor, 7,500 × g, 4 mL starting volume, room temperature, n=6 (mean value of 3 membrane lots). The shaded volumes were taken from Table 1.

## Maximizing Sample Recovery

Low sample recovery in the concentrate may be due to adsorptive losses, over-concentration, or passage of sample through the membrane.

- Adsorptive losses depend upon solute concentration, its hydrophobic nature, temperature and time of contact with filter device surfaces, sample composition, and pH. To minimize losses, remove concentrated samples immediately after centrifugal spin.
- If the starting sample concentration is high, monitor the centrifugation process in order to avoid over-concentration of the sample. Over-concentration can lead to precipitation and potential sample loss.
- If the sample appears to be passing through the membrane, choose a lower MWCO Amicon® Ultra-4 device.

## How to Quantify Recoveries

Calculate total recovery, percent concentrate recovery, and percent filtrate recovery using the method below. The procedure provides a close approximation of recoveries for solutions having concentrations up to roughly 20 mg/mL.

**NOTE:** Appropriate assay techniques include absorption spectrophotometry, radioimmunoassay, refractive index, and conductivity.

### Direct Weighing Procedure

The density of most dilute proteins is nearly equal to the density of water (i.e., 1 g/mL). Using this property, the concentrate and filtrate volumes can be quantified by weighing them and converting the units from grams to milliliters. This technique is valid only for solutions with concentrations of approximately 20 mg/mL or less.

1. Before use, separately weigh the empty filter device, the centrifuge tube, and an empty tube for concentrate collection.
2. Fill filter device with solution and reweigh.
3. Assemble device and centrifuge per instructions.
4. Collect the concentrate with a pipettor and dispense it into the preweighed concentrate collection tube.
5. Remove the device from the centrifuge tube and weigh the centrifuge tube and concentrate collection tube.
6. Subtract weight of empty device/tubes to calculate weights of starting material, filtrate, and concentrate.
7. Assay the starting material, filtrate, and concentrate to determine solute concentration.
8. Calculate recoveries using the weight/volume data and the measured concentrations as follows:

$$\% \text{ concentrate recovery} = 100 \times \frac{W_c \times C_c}{W_o \times C_o}$$

$$\% \text{ filtrate recovery} = 100 \times \frac{W_f \times C_f}{W_o \times C_o}$$

$$\% \text{ total recovery} = \% \text{ concentrate recovery} + \% \text{ filtrate recovery}$$

$W_c$  = total weight of concentrate before assay

$W_o$  = weight of original starting material

$W_f$  = weight of filtrate

$C_c$  = concentrate concentration

$C_o$  = original starting material concentration

$C_f$  = filtrate concentration

## Specifications

### Maximum initial sample volume

Swinging-bucket and  
fixed-angle rotors (45° and 35°) 4.0 mL

Fixed-angle rotor (23°) 3.5 mL

Typical final concentrate volume 50–100  $\mu$ L

### Maximum relative centrifugal force

Swinging-bucket rotor 4,000  $\times$  g

Fixed-angle rotor 7,500  $\times$  g

Active membrane area 3 cm<sup>2</sup>

### Dimensions

Filter device in tube (capped)

Length: 124 mm (4.9 in.) Diameter: 17.3 mm (0.7 in.)

Filter device

Length: 73.4 mm (2.9 in.) Diameter: 17.2 mm (0.7 in.)

### Materials of Construction

Filter device Copolymer styrene/butadiene

Membrane Ultracel® low binding regenerated cellulose

Filtrate tube Polypropylene

Filtrate cap and liner Polyethylene

## Chemical Compatibility

Amicon® Ultra centrifugal devices are intended for use with biological fluids and aqueous solutions. Before use, check the sample for chemical compatibility with the device.

Table 4. Chemical Compatibility of Amicon® Ultra Filter Devices

Acids	Concentration		Concentration
Acetic acid	≤ 50%*	Phosphoric acid	≤ 30%
Formic acid	≤ 5%*	Sulfamic acid	≤ 3%
Hydrochloric acid	≤ 1.0 M	Sulfuric acid	≤ 3%
Lactic acid	≤ 50%	Trichloroacetic acid (TCA)	≤ 10%*
Nitric acid	≤ 10%	Trifluoroacetic acid (TFA)	≤ 30%*

### Alkalis

Ammonium hydroxide	≤ 10%	Sodium hydroxide	≤ 0.5 M
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### Alcohols

n-Butanol	≤ 70%	Isopropanol	≤ 70%
Ethanol	≤ 70%	Methanol	≤ 60%

### Detergents

Alconox® detergent	≤ 1%	Sodium dodecyl sulfate (SDS)	≤ 0.1%
CHAPS detergent	≤ 0.1%	Tergazyme® detergent	≤ 1%
Lubrol® PX detergent	≤ 0.1%	Triton® X-100 surfactant	≤ 0.1%
Nonidet™ P-40 surfactant	≤ 2%	Tween® 20 surfactant	≤ 0.1%
Sodium deoxycholate	≤ 5%		

### Organic solvents

Acetone	not recommended	Ethyl acetate	not recommended
Acetonitrile	≤ 20%	Formaldehyde	≤ 5%
Benzene	not recommended	Pyridine	not recommended
Carbon tetrachloride	not recommended	Tetrahydrofuran	not recommended
Chloroform	not recommended	Toluene	not recommended
Dimethyl sulfoxide (DMSO)	≤ 5%*		

### Miscellaneous

Ammonium sulfate	Saturated	Phenol	≤ 1%
Diethyl pyrocarbonate	≤ 0.2%	Phosphate buffer (pH 8.2)	≤ 1 M
Dithiothreitol (DTT)	≤ 0.1 M	Polyethylene glycol	≤ 10%
Glycerine	≤ 70%	Sodium carbonate	≤ 20%
Guanidine HCl	≤ 6 M	Tris buffer (pH 8.2)	≤ 1 M
Imidazole	≤ 100 mM	Urea	≤ 8 M
Mercaptoethanol	≤ 0.1 M		

\* Contact with this chemical may cause materials to leach out of the component parts. Solvent blanks are recommended to determine whether leachables represent potential assay interferences.

## Product Labeling Symbols

The following table defines the symbols found on Amicon® Ultra-4 10K device labels.

Symbol	Definition	Symbol	Definition
	In vitro diagnostic medical device		Date of manufacture
	Catalogue number		Manufacturer
	Do not reuse		Non-sterile product
	Use-by date		CE conformity marking
	Batch code		

## Product Ordering Information

This section lists the catalogue numbers for Amicon® Ultra Ultrafiltration Devices. See the Technical Assistance section for contact information. You can purchase these products on-line at [www.millipore.com/products](http://www.millipore.com/products).

Initial volume (mL)	Final concentrate volume (µL)	Product	Qty / pk	3K	10K	30K	50K	100K
0.5	15–20	Amicon® Ultra-0.5 device	8	UFC500308	UFC501008	UFC503008	UFC505008	UFC510008
			24	UFC500324	UFC501024	UFC503024	UFC505024	UFC510024
			96	UFC500396	UFC501096	UFC503096	UFC505096	UFC510096
			500	UFC5003BK	UFC5010BK	UFC5030BK	UFC5050BK	UFC5100BK
Amicon® Ultra-0.5 Collection Tubes			96	UFC50VL96				
2	30–70	Amicon® Ultra-2 device	24	UFC200324	UFC201024	UFC203024	UFC205024	UFC210024
4	50–100	Amicon® Ultra-4 device	8	UFC800308	UFC801008*	UFC803008	UFC805008	UFC810008
			24	UFC800324	UFC801024*	UFC803024	UFC805024	UFC810024
			96	UFC800396	UFC801096*	UFC803096	UFC805096	UFC810096
15	150–300	Amicon® Ultra-15 device	8	UFC900308	UFC901008*	UFC903008	UFC905008	UFC910008
			24	UFC900324	UFC901024*	UFC903024	UFC905024	UFC910024
			96	UFC900396	UFC901096*	UFC903096	UFC905096	UFC910096

\* Amicon® Ultra-4 and -15 10K devices are for in vitro diagnostic use. All other devices are for research use only.

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## Standard Warranty

The applicable warranty for the products listed in this publication may be found at [www.millipore.com/terms](http://www.millipore.com/terms) (within the "Terms and Conditions of Sale" applicable to your purchase transaction).



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