SIGMA-ALDRICH®

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Product Information

CDK1/CyclinA1, active, GST-tagged, human PRECISIO[®] Kinase recombinant, expressed in Sf9 cells

Catalog Number **SRP5008** Storage Temperature –70 °C

Synonyms:

CDK1: CDC2 CyclinA1: CCNA1

Product Description

CDK1 or Cell Division Control protein 1 is essential for the completion of START, the controlling event in the cell cycle that is required to initiate mitosis. CDK1 is a catalytic subunit of a protein kinase complex, called the M-Phase Promoting Factor that induces entry into mitosis and is universal among eukaryotes.¹ Phosphorylation of Bcl-2 in G_2 /M phase-arrested cells following photodynamic therapy with hypericin involves a CDK1-mediated signal and delays the onset of apoptosis. Therapeutic potential of CDK inhibitor NU2058 in androgen-independent prostate cancer has also been demonstrated.²

Recombinant full-length human CDK1 and CyclinA1 were co-expressed by baculovirus in *Sf9* insect cells using an N-terminal GST tag on both proteins. The CDK1gene accession number is NM_001786; CyclinA1 is BC036346. Recombinant protein stored in 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM glutathione, 0.1 mM EDTA, 0.25 mM DTT, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: CDK1 ~58 kDa and CyclinA1 ~81 kDa

Purity: 70-95% (SDS-PAGE, see Figure 1)

Specific Activity: 32-48 nmole/min/mg (see Figure 2)

Precautions and Disclaimer

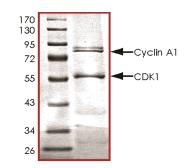
This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

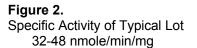
Storage/Stability

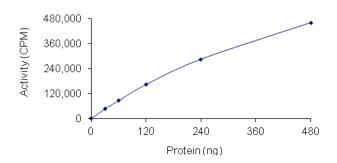
The product ships on dry ice and storage at -70 °C is recommended. After opening, aliquot into smaller quantities and store at -70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Typical Lot 70–95% (densitometry)







Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7. 2, 12.5 mM glycerol 2-phosphate, 25 mM MgC1₂, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 $ng/\mu I$ BSA solution.

Kinase Solution – Dilute the active CDK1/CyclinA1 $(0.1 \ \mu g/\mu)$ with Kinase Dilution Buffer to the desired concentration.

<u>Note</u>: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active CDK1/CyclinA1 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μl aliquots at –20 °C.

 γ -³³P-ATP Assay Cocktail (250 μ M) – Combine 5.75 ml of Kinase Assay Buffer, 150 μ l of 10 mM ATP Stock Solution, 100 μ l of γ -³³P-ATP (1 mCi/100 μ l). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Histone H1 diluted in distilled water to a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

<u>Kinase Assay</u>

This assay involves the use of the ³³P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active CDK1/CyclinA1, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ -³³P-ATP Assay Cocktail may be thawed at room temperature.
- 2. In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl:
 - 10 µl of Kinase Solution
 - 5 µl of Substrate Solution
 - 5μ of cold water (4 °C)
- Set up a blank control as outlined in step 2, substituting 5 μl of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ -³³P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- 5. After the 15 minute incubation, stop the reaction by spotting 20 μ l of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- 6. Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ^{-33} P-ATP counts introduced into the reaction. Spot 5 µl of the γ^{-33} P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR = <u>cpm of 5 μ l of γ -³³P-ATP Assay Cocktail nmole of ATP</u>

cpm – value from control (step 7) nmole – 1.25 nmole (5 μ l of 250 μ M ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\frac{\Delta \text{cpm} \times (25/20)}{\text{SR} \times \text{E} \times \text{T}}$$

SR = specific radioactivity of the ATP (cpm/nmole ATP) \triangle cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

- 20 = spot volume
- T = reaction time (minutes)
- E = amount of enzyme (mg)

References

- Vantieghem, A., et al., Phosphorylation of Bcl-2 in G₂/M phase-arrested cells following photodynamic therapy with hypericin involves a CDK1-mediated signal and delays the onset of apoptosis. J. Biol. Chem., 277(40), 37718-31 (2002).
- Rigas, A.C. et al., Therapeutic potential of CDK inhibitor NU2058 in androgen-independent prostate cancer. Oncogene, 26(55), 7611-9 (2007).

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