For life science research only.
Not for use in diagnostic procedures.

# Taq DNA Polymerase, 1 U/µl
## from *Thermus aquaticus* BM, recombinant (*E. coli*)

**Version: 15**
Content Version: October 2020

Deoxynucleoside-triphosphate: DNA deoxynucleotidyltransferase

| | |
|---|---|
| **Cat. No. 11 647 679 001** | 250 U<br>1 U/µl<br>200 reactions in a final volume of 50 µl |
| **Cat. No. 11 647 687 001** | 1,000 U<br>4 x 250 U<br>800 reactions in a final volume of 50 µl |

**Store the product at −15 to −25°C.**

# 1. General Information

## 1.1. Contents

| Vial / Bottle | Label | Function / Description | Catalog Number | Content |
|---|---|---|---|---|
| 1 | Taq DNA Polymerase | Enzyme storage buffer: 20 mM Tris-HCl, 1 mM DTT, 0.1 mM EDTA, 0.1 M KCl, 0.5% Nonidet P-40 (v/v), 0.5% Tween 20 (v/v), 50% glycerol (v/v), pH 8.0 (+4°C). | 11 647 679 001 | 1 vial, 250 µl |
| | | | 11 647 687 001 | 4 vials, 250 µl each |
| 2 | Taq DNA Polymerase, PCR buffer, 10x conc. with $MgCl_2$ | Buffer composition: 100 mM Tris-HCl, 15 mM $MgCl_2$, 500 mM KCl, pH 8.3 (+20°C). | 11 647 679 001 | 1 vial, 1 ml |
| | | | 11 647 687 001 | 4 vials, 1 ml each |
| 3 | Taq DNA Polymerase, $MgCl_2$ stock solution | 25 mM $MgCl_2$ | 11 647 679 001 | 1 vial, 1 ml |
| | | | 11 647 687 001 | 2 vials, 1 ml each |
| 4 | Taq DNA Polymerase, PCR buffer, 10x conc. without $MgCl_2$ | Buffer composition: 100 mM Tris-HCl, 500 mM KCl, pH 8.3 (+20°C). | 11 647 679 001 | 1 vial, 1 ml |
| | | | 11 647 687 001 | 4 vials, 1 ml each |

## 1.2. Storage and Stability

### Storage Conditions (Product)

When stored at −15 to −25°C, the product is stable through the expiry date printed on the label.

| Vial / Bottle | Label | Storage |
|---|---|---|
| 1 | Taq DNA Polymerase | Store at −15 to −25°C. |
| 2 | PCR buffer, 10x conc. with $MgCl_2$ | |
| 3 | $MgCl_2$ stock solution | |
| 4 | PCR buffer, 10x conc. without $MgCl_2$ | |

## 1.3. Additional Equipment and Reagent required

### Standard laboratory equipment

- Nuclease-free, aerosol-resistant pipette tips
- Pipettes with disposable, positive-displacement tips
- Sterile reaction tubes for preparing PCR mixes and dilutions
- PCR reaction vessels, such as 0.2 ml thin-walled PCR tubes or plates
- Standard benchtop microcentrifuge
- Thermal block cycler

### For PCR

- PCR primers
- Template DNA
- PCR Nucleotide Mix, PCR Grade*
- Water, PCR Grade*

### For DIG DNA labeling

- Digoxigenin-11-dUTP, alkali-stabile*, or
- Digoxigenin-11-dUTP, alkali-labile*

## 1.4. Application

Taq DNA Polymerase is used in a variety of techniques:

- The enzyme activity is stable during prolonged incubation at high temperatures (+95°C) and can therefore be used to amplify DNA fragments by PCR.
- DNA labeling reactions
- Sequencing/cycle sequencing

# 2. How to Use this Product

## 2.1. Before you Begin

### Sample Materials

Use any template DNA such as genomic or plasmid DNA, cDNA suitable for PCR in terms of purity, concentration, and absence of inhibitors. For reproducible isolation of nucleic acids, use:

- Either a MagNA Pure System together with a dedicated reagent kit (for automated isolation),
- or a High Pure Nucleic Acid Isolation Kit (for manual isolation).
- Use 10 to 250 ng complex genomic DNA or 0.1 to 15 ng plasmid DNA/cDNA.

⚠ ***Store the template DNA in either Water, PCR Grade* or 5 to 10 mM Tris-HCl, pH 7 to 8. Avoid dissolving the template in TE buffer since EDTA chelates $Mg^{2+}$.***

### Mg2+ Concentration

1 to 10 mM (as $MgCl_2$) (optimal)
1.5 mM (as $MgCl_2$) when used with 200 µM of each dNTP (standard)

### General Considerations

The optimal conditions, including incubation times and temperatures, concentration of enzyme, template DNA, $Mg^{2+}$ vary from system to system and must be determined for each individual experimental system. At the very least, titrate the $Mg^{2+}$ concentration and the amount of enzyme used per assay to ensure optimal efficiency of DNA synthesis.
As a starting point, use the following guidelines:

- Optimal enzyme concentration: 0.5 to 2.5 U/50 µl. A concentration of 1.25 U/50 µl will usually produce satisfactory results.
- Optimal $Mg^{2+}$ concentration can vary between 1.5 mM and 10 mM. In most cases, a $Mg^{2+}$ concentration of 1.5 mM will produce satisfactory results if you use 200 µM of each dNTP.

For individual optimization of the $Mg^{2+}$ concentration, a buffer without $MgCl_2$ and a $MgCl_2$ stock solution are supplied separately. The table shows the volumes of the $MgCl_2$ stock solution which give the designated $MgCl_2$ concentrations when added to a 50 µl PCR mixture. All other steps for preparing of the reaction mix are the same as described.

| **$MgCl_2$ [mM]** | 1 | 1.25 | 1.5 | 1.75 | 2 | 2.5 | 5 |
|---|---|---|---|---|---|---|---|
| **Volume [µl]** | 2 | 2.5 | 3 | 3.5 | 4 | 5 | 10 |

- dNTP concentration: Always use equal concentrations of all four dNTPs. The final concentration of each dNTP should be between 50 and 500 µM; the most commonly used concentration is 200 µM. If you increase the dNTP concentration, you must also increase the $Mg^{2+}$ concentration.

### Safety Information

#### For customers in the European Economic Area

Contains SVHC: octyl/nonylphenol ethoxylates. For use in research and under controlled conditions only – acc. to Art. 56.3 and 3.23 REACH Regulation.

# 2.2. Protocols

## Preparation of PCR master mixes

Prepare two PCR master mixes. Master Mix 2 contains enzyme and reaction buffer; Master Mix 1 contains all other reaction components. This circumvents the need for hot start and avoids that the enzyme interacts with primers or template during the reaction setup. If you are setting up multiple reactions, the volume of each master mix should be 110% of the volume needed for all the samples. For example, to prepare Master Mix 2 for 20 reactions, make 550 µl of the mix. The extra volume allows for losses during pipetting.

### Preparation of master mix 1

1. Thaw the reagents and store on ice.
   – Briefly vortex and centrifuge all reagents before setting up the reactions.

2. Prepare a 10x-concentrated solution of each respective primer.
   *If you are using, for example, the final concentration of 0.5 µM for each primer, the 10x-concentrated solution would contain a 5 µM concentration of the respective primer.*

3. To a sterile 1.5 ml reaction tube on ice, add the components in the order listed:

| Reagent | Volume [µl] | Final conc. |
|---|---|---|
| Water, PCR Grade* | add up to a final volume of 25 | – |
| PCR Grade Nucleotide Mix* (10 mM of each dNTP) | 1 | 200 µM of each dNTP |
| Forward primer 1 | 5 | 0.1 – 1 µM |
| Reverse primer 2 | 5 | 0.1 – 1 µM |
| Template DNA | variable | 10 – 250 ng gDNA<br>0.1 – 15 ng cDNA |
| **Final Volume** | **25** | |

4. Mix and centrifuge briefly.

### Preparation of master mix 2

1. Thaw the reagents and store on ice.
   – Briefly vortex and centrifuge all reagents before setting up the reactions.

2. To a sterile reaction tube on ice, add the components in the order listed for each 50 µl reaction:

| Reagent | Volume [µl] | Final conc. |
|---|---|---|
| Water, PCR Grade* | add up to a final volume of 25 | – |
| PCR reaction buffer, 10x | 5 | 1x |
| Taq DNA Polymerase (1 U/µl) | 0.5 – 2.5 | 0.5 – 2.5 U/reaction |
| **Final Volume** | **25** | |

3. Mix and centrifuge briefly.

## PCR protocol

*The following thermal profiles are an example. Different thermal cyclers may require different profiles.*

1. For each reaction, combine 25 µl Master Mix 1 and 25 µl Master Mix 2 in a thin-walled PCR tube on ice.
   – Gently vortex the mixture to produce a homogeneous reaction, then centrifuge briefly to collect the solution at the bottom of the tube.

   ⚠ ***Start thermal cycling immediately. Do not store the combined reaction mix on ice.***

2. Place your samples in a thermal block cycler and perform PCR.
   – Run 30 cycles for DNA and 35 cycles for cDNA.

| Step | Temperature [°C] | Time | Number of Cycles |
|---|---|---|---|
| Pre-Incubation | 94 | 2 min | 1 |
| Denaturation<br>Annealing<br>Elongation | 94<br>45 – 68[1]<br>72 | 15 – 30 sec<br>30 – 60 sec<br>45 sec – 3 min[2] | 25 – 30 |
| Final Elongation | 72 | 7 min | 1 |
| Cooling | 4 | indefinitely | |

3. After cycling, use samples immediately or store them frozen for later use.

*For best results, check the PCR product on an agarose gel for size and specificity. Use an appropriate size marker. In addition, purify the PCR product with the High Pure PCR Product Purification Kit, for example, before performing nested PCR.*

(1) Annealing temperature depends on the melting temperature of the primers used.

(2) Elongation time depends on the length of the fragment to be amplified. Use 45 seconds for targets up to 1 kb, 1 minute for fragments up to 1.5 kb, and 2 minutes for fragments up to 3 kb.

## DIG DNA labeling

Digoxigenin 11-dUTP* is incorporated into DNA by Taq DNA Polymerase. See the Instructions for Use of the Digoxigenin-11-dUTP for additional information.

## 2.3. Parameters

### EC-Number

EC 2.7.7.7

### Incorporation of Modified Nucleotides

Enzyme accepts modified nucleotides such as radiolabeled nucleotides, DIG-dUTP, and biotin-dUTP.

### Molecular Weight

Approximately 95 kD.

### pH Optimum

Approximately 9 (+20°C).

### Temperature Optimum

Approximately +75°C (elongation).
Optimal elongation temperature

### Unit Assay

#### Unit assay on activated DNA

##### Incubation buffer

67 mM Tris-HCl; pH 8.3 (+25°C), 5 mM $MgCl_2$, 10 mM 2-mercaptoethanol, 0.2% polydocanol, 0.2 mg/ml gelatin, 0.2 mM each dATP, dGTP, dTTP, and 0.1 mM dCTP.

##### Incubation procedure

1. M13mp9ss, M13 primer (17mer), and 1 µCi ($\alpha^{32P}$) dCTP are incubated with suitable dilutions of Taq DNA Polymerase in 50 µl Incubation buffer at +65°C for 60 minutes.

2. The amount of incorporated dNTPs is determined by trichloroacetic acid precipitation.

### Unit Definition

One unit of Taq DNA Polymerase is defined as the amount of enzyme that incorporates 10 nmol of total deoxyribonucleoside triphosphates into acid precipitable DNA within 60 minutes at +65°C under the described assay conditions.

### Volume Activity

1 U/µl
As determined in the assay on activated DNA.

### Working Concentration

0.5 to 2.5 U per 50 µl reaction (optimal).
1.25 U per 50 µl reaction (standard).

# 3. Troubleshooting

| Observation | Possible cause | Recommendation |
|---|---|---|
| Little or no PCR product. | Difficult templates, such as GC-rich templates. | Perform PCR with the GC-RICH PCR System*. |
| | | Add DMSO (final concentration, 8%), and reduce enzyme concentration, for example, down to 0.5 U per reaction. |
| | DNA template problems. | Check quality and concentration of template:<br>▪ Analyze an aliquot on an agarose gel to check for possible degradation.<br>▪ Test the template with an established primer pair or PCR system.<br>▪ Check or repeat template purification. |
| | Enzyme concentration too low. | Increase enzyme concentration to 2 U Taq DNA Polymerase per 50 µl reaction. |
| | | If necessary, increase the amount of polymerase in 0.5 U steps. |
| | $MgCl_2$ concentration too low. | Increase the $MgCl_2$ concentration in 0.25 mM steps; the minimal acceptable concentration is 1.5 mM $MgCl_2$. |
| | Cycle conditions not optimal. | Decrease annealing temperature. |
| | | Increase cycle number. |
| | | Make sure that the final elongation step is included in the program. |
| | Primer design not optimal. | Design alternative primers. |
| | Primer concentration not optimal. | Both primers must have the same concentration. |
| | | Titrate primer concentration (0.1 to 1 µM). |
| | Primer quality or storage problems. | If you use an established primer pair, check performance in an established PCR system, for example, with a control template. |
| | | Make sure that the primers are not degraded. |
| | | Always store primers at −15 to −25°C. |
| | Formation of primer-dimers. | Use two master mixes, as described in the protocol. |
| | | Use FastStart Taq DNA Polymerase* instead of Taq DNA Polymerase. |
| Multiple bands or background smear. | Annealing temperature too low. | Increase annealing temperature.<br>*Longer primers have higher annealing temperatures.* |
| | Primer design or concentration not optimal. | Review primer design. |
| | | Titrate primer concentration (0.1 to 0.6 µM). |
| | | Both primers must have the same concentration. |
| | | Perform nested PCR with nested primers. |
| | Difficult templates, such as GC-rich templates. | Perform PCR with the GC-RICH PCR System*. |
| | DNA template problems. | Use serial dilution of template. |
| PCR products in negative control experiments. | Carryover contamination present. | Replace all reagents, especially water. |
| | | Use aerosol-resistant pipette tips. |
| | | Set up PCR reactions in an area separate from that used for PCR product analysis. |
| | | To eliminate carryover contaminants:<br>▪ Use dUTP (600 µM) instead of dTTP (200 µM), and Uracil-DNA Glycosylase* (1 U/50 µl reaction).<br>▪ Increase $Mg^{2+}$ concentration to a maximum of 4 mM to compensate for higher dNTP concentration. |
| Problems specific to RT-PCR. | No product, additional bands, background smear observed. | The volume of cDNA template (RT reaction) should not exceed 10% of the final volume of the PCR reaction. |
| | | Follow all troubleshooting tips. |
| | | Increase $MgCl_2$ in 0.25 mM steps. |

# 4. Additional Information on this Product

## 4.1. Test Principle

Taq DNA Polymerase is a highly processive 5′→3′ DNA polymerase that lacks 3′→5′ exonuclease activity. It is a single polypeptide chain with a molecular weight of approximately 95 kD.
Taq DNA Polymerase was originally isolated from the thermophilic eubacterium *Thermus aquaticus* BM, a strain lacking Taq I restriction endonuclease. The enzyme was cloned in *E. coli*.

## 4.2. Quality Control

For lot-specific certificates of analysis, see section, **Contact and Support**.

# 5. Supplementary Information

## 5.1. Conventions

To make information consistent and easier to read, the following text conventions and symbols are used in this document to highlight important information:

| Text convention and symbols | |
|---|---|
| *Information Note: Additional information about the current topic or procedure.* | |
| **Important Note: Information critical to the success of the current procedure or use of the product.** | |
| ① ② ③ etc. | Stages in a process that usually occur in the order listed. |
| ❶ ❷ ❸ etc. | Steps in a procedure that must be performed in the order listed. |
| * (Asterisk) | The Asterisk denotes a product available from Roche Diagnostics. |

## 5.2. Changes to previous version

Layout changes.
Editorial changes.
New information added related to the REACH Annex XIV.

## 5.3. Ordering Information

| Product | Pack Size | Cat. No. |
|---|---|---|
| Reagents, kits | | |
| Digoxigenin-11-dUTP, alkali-labile | 25 nmol, 25 µl, 1 mM | 11 573 152 910 |
| | 125 nmol, 125 µl, 1 mM | 11 573 179 910 |
| PCR Nucleotide Mix | 200 µl, 500 reactions of 20 µl final reaction volume | 11 581 295 001 |
| | 5 x 200 µl, 2,500 reactions of 20 µl final reaction volume. | 04 638 956 001 |
| | 10 x 200 µl, 5,000 reactions of 20 µl final reaction volume. | 11 814 362 001 |
| GC-RICH PCR System | 100 U, 50 reactions in a final volume of 50 µl | 12 140 306 001 |
| Water, PCR Grade | 25 ml, 25 x 1 ml | 03 315 932 001 |
| | 25 ml, 1 x 25 ml | 03 315 959 001 |
| | 100 ml, 4 x 25 ml | 03 315 843 001 |
| Digoxigenin-11-dUTP, alkali-stable | 25 nmol, 25 µl, 1 mM | 11 093 088 910 |
| | 125 nmol, 125 µl, 1 mM | 11 558 706 910 |
| | 5 x 125 nmol, 5x 125 µl, 1 mM | 11 570 013 910 |
| Taq DNA Polymerase, 5 U/µl | 100 U, 1 x 100 U, 50 reactions in a final volume of 50 µl | 12 032 902 001 |
| | 500 U, 2 x 250 U, 250 reactions in a final volume of 50 µl | 12 032 929 001 |
| | 1,000 U, 4 x 250 U, 500 reactions in a final volume of 50 µl | 12 032 937 001 |
| | 2,500 U, 10 x 250 U, 1,250 reactions in a final volume of 50 µl | 12 032 945 001 |
| | 5,000 U, 20 x 250 U, 2,500 reactions in a final volume of 50 µl | 12 032 953 001 |
| Uracil-DNA Glycosylase, heat-labile | 100 U, 1 U/µl | 11 775 367 001 |
| | 500 U, 1 U/µl | 11 775 375 001 |

## 5.4. Trademarks

FASTSTART and MAGNA PURE are trademarks of Roche.
All other product names and trademarks are the property of their respective owners.

## 5.5. License Disclaimer

For patent license limitations for individual products please refer to:
**List of biochemical reagent products**.

## 5.6. Regulatory Disclaimer

For life science research only. Not for use in diagnostic procedures.

## 5.7. Safety Data Sheet

Please follow the instructions in the Safety Data Sheet (SDS).

## 5.8. Contact and Support

To ask questions, solve problems, suggest enhancements or report new applications, please visit our **Online Technical Support Site**.

To call, write, fax, or email us, visit **sigma-aldrich.com**, and select your home country. Country-specific contact information will be displayed.

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