



## Product Information

### Chemichrome™ Ultimate

For Western blotting systems using anti-mouse, anti-rabbit, or anti-goat secondary antibodies

Product Code **C 8738**

Storage Temperature  $-20\text{ }^{\circ}\text{C}$

### Product Description

Chemichrome™ Ultimate is a positive control for use in Western blotting. It is designed for qualitative molecular weight determinations in Laemmli SDS-PAGE systems<sup>1</sup> and for use as a visual confirmation of Western blot transfer efficiency.

Chemichrome Ultimate contains eight proteins that have been chemically reduced, alkylated, and conjugated to brilliantly colored dyes. These eight proteins can be readily visualized in a gel or on a membrane after transfer. Chemically reduced and alkylated mouse, rabbit, and goat IgG have also been added, so that Chemichrome Ultimate can be used as a positive control for anti-mouse, anti-rabbit, or anti-goat secondary antibody conjugates in either colorimetric or chemiluminescent systems. The heavy chain of the IgG (approximately 50 kDa) is usually visualized after Western blotting. The light chain of IgG (25 kDa) may be visualized if a secondary antibody specific to the Fab region of the primary antibody is used. A 100 kDa band (heavy chain dimer) may also be present if using a rabbit system.

Chemichrome Ultimate is supplied as a ready-to-use solution that resists freezing.

- No need for the chemical reduction of the markers before loading the gel.
- No boiling is required.
- No freeze/thaw cycles means diminished degradation and longer shelf life.
- Storage at  $-20\text{ }^{\circ}\text{C}$  conserves valuable  $-70\text{ }^{\circ}\text{C}$  freezer space.

The product is supplied as a ready-to-use solution. Simply allow Chemichrome Ultimate to warm to room temperature before use and then load onto a gel. Chemichrome Ultimate transfers cleanly to PVDF or nitrocellulose membranes using Towbin's<sup>2</sup> or CAPS buffer, respectively.

Each vial of Chemichrome Ultimate contains 200  $\mu\text{l}$  of solution.

### Reagents and Equipment Required But Not Provided

- SDS-PAGE gels, running buffer, and gel unit or apparatus
- Nitrocellulose (Product Code N 5891) or PVDF (Product Code P 4188) membrane
- Blotting paper (Product Code P 7796)
- Western transfer buffer (Product Code T 4904)
- Western blotting apparatus
- Blocking Solution (see Related Products)
- Methanol (Product Code M 1775)
- Primary antibody
- Anti-mouse, rabbit, or goat secondary antibody
- For colorimetric detection, TMB substrate (Product Code T 0565) for horseradish peroxidase (HRP) detection, BCIP/NBT substrate (Product Code B 6404) for alkaline phosphatase (AP) detection, or a suitable chemiluminescent substrate for HRP or AP detection should be used.
- Tris buffered saline with TWEEN<sup>®</sup> 20 (TBST, Product Code T 9039) or phosphate buffered saline with TWEEN 20 (PBST, Product Code P 3563).

### Precautions and Disclaimer

This product is for laboratory research use only. Consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Note: It is **not** recommended that Chemichrome Ultimate be used as standards for quantitative molecular weight determinations, but only as a qualitative tool. For quantitative molecular weight determinations use the following:  
SigmaMarker, Low Range, 6.5-66 kDa,  
(Product Code M 3913)  
SigmaMarker, High Range, 36-205 kDa,  
(Product Code M 3788)  
SigmaMarker, Wide Range, 6.5-205 kDa,  
(Product Code M 4038)

## Storage/Stability

It is recommended to store the product at  $-20\text{ }^{\circ}\text{C}$ . Chemichrome Ultimate is stable for at least one year as supplied.

## Procedure

### Load Amount

Different substrates and different secondary antibodies in conjunction with different blocking agents will cause variation in the detection of the target protein(s). As a result, different amounts of Chemichrome Ultimate should be loaded onto the gel depending on the blocking agent, secondary antibody, and substrate used. High sensitivity substrates and blockers need less Chemichrome Ultimate than do the less sensitive substrates and blockers. See Table 1 for the recommended loading volumes for different blockers and substrates. If a Western blot is performed and the Chemichrome Ultimate signal is too strong, either load less Chemichrome Ultimate or dilute the Chemichrome Ultimate in Laemmli sample buffer (Product Code S 3401) before loading onto the gel.

**Table 1.**

Recommended loading volumes of Chemichrome Ultimate per well

Blocking Reagent Used	Detection System		
	ECL™ and ECL Plus™	SuperSignal® West Femto and SuperSignal® West Dura	TMB and BCIP/NBT
	μl of Chemichrome Ultimate per well*		
Western Blocker Solution	5 μl	2.5 μl	5 μl
Non-fat Dry Milk Blocker	10 μl	5 μl	5 μl

\*This recommendation is appropriate for a gel of 1 mm thickness and a well of 5 mm width.

## Western Blotting Procedure

This procedure assumes that the optimal amounts of primary and secondary antibodies are known. This is a general guideline procedure; each antibody-antigen pair must be optimized for signal by the end user.

1. All steps below should be performed with slight agitation on a rocker or orbital shaker such that the membrane is freely floating. All incubations should be performed at room temperature.
2. Load the gel with Chemichrome Ultimate and samples (see Table 1).
3. Run the gel and then transfer to either a PVDF or nitrocellulose membrane.
4. Wash the membrane for at least 2 minutes with high purity water.
5. Block the membrane in appropriate blocking agent for 30–60 minutes (see Related Products for suggestions).
6. Add the primary antibody to the blocking agent. The final concentration of the primary antibody in this solution can range from 1-20 μg/ml.
7. Incubate the membrane with the primary antibody solution for at least 30 minutes.
8. Wash with TBST (Product Code T 9039) or PBST (Product Code P 3563) for at least 1 minute.
9. Remove the TBST or PBST and add at least 10 ml of appropriate blocking agent to the membrane. Add the appropriate amount of secondary antibody recommended by the detection substrate used.
10. Incubate the membrane with the secondary antibody solution for 30–60 minutes.
11. Remove the blocking solution and wash the membrane 4 times for 5 minutes each with TBST or PBST.
12. Remove the membrane from the wash buffer and drain any excess liquid from the membrane. Keep the membrane damp; do not allow the membrane to dry out.
13. Place the membrane on a flat sheet of plastic wrap or on any clean plastic surface.
14. Use the detection substrate (colorimetric or chemiluminescent) compatible with the secondary antibody (HRP or AP) used (see Related Products for suggestions).

15.

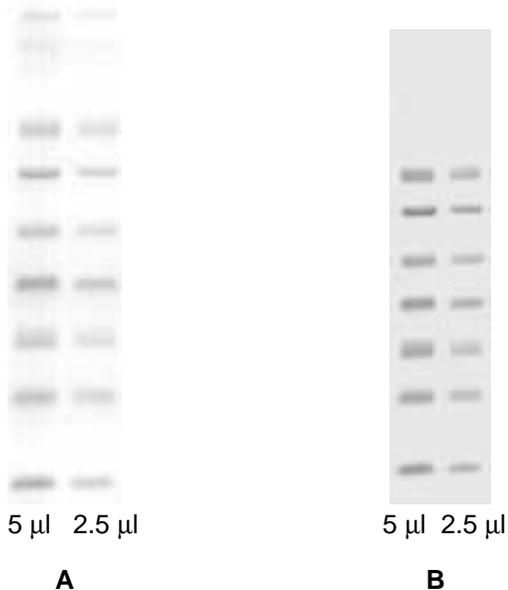
**Table 2.**

Apparent Molecular Weights (kDa) of the Proteins in Chemichrome Ultimate		
Band Color	4 → 20% Tris-Glycine	10 → 20% Tris-Tricine
Violet	220	210
Pink	100	90
Blue	60	65
Pink	45	40
Orange	30	30
Blue	20	20
Pink	12	13
Blue	8	8

Apparent Molecular Weights were determined by using SigmaMarker, Wide Range, 6.5-205 kDa, (Product Code M 4038) as a standard. The molecular weight of the violet band, which is outside the range of the standard, is an approximation.

**Figure 1.**

Chemichrome Ultimate in SDS-PAGE Gel and PVDF membrane



- A. 10-20% Tris-tricine gel was loaded with 5 and 2.5 µl of Chemichrome Ultimate. The gel was run using standard conditions on an 8 x 8 cm, 1 mm thick, 12-well precast gel.
- B. Chemichrome Ultimate transferred to PVDF membrane from the above Tris-tricine gel. Transfer was completed in 60 minutes at 70 volts with Towbin's buffer (Tris-Glycine in 20% methanol).<sup>2</sup>

**Related Products**

Product Name	Package Size	Product Code
TBS	10 packets	T 6664
PBS	10 packets	P 3813
Western Blocker Solution	400 ml	W 0138
TBS + 3% milk	10 packets	T 8793
PBS + 3% milk	10 packets	P 2194
TBS + TWEEN 20	10 packets	T 9039
PBS + TWEEN 20	10 packets	P 3563
Anti-mouse HRP Ab	2 ml	A 9044
Anti-rabbit HRP Ab	1 ml	A 0545
Anti-goat HRP Ab	1 ml	A 5420
Anti-mouse AP Ab	0.25, 0.5, 1 ml	A 3562
TMB substrate	100 ml	T 0565
BCIP / NBT substrate	100 ml	B 6404

**References**

1. Laemmli, U.K., Nature, **227**, 680-685 (1970).
2. Towbin, H., et al., Proc. Natl. Acad. Sci. USA, **76**, 4350-4354 (1979).

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