

Technical Bulletin

Jurkat HLA Panel Cell Lines

Catalog Number HLA002**Product Description**

The Jurkat HLA panel is comprised of eleven (11) genetically modified cell lines targeting MHC Class I HLA molecules in a Jurkat T lymphocyte parental cell line background. The full HLA panel consists of a Beta-2 microglobulin (β2M) knockout cell line and an additional ten (10) monoallelic HLA expression cell lines.

The panel consists of the following cell lines (Figure 1):

1. HLA002A-1VL: Jurkat B2M KO Cells
2. HLA002B-1VL:
Jurkat HLA-A*02:01 Cells
3. HLA002C-1VL:
Jurkat HLA-A*01:01 Cells
4. HLA002D-1VL:
Jurkat HLA-A*03:01 Cells
5. HLA002E-1VL:
Jurkat HLA-A*11:01 Cells
6. HLA002F-1VL:
Jurkat HLA-A*24:02 Cells
7. HLA002G-1VL:
Jurkat HLA-B*15:10 Cells
8. HLA002H-1VL:
Jurkat HLA-B*07:02 Cells
9. HLA002I-1VL:
Jurkat HLA-B*08:01 Cells
10. HLA002J-1VL:
Jurkat HLA-B*35:01 Cells
11. HLA002K-1VL:
Jurkat HLA-B*40:01 Cells

CompoZr® zinc finger nuclease (ZFN) technology was used to create a targeted knockout (KO) of the Beta-2 microglobulin (β2M) gene in wild type Jurkat cells (Catalog Number 88042803). Generation of the Jurkat β2M KO cell line was confirmed to not express endogenous cell surface MHC Class I HLA molecules (HLA-A or HLA-B) via next-generation sequencing (NGS) analysis as shown in Figure 2 and Figure 3 and fluorescence-activated cell sorting (FACS) as shown in Figure 4.

Following single cell cloning and expansion of the Jurkat β2M KO cell line, MISSION® lentiviral particles were used to generate the ten (10) monoallelic HLA cell lines which express individual HLA-A or HLA-B subtypes on the cell surface via a β2M:HLA fusion protein.

Individual HLA-A or HLA-B subtype expression was measured in the HLA panel via fluorescent-activated cell sorting (FACS) as shown in Figure 4.

Figure 1
Generation of the HLA Panel Cell Lines

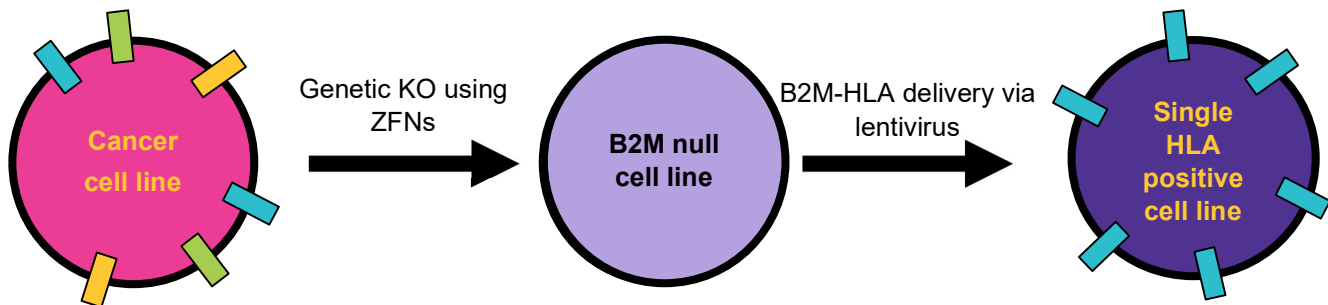


Figure 2. Genomic mutations in the $\beta 2M$ locus. -5 and -1 bp deletions within exon 2 of $\beta 2M$ in Jurkat cells. Reference sequence in wild type Jurkat cells, ZFN target site is highlighted in yellow.

REFERENCE	CTTTCTATCTCTTGTACTACACTGAATTCACCCCCACTGAAAAAGATGAGTATGCCTGCCGTGTGAACCATGTGACTTTG	
CALL #1	CTTTCTATCTCTTGTACTACACTGAATTCACCCCCACT - - - - AGATGAGTATGCCTGCCGTGTGAACCATGTGACTTTG	50% 17491 reads
CALL #2	CTTTCTATCTCTTGTACTACACTGAATTCACCCCCACTGAAAA - GATGAGTATGCCTGCCGTGTGAACCATGTGACTTTG	47% 16552 reads
BELOW CALLING THRESHOLD		3% (978 reads)

Figure 3. Exonic alignment -5 and -1 bp deletions. Both deletions result in predicted premature stop codons in exon 2 of $\beta 2M$ (indicated by red asterisks). ZFN target sites are shown in cyan.



Genomic sequence at the target region recognized by the ZFN pair.
TGTACTACACTGAATTCACCCCCACTGAAAAAGATGAGTATGCCTGCCGTGTGAACC

NGS PCR for genotyping

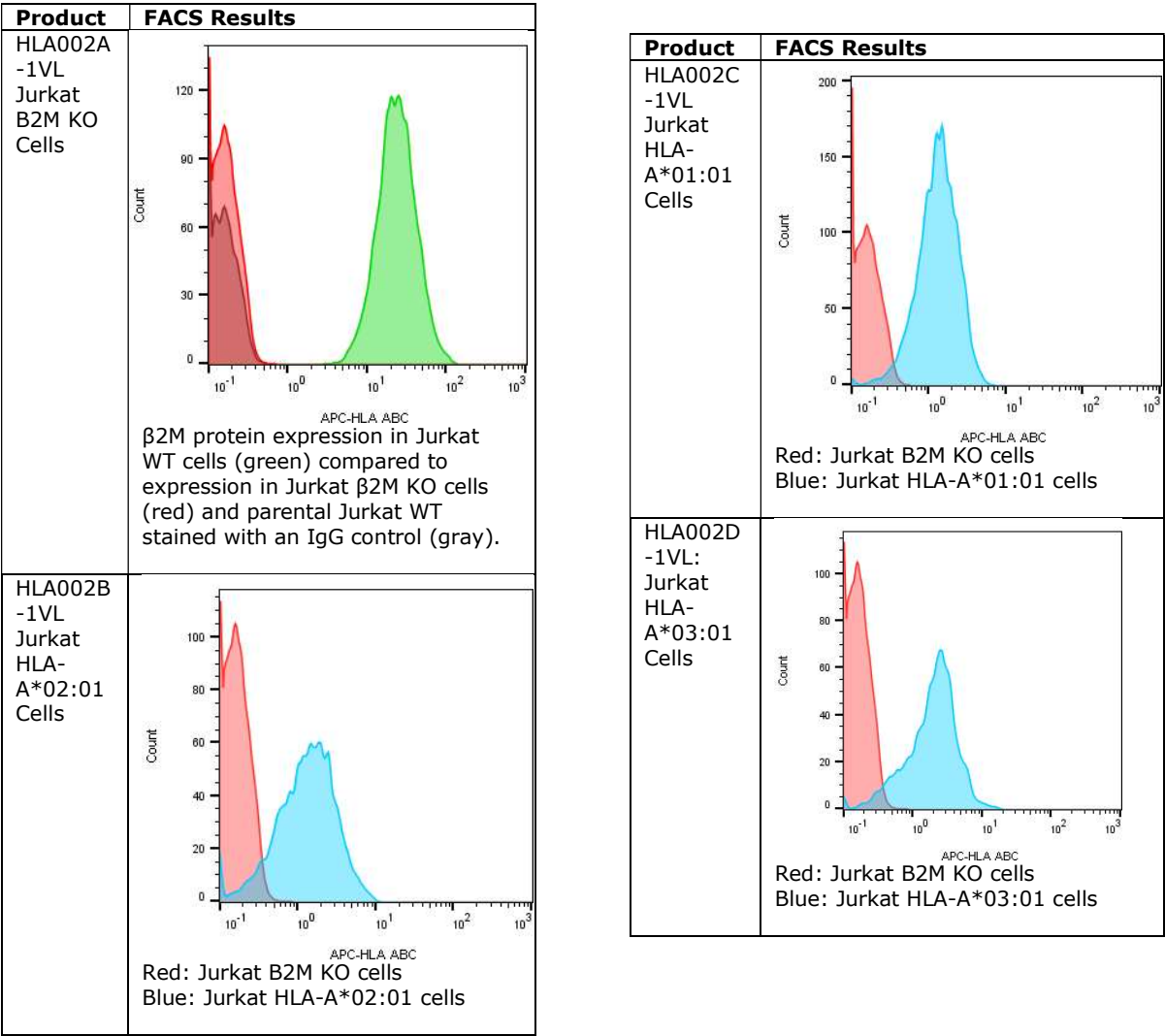
Forward: TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGNNNNNNctgggtttcatccatccgaca
Reverse: GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGNNNNNNtgggatgggactcattcagg

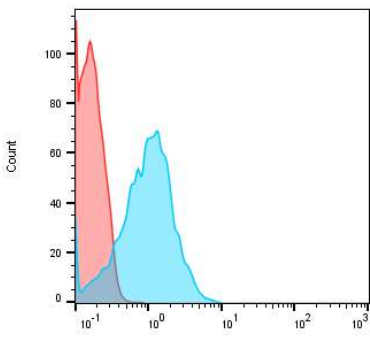
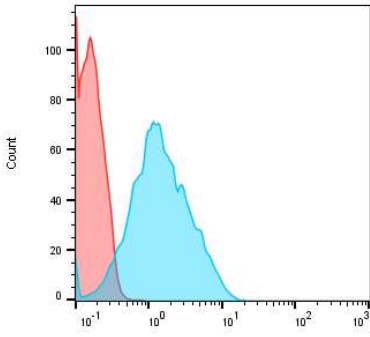
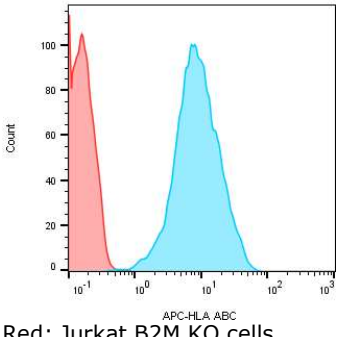


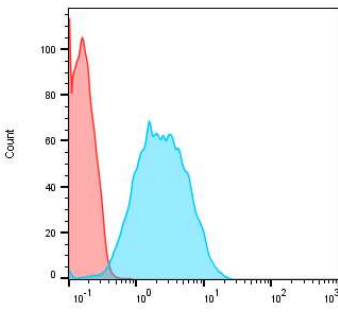
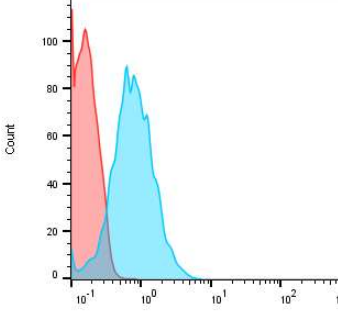
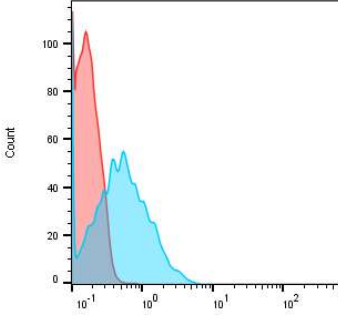
Wildtype amplicon sequence (321 bp)

CTGGGTTTCATCCATCCGACATTGAAGTTGACTTACTGAAGAATGGAGAGAGAATTGAAAAAGTGGAGCATT
CAGACTTGTCTTTCAGCAAGGACTGGTCTTTCTATCTCTTGACTACACTGAATTCACCCCCACTGAAAAAGA
TGAGTATGCCTGCCGTGTGAACCATGTGACTTTGTCACAGCCCAAGATAGTTAAGTGGGGTAAGTCTTACAT
TCTTTTGAAGCTGCTGAAAGTTGTGTATGAGTAGTCATATCATAAAGCTGCTTTGATATAAAAAAGGTCTAT
GGCCATACTACCCTGAATGAGTCCCCATCCCA

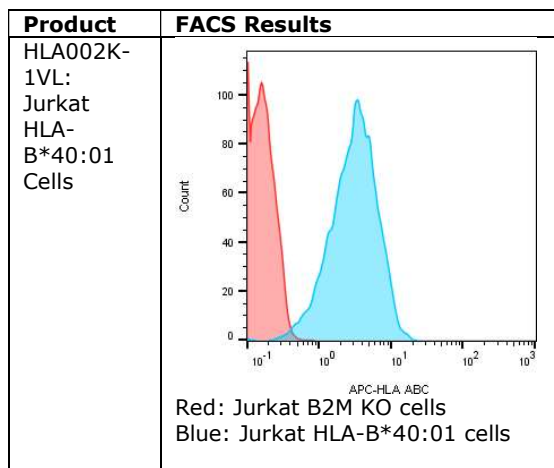
Figure 4. Confirmation of β 2M KO and expression of individual HLA-A or HLA-B subtypes in the Jurkat HLA Panel via FACS Analysis. Cells were stained with APC-IgG or APC-HLA ABC antibody (clone W6/32).



Product	FACS Results
HLA002E-1VL: Jurkat HLA-A*11:01 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-A*11:01 cells</p>
HLA002F-1VL: Jurkat HLA-A*24:02 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-A*24:02 cells</p>
HLA002G-1VL: Jurkat HLA-B*15:10 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-B*15:10 cells</p>

Product	FACS Results
HLA002H-1VL: Jurkat HLA-B*07:02 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-B*07:02 cells</p>
HLA002I-1VL: Jurkat HLA-B*08:01 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-B*08:01 cells</p>
HLA002J-1VL: Jurkat HLA-B*35:01 Cells	 <p>APC-HLA ABC</p> <p>Red: Jurkat B2M KO cells Blue: Jurkat HLA-B*35:01 cells</p>

MERCK



Components

This product is eleven (11) cryovials containing a minimum of 1 million Jurkat cells in each vial.

The cryoprotectant medium used is CryoStor® cell cryopreservation medium containing 10% DMSO (Catalog Number C2874).

Cell Line Description

Organism: Homo sapiens (human)
Tissue: Blood (leukemic T-cell lymphoblast)
Gender: Male
Morphology: Lymphoblastoid
Growth Properties: Suspension

DNA Profile

STR-PCR Data:
Amelogenin: X,Y
CSF1PO: 12
D13S317: 8, 11
D16S539: 11
D18S51: 13, 22
D21S11: 31.2, 33.2
D3S1358: 16
D5S818: 9
D7S820: 8, 12
D8S1179: 13, 14
FGA: 20
Penta_D: 11, 13
Penta_E: 9, 12

TH01: 6, 9.3
TPOX: 8, 10
vWA: 18, 19

The STR profile of this cell line matches that of its parental cell line European Collection of Authenticated Cell Cultures (ECACC) Catalog Number 88042803. Please see the ECACC Catalog Number 88042803 datasheet for additional information about the origin of this cell line.

Reagents and Equipment Required but Not Provided

- RPMI-1640 Medium with L-glutamine and sodium bicarbonate, Catalog Number R8758
- Fetal Bovine Serum, USA origin, sterile-filtered, Catalog Number F2442
- Biological safety cabinet
- 70% ethanol (prepared from Ethanol, Catalog Number E7148)
- Bio-Pure™ alcohol wipes, Catalog Number Z688487
- 37 °C water bath (operating range 35-38 °C)
- Sterile 15 mL conical tubes
- Centrifuge
- Serological pipettor with 1, 2, 5, 10, and 25 mL sterile pipettes
- Vacuum aspiration system and sterile plastic or glass aspiration tips
- Sterile 25 cm² or 75 cm² culture flasks
- 37 °C, 5% CO₂ incubator

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Precaution: It is recommended that protective gloves and clothing always be used, and a full-face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to the gas phase may result in the rapid expansion of the vessel, potentially blowing off its cap with dangerous force creating flying debris.

Storage/Stability

Store cells at -196 °C (liquid nitrogen)

Upon receiving a shipment of frozen cells, it is important the end user gives the shipment attention without delay. To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70 °C. Storage at -70 °C will result in loss of viability.

At the time a cell line is ordered, end users should also consider the culture conditions for the new cell line and make sure the appropriate medium will be available when the cells arrive.

Procedure

Medium Preparation Instructions

The base medium for this cell line is RPMI 1640 with L-Glutamine (Catalog Number R8758).

Complete Medium: To make the complete growth medium, add Fetal Bovine Serum (Catalog Number F2442) to a final concentration of 10%.

Thawing of Frozen Cells

1. Thaw the vial by gentle agitation in a 37 °C water bath for ~1 minute. To reduce the possibility of contamination, keep the O-ring and cap out of the water.
2. Remove the vial from the water bath as soon as the contents are thawed and decontaminate by dipping in or spraying with 70% ethanol solution. All the operations from this point on should be carried out under aseptic conditions.
3. Transfer the cell suspension to a 15 ml conical tube containing 9 mL of warmed Complete Medium.
4. Centrifuge the cells at 125 × g for 5-7 minutes at room temperature.
5. Aspirate the media from the tube. Resuspend the cell pellet with 6 mL of warmed Complete Medium and plate into a 25 cm² or 75 cm² culture flask.
6. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested, prior to the addition of the vial contents, the culture vessel containing the Complete Medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0-7.6) and temperature (37 °C).
7. Incubate the culture at 37 °C in an incubator containing an atmosphere of 5% CO₂ in air.



Sub-culturing Procedure

Cells should be sub-cultured when at $0.9 - 1.1 \times 10^6$ cells/mL

1. Perform cell count and dilute with fresh media to a cell concentration of 400,000 cells/mL.
2. Add appropriate aliquots of the cell suspension into new culture vessels.
3. Incubate cultures at 37 °C in an incubator containing an atmosphere of 5% CO₂ in air.

References

1. Giard, D.J., et al., *In vitro* cultivation of human tumors: establishment of cell lines derived from a series of solid tumors. *J. Natl. Cancer Inst.*, **51(5)**, 1417-23 (1973). PMID: 4357758
2. Gornalusse, G., Hirata, R., Funk, S. et al., HLA-E-expressing pluripotent stem cells escape allogeneic responses and lysis by NK cells. *Nat. Biotechnol.* **35**, 765-772 (2017). PMID: 28504668

Additional product and technical information can be obtained by searching for the catalog number at sigmaaldrich.com.

These products are covered by the Purchase Agreement as described in Exhibit 1.

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HLA Panel Cell Lines

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