



# **GenElute<sup>™</sup>-E Single Spin Blood DNA 96 Kit**

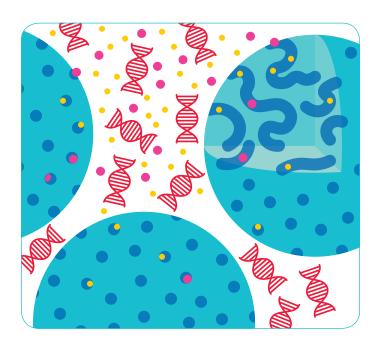


# What's Inside: Reduced waste for a better environment A simplified workflow Specifications . . . . . . . . . . . . . . . . 4 Intended Use . . . . . . . . . . . . . . . . . 4 Typical Results Storage and Stability . . . . . . . . . . 5 Kit Storage Sample Storage and Variability Disposal Materials and Equipment Needed ..... 5 Kit Contents Not Supplied in Kit Preparation before starting Standard Protocol..........6 Lysis Preparation of Purification Plate Purification of DNA Product Ordering ..... 9 Precautions and Disclaimer Notice Standard Warranty Contact Information Technical Assistance GenElute™-E Single Spin Checklist ...... 10 EC196

For research use only.

#### Introduction

GenElute™-E Single Spin Kit is a nucleic acid purification system that eliminates the need for high salt binding and ethanol wash steps, yielding DNA and RNA preparations with fewer impurities for more robust results. GenElute™-E DNA and RNA purification kits employ a **negative chromatography** method dependent on size exclusion to separate large DNA and RNA nucleic acid molecules from smaller protein, lipid, and ionic components in cell, tissue, blood, and other samples.



Using negative chromatography, Single Spin columns efficiently absorb and retain sample contaminants while allowing nucleic acids to flow through the column, reducing the number of steps and plastic materials required for purification. The key is the novel lysis that allows negative chromatography to be used for high quality nucleic acid purification.

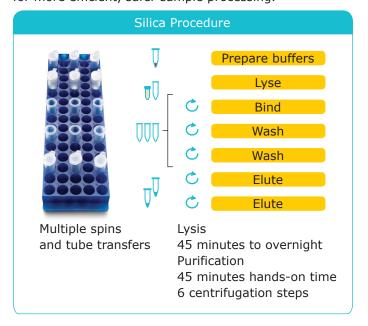
Three key advantages over silica:

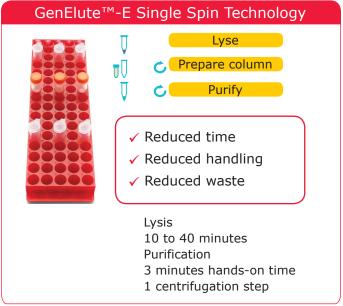
- · Simplified workflow
- Superior performance
- Waste reduction

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#### A simplified workflow

Purification in one spin, eliminating all wash steps and reducing tube handling for more efficient, safer sample processing.





#### Reduced waste for a better environment

With fewer plastic tubes and no hazardous liquids, GenElute™-E DNA and RNA purification kits provide an ecofriendly alternative to silica-based purification.

GenElute™-E purification kits greatly reduce the amounts of plastic-based components packaged with each kit and consumed while executing protocols in the lab. All tedious binding and washing steps associated with silica-based procedures are omitted, with no use of hazardous materials such as chaotropic salts or organic solvents that require special disposal. Plastic waste is reduced by 55% compared to a common silica kits, resulting in disposal cost savings and reduced environmental impact.

GenElute<sup>™</sup>-E Single Spin nucleic acid purification kits provide easier workflows for DNA and RNA isolation, better nucleic acid quality with fewer impurities, and reduced plastic and hazardous waste disposal compared to silica bind-wash-elute spin prep kits.

#### **GenElute™-E Single Spin Purification supports:**

- Significantly reduced plastic waste
- No hazardous bind and wash steps
- Responsible and sustainable nucleic acid purification
- Disposal cost savings



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## **Specifications**

Sample Input	Up to 60 µl	
Sample Type	Human and animal whole blood	
Sample Condition	Fresh, frozen, stabilized	
Required time after lysis	2 minutes	
Purified Nucleic Acid	DNA > 200 bp	
Final Volume	90-110 μl	
The purified genomic	<ul> <li>Restriction digestions</li> </ul>	
DNA is ready for immediate use in	<ul> <li>PCR and qPCR</li> </ul>	
these downstream	<ul> <li>Southern blots</li> </ul>	
applications	<ul> <li>Sequencing reactions</li> </ul>	

#### **Intended Use**

For 96-well plate purification of genomic DNA from liquid blood samples. This protocol has been developed for up to 60  $\mu L$  of human or animal whole blood (EDTA-, Citrate- or Heparin-stabilized) or buffy coats. Erythrocytes from non-mammals (e.g. birds, fish, reptiles) contain DNA. Here, a volume of 5-10  $\mu L$  of blood per purification is recommended.

#### **Typical Results**

Table 1.

Sample Type	Typical Yield (μg/200μl)
EDTA-Stabilized Human	2-10
Citrate-Stabilized Human	2-4
Heparin-Stabilized Human	1-3
Buffy Coat	5-25
Mouse Blood	3-5
Chicken Blood	3-15
Tilapia Blood	5-10
Dried Blood Spots	1-15

Typical yields reflect  $\mu g$  of gDNA per 200  $\mu l$  of initial sample. Due to sample variability, results may vary.

Figure 1.

Yield and Quality of gDNA Isolated from Whole Blood Using GenElute™-E

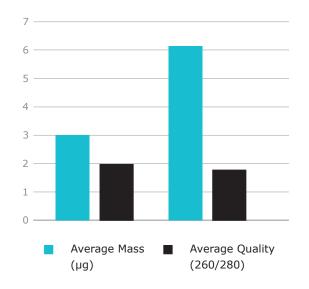


Table 2.

	Average Concentration (ng/µl)	Average Quality (260/280)	Average Mass (µg)
Tilapia Blood	49.87	1.97	4.99
Chicken Blood	61.23	1.82	6.12

Average spectrophotometric results of from forty-eight replicates of gDNA isolation from chicken blood and thirty-two replicates of gDNA isolation from tilapia blood using GenElute $^{\text{TM}}$ -E Single Spin Blood DNA 96 Kit. Graph shown in Figure 1. Due to sample variability, results may vary.

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#### Storage and Stability

#### Kit Storage

Store SmartLyse™ B Protease ② and Purification Plates at 2-8 °C. The remaining components should be stored at room temperature. Use the kit within 12 months of receipt.

#### Sample Storage and Variability

Nucleic acid degrades over time, potentially leading to reduced fragment length and overall yield. Therefore, it is best if samples are acquired from fresh material. Often when working with samples, this is not feasible. Stabilizing the sample through the addition of stabilizing reagents can assist in improving results. Be aware that the DNA yield to be expected is donor-dependent.

#### **Disposal**

GenElute<sup>™</sup>-E kits adhere to the principles of "SMASH Packaging", our plan that drives improvement in the sustainability of our packaging through less packaging, more sustainable materials and easier recycling.

The box and insert material comes from sustainably managed forests and/or more than 70% of recycled content. The kit componet bags are composed of starch-based, compostable material. Please recycle.

Kit components exposed to samples should be disposed of with biological waste. Other kit materials should be disposed of according to all applicable international, federal, state, and local regulations.

### **Materials and Equipment Needed**

#### **Kit Contents**

- Lysis Plate: 96-well plate for lysis of blood samples in a 96-well thermal shaker.
- Purification Plate: 96-well plate containing the resin matrix for DNA purification.
- DNA Storage Plate: 96-well plate for the collection of the purified DNA.
- Adhesive Foil for plate sealing.
- Reagents:
  - Blood Lysis Buffer
  - SmartLyse<sup>™</sup> B Protease <sup>¹</sup>
  - Clearing Solution B
  - 1x Tris Buffer ①.

#### **Not Supplied in Kit**

- Conditioning Plate: 96-deep well plate with minimum of 800 µL well volume for the collection of void volume during preparation of the Purification Plate. Reusable.
- 96-well swing-out centrifuge.

**Important:** Switch centrifuge to relative centrifugal force, rcf (x g); if this is not possible please use formula to calculate the conversion of round per minute (rpm) into rcf. Most centrifuges offer the choice between rpm and g-force (rcf); if not, calculate the rpm matching the g-force using the formula:

rpm = 1,000 x  $\sqrt{(g/(1.12 \times r))}$ , where r = radius of rotor in mm and g is the required g-force.

• 96-well Plate Thermal Shaker with agitation, capable of heating to 60 °C and 80 °C.

Alternative: Heating Block or heat chamber.

- · Vortex device.
- Pipets for 10 μL, 200 μL, and 1,000 μL scales, corresponding pipet tips.
- 8-channel pipet for 200 μL scale, corresponding pipet tips.
- Troughs for Master Mix preparation holding > 10 mL.
- Balance Plate(s) to be used in the centrifuge in case an odd number of plates are being processed.

#### Preparation before starting

- Heat the thermal shaker or heating block/chamber to 60 °C.
- Set the microcentrifuge to 1,000 x g.

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#### Standard Protocol

#### Lysis

For each sample, transfer 50 μL Blood Lysis
 Buffer 
 and 10 μL SmartLyse™ B Protease Mix 
 .

If working with more than two samples, prepare a Lysis Master Mix with 20% excess volume for the number of blood samples (see table).

Number of samples	1	96 (+20%)
Blood Lysis Buffer 🕕	50 μL	5,760 μL
SmartLyse™ B Protease •	10 μL	1,152 μL
Final Volume	60 µL	6,912 μL

Add 60  $\mu L$  of the Lysis Master Mix per well of Lysis Plate.

- 2. Add up to 60  $\mu L$  of blood sample per well of the Lysis Plate.
- 2. Seal Lysis Plate tightly with Adhesive Foil.
- 3. Place the Lysis Plate in the thermal shaker and incubate at 60 °C for 30 minutes with maximum agitation.

If using Heating Block or heat chamber, vortex halfway through incubation time to re-suspend, and return to incubation.

Lysis time can be shortened to 15 minutes without loss in PCR performance but A260/A230 purity ratio may be lowered.

**Note:** Agitation during lysis is mandatory for blood samples (in contrast to other sample types).

Meanwhile during lysis, proceed with "Preparation of Purification Plate".

- 4. After incubation at 60 °C, increase the temperature to 80 °C and incubate for additional 10 minutes with maximum agitation.
- 5. After having performed lysis, detach the Adhesive Foil from the incubated Lysis Plate and add 10  $\mu$ L Clearing Solution B  $\odot$  to each well of the Lysis Plate. Mix by pipetting up and down. The sample will become cloudy.

**Note**: Usually, the addition of RNase is not required due to endogenous RNases. If RNA needs to be stringently degraded, add 1  $\mu$ L RNase before the addition of Clearing Solution B  $^{\odot}$  and incubate for 2 minutes at room temperature.

**Note**: For extraction from buffy coat, pipet the lysate up and down 10 times before proceeding to centrifugation.

6. Centrifuge Lysis Plate for 3 minutes at maximum speed.

#### **Preparation of Purification Plate**

7. Carefully detach the lower and upper sealing foils from the Purification Plate.

**Note**: If the Purification Plate was not shipped or stored upright, resin may stick to the upper foil. In this case, horizontally shake plate until resin is removed from upper foil.

- Plate preparation: Place the Purification Plate on top of the Conditioning Plate (a 96-deep well plate with a minimum well volume of 800 μL, not supplied) and centrifuge for 1 minute at 1,000 x g to collect the void buffer from the Purification Plate. Discard the flow-through ("void volume") collected in the Conditioning Plate (Conditioning Plate can be re-used).
- 9. Place conditioned Purification Plate on top of the Storage Plate for collection of purified DNA.

#### **Purification of DNA**

- 10. Transfer the lysis supernatant containing the DNA into the prepared Purification Plate. Important loading instructions:
  - Using the 8-channel pipette, carefully obtain the supernatant containing the DNA. Avoid any cellular debris at the bottom of the wells as it may clog the pipette tips. It is recommended to use wide-bore pipette tips for this step.

**Note:** Residual blood precipitate may be loaded and will not interfere with purification.

- Slowly and vertically release the supernatant onto the middle of the resin surface.
- Do not punch pipette tip into the resin bed during loading of supernatant.
- 11. Centrifuge the Purification Plate on top of the Storage Plate for 1 minute at 1,000 x g. The purified DNA flows through the Purification Plate into the Storage Plate. Discard the Purification Plate.

The collected DNA can be used immediately or kept at 2-8 °C or for long-term storage at -20 °C. For spectrophotometric analysis, use the 1x Tris Buffer  $\odot$  supplied with the kit.

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# **Troubleshooting**

Problem  Probable Cause  Individual samples have inherent variability. In addition, there is variability across different sample types. Optimization needs to be performed by the user to validate for their sample type.  Using too much sample may result in overloading the Purification. Using variability across different sample type.  Using too little of sample may result  Use the recommended sample load.  Optimization needs to be performed by user to validate for their sample type.  Use the recommended sample load.  Optimization needs to be performed by user to validate for their sample type.	fresh itions or gate low the
variability. In addition, there is variability across different sample types. Optimization needs to be performed by the user to validate for their sample type.  Using too much sample may result in overloading the Purification Plate's capacity for separation.  Use the recommended sample load. Optimization needs to be performed by user to validate for their sample type.  Use the recommended sample type.  Use the recommended sample load.  Use the recommended sample load.	fresh itions or gate low the
in overloading the Purification Plate's Optimization needs to be performed by capacity for separation.  Use the recommended sample load.	
LICING TOO LITTLE OF CAMPIE MAY PACIFIT	the
in low sample yields.  Optimization needs to be performed by user if their sample type is low yielding	
Small, possibly degraded, DNA- fragments (< 60 bp) and/or RNA (if RNase was used) are removed during purification. Since these components are co-purified with silica-based kits, there may be artificially lower oD 260 readings with GenElute™-E kits. Thus, the calculation of sample concentration and subsequent yield may appear lower.  Quantitation by measuring signal intent of bands via gel electrophoresis fragment separation, using a fragment analyzer, comparing qPCR Ct values will provide reliable measurement of full-length gDI	nt or a more
Centrifugation speeds and spin times have been optimized to acquire the fraction of sample containing the nucleic acid.  Verify that centrifugation was performe the recommended conditions.	d under
Incomplete lysis of the sample may lead to reduced yields. Lysis times may need to be extended depending on the sample type.  Additionally, the SmartLyse™ enzymes optimized to work at the recommended temperature. Verify that the heating until thermal shaker) is heating correctly.	
If the preparation steps were performed incorrectly, then the separation resin will be packed incorrectly.  Verify that the preparation steps for the Purification Plate was performed according to the protocol.	
Loading too low of sample or too high of sample may result in sample volume loss. The loaded sample volume is required to be within 90-110 $\mu$ L as that volume is required to displace the Purification Plate buffer.  Low sample  Low sample  Loading too low of sample or too high onto the Purification Plate is below 90 $\mu$ bring the sample within the recommendary available to be loaded onto the Purification Plate is above 110 $\mu$ l (impacting results only load up to the recommended volume)	ol, then ded volume tion s), then
Blocking of pipette tips by lysate debris during aspiration of the supernatant may result in a lower transfer volume and, consequently, in a lower DNA yield.  Loading of debris onto the Purification on timpact the ability of the Purification to purify the nucleic acid from the same However, avoid aspirating the debris in pipette tip or use wide-bore tips to aspir the sample has an abundance of debric prevent clogging	Plate ole. to the irate
260/230 ratios appear to be "too low."  In some cases, the 260/230 ratios may be below the recommended range.  Downstream assays have not been show be compromised by lower 260/230 ration nucleic acid isolated using GenElute™-F	os using

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The purified sample has a reddish color.	In these cases, the Purification Plate may be overloaded due to variations in sample type.	Reduce the amount of sample used with the kit.
	Reddish coloration may indicate incomplete digestion for the sample type.	Try increasing the lysis time.
Purification Plates with dried resin	In rare cases, the Purification Plates dry out during storage. This may be due to not storing the plates according to the recommended conditions.	Store GenElute™-E kits according to the recommended conditions.
Purification Plates with leaks	Although rare, improper sealing or too robust of turbulence can break the seal of the covering of the Purification Plates.	Ensure Purification Plates are efficiently sealed before introducing them to any agitation. If problem persists, reduce the turbulence by reducing the rpm.

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#### **Product Ordering**

Description	Qty	Catalogue No.	
	10	EC100-10RXN	
GenElute™-E Single Spin Blood DNA Kit	50	EC100-50RXN	
DIOOG DIVA KIL	250	EC100-250RXN	
	10	EC200-10RXN	
GenElute™-E Single Spin Blood DNA High Yield Kit	50	EC200-50RXN	
	250	EC200-250RXN	
	10	EC300-10RXN	
GenElute™-E Single Spin Tissue DNA Kit	50	EC300-50RXN	
	250	EC300-250RXN	
	10	EC400-10RXN	
GenElute™-E Single Spin Cell Culture DNA Kit	50	EC400-50RXN	
30 3a 3 2	250	EC400-250RXN	
	10	EC500-10RXN	
GenElute™-E Single Spin Plant DNA Kit	50	EC500-50RXN	
Traile BNA Ric	250	EC500-250RXN	
	10	EC600-10RXN	
GenElute™-E Single Spin DNA Cleanup Kit	50	EC600-50RXN	
DNA Cicanap Kit	250	EC600-250RXN	
GenElute™-E	10	EC700-10RXN	
Organic Solvent DNA	50	EC700-50RXN	
Cleanup	250	EC700-250RXN	
	10	EC800-10RXN	
GenElute™-E Single Spin RNA Cleanup Kit	50	EC800-50RXN	
Tara Cicaria Paric	250	EC800-250RXN	
GenElute™-E FFPE	10	EC900-10ML	
Deparaffinization Solution	100	EC900-100ML	
GenElute™-E	100	EC111-100ML	
Tissue Stabilizer	500	EC111-500ML	
GenElute™-E	1	EC222-1EA	
RNA Gel Loading Buffer	5	EC222-5EA	
GenElute™-E Single Spin	2	EC396-2EA	
Tissue DNA 96 Kit	8	EC396-8EA	
GenElute™-E Single Spin	2	EC596-2EA	
Plant DNA 96 Kit	8	EC596-8EA	
GenElute™-E Single Spin	2	EC196-2EA	
Blood DNA 96 Kit	8	EC196-8EA	
GenElute™-E Single Spin Cap Puncher	1	EC9999-1EA	

#### **Precautions and Disclaimer**

This product is for research use only. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

#### **Notice**

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# GenElute™-E Single Spin Checklist for Blood DNA 96 Kit

#### **Preparation Before Starting EC196**

- Heat the thermal shaker or heating block/chamber to 60 °C.
- Set the centrifuge to  $1,000 \times g$ .

## Lysis

Prepare Lysis Master Mix, add 60 µL of the Lysis Master Mix per well of Lysis Plate.

Number of samples	1	96 (+20%)
Blood Lysis Buffer (1)	50 μL	5,760 μL
SmartLyse™ B Protease ©	10 μL	1,152 μL
Final Volume	60 µL	6,912 μL

Add up to 60 µL of blood sample per well of Lysis Plate. Seal with Adhesive Foil.

Plate **Preparation** during lysis

Incubate







- Detach Adhesive Seal and add 10 µL Clearing Solution B . Mix by pipetting.
- Centrifuge Lysis Plate for 3 minutes at maximum speed.

# Preparation of Purification Plate (during 60 °C and 80 °C incubation)

- Detach lower and upper sealing foils from Purification Plate.
- Place the Purification Plate on top of a Conditioning Plate.
- Centrifuge 1 minute at 1,000 x g to collect void buffer.
- Place conditioned Purification Plate on top of Storage Plate.

#### Purification of DNA

- Transfer lysis supernatant from Lysis Plate to Purification Plate.
- Centrifuge 1 minute at 1,000 x g to collect DNA into the Storage Plate.
- Collected DNA is ready to use.

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