

## Novabiochem®

Letters: 3/06

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# Product Focus: New reagents for peptide synthesis

## **NEW** Asn and Gln derivatives for Fmoc SPPS

Fmoc-Asn(Dmcp)-OH

Fmoc-Gln(Dmcp)-OH

### Features & Benefits

- Dmcp provides complete protection to side-chain carboxamide
- Dmcp is more easily cleaved than Trt group
- Faster coupling than Trt-protected derivatives
- Higher solubility than Trt-protected derivatives

Since the introduction of Fmoc-Asn(Trt)-OH and Fmoc-Gln(Trt)-OH by Sieber [1], they have become the standard derivatives for the routine synthesis of asparagine- and glutamine-containing peptides by Fmoc SPPS. These derivatives, however, do have certain limitations. Firstly, cleavage of the N-Trt group can be sluggish, particularly in the case of N-terminal Asn residues which can require over 4 hours for complete Trt removal with 95% TFA [2]. Secondly, these derivatives have low solubility in DMF compared to other Fmoc-protected amino acids. Thirdly, protected peptides containing Asn(Trt) and Gln(Trt) often have poor solubility. Finally, Asn(Trt) in particular



is notorious for inducing aggregation during solid phase synthesis.

Novabiochem® is, therefore, pleased to introduce L. A. Carpino's N-dimethylcyclopropylmethyl (Dmcp) derivatives of Fmoc-protected asparagine and glutamine [3]. Cleavage of the Dmcp group from the side-chains of Asn and Gln is rapid, even when the residue is located at the N-terminus of a peptide. Coupling of Dmcp-protected derivatives appears to be faster than that of the corresponding hindered Trt derivatives: for example, coupling of Fmoc-Asn(Trt)-OH with TFFH in the synthesis of ACP gave sigificant quantities of des-Asn peptide, whereas with Fmoc-Asn(Dmcp)-OH none of this by-product was generated [3]. Furthermore, protected peptides containing Asn(Dmcp) and Gln(Dmcp) residues appear to have enhanced solubility. This observation would indicate that peptides containing these residues would be less prone to aggregation during SPPS. Finally, Fmoc-Asn(Dmcp)-OH and Fmoc-Gln(Dmcp)-OH are also more soluble in DMF than Fmoc-Asn(Trt)-OH and Fmoc-Gln(Trt)-OH, thereby facilitating coupling reactions at higher concentration.

## *NEW* orthogonally-protected Gluderivative for Fmoc SPPS

Fmoc-Glu-O-2-PhiPr

### Features & Benefits

- 2-PhiPr group is removed with 1% TFA in DCM
- Ideal tool for on-resin synthesis of head-to-tail cyclic peptides

Fmoc-Glu-O-2-PhiPr is an excellent tool for the synthesis of head-to-tail cyclic peptides by Fmoc SPPS, as the O-2-PhiPr group is stable to bases but can be selectively cleaved on-resin with 1% TFA in DCM, in the presence of standard tBu-based protecting groups [4]. Typically, the amino acid is anchored via the side-chain  $\gamma$ -carboxyl group to Wang or Rink amide-type resins. Following chain extension, the  $\alpha$ -carboxyl is unmasked by treatment with 1% TFA in DCM, and on-resin cyclization can be carried out (Figure 1).

Fig. 1: Synthesis of cyclic peptides via side-chain anchoring of Glu-0-2-PhiPr.

For side-chain to tail lactam-bridged peptides, the combination of Lys(Mtt) and Glu-0-2-PhiPr is particularly advantageous since both side-chain protecting groups can be removed in a single step.

## *NEW* reagent for the synthesis of labeled peptides

Fmoc-Asp(EDANS)-OH

### Features & Benefits

- Direct synthesis of peptide fluorescent substrate containing EDANS
- Compatible with Fmoc SPPS
- TFA cleavage provides labeled peptide directly

The EDANS/Dabcyl fluorophore-quencher pair is one of the most commonly used for FRET applications, owing to excellent spectral overlap between the emission spectrum of EDANS ( $\lambda_{ex}$  341 nm,  $\lambda_{em}$  471 nm) and absorbance spectrum of Dabcyl ( $\lambda_{max}$  453 nm) [5, 6] (Figure 2). Quenching of the fluorescence of EDANS by Dabcyl is consequently highly efficient, with up to 40-fold enhancements in fluorescence having been observed upon proteolysis of Dabcyl/EDANS-labeled peptides [5].

To incorporate EDANS within the peptide chain, the simplest approach is to use Fmoc-Asp(EDANS)-OH or Fmoc-Glu(EDANS)-OH during peptide assembly [7, 8]. Introduction

of the analogous Glu derivative during SPPS has been achieved using PyBOP®/DIPEA activation in conjunction with an extended coupling time [8]. Powerful acylating reagents such as PyBrOP® should be avoided as their use can lead to acylation of the naphthylamine nitrogen.

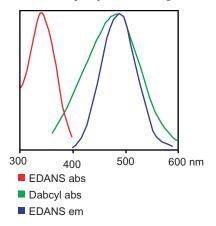


Fig. 2: Absorbance and emission spectra of Dabcyl and EDANS [7].

04-12-1288	Fmoc-Asp(EDANS)-OH	500 mg
NEW		1 g
04-12-1291	Fmoc-Glu(EDANS)-OH	500 mg
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# *NEW* resins for the synthesis of P1-labeled protease substrates

### Fmoc-Asp(Wang resin)-AFC

### Fmoc-Asp(Wang resin)-pNA

Fmoc-Lys(carbamate Wang resin)-AMC

#### Features & Benefits

- Direct synthesis of peptide fluorescent substrates containing C-terminal Asp-pNA, Asp-AFC or Lys-AMC
- Compatible with Fmoc SPPS
- TFA cleavage provides labeled peptide directly
- Ideal for peptide substrate optimization and enzyme profiling

P1-labeled peptide substrates are the most commonly used tools for studying protease activity and specificity [9]. They comprise a peptide in which the carboxyl group of the P1 residue of the protease substrate recognition sequence forms an amide bond with a dye. When proteolysis of this amide bond occurs, the dye is released enabling the process to be detected by changes in optical density or fluorescence (Figure 3).

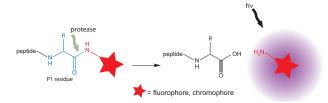


Fig. 3: Principle of P1-labeled fluorogenic substrates.

Fmoc-Asp(Wang resin)-pNA, Fmoc-Asp(Wang resin)-AFC and Fmoc-Lys(carbamate Wang resin)-AMC are the latest editions to Novabiochem's range of pre-loaded resins for the synthesis of P1-labeled peptides. These resins are ideal tools for the production of enzyme substrate libraries for protease profiling as their use allows aspartyl p-nitroanilides and 7amido-4-trifluoromethylcoumarins, and lysyl 7-amido-4methylcoumarins, respectively, to be prepared directly by Fmoc SPPS. They are extremely simple to use as they are fully compatible with standard Fmoc SPPS protocols. The Fmoc group is removed with 20% piperidine in DMF under standard conditions, and the free amine group can be acylated with Fmoc amino acids activated with PyBOP® or TBTU. Following peptide assembly, cleavage with 95% TFA releases the labeled peptide directly from the solid support without any additional steps.

Table 1: Optical properties of dyes used in P1-labeled substrates.

Dye	λ <sub>max/ex</sub> (nm)	<b>E</b> (M <sup>−1</sup> cm <sup>−1</sup> )	λ <sub>em</sub> (nm)
pNA	405	10,500	
AMC	342	16,000 (354 nm)	441
AFC	400	12,600 (380 nm)	505

04-12-3918 NEW	Fmoc-Asp(Wang resin)-AFC	500 mg 1 g
04-12-3919 NEW	Fmoc-Asp(Wang resin)-pNA	500 mg 1 g
04-12-3917 <i>NEW</i>	Fmoc-Lys(carbamate Wang resin)-AMC	500 mg 1 g
04-12-3912	Fmoc-Arg(bis-Boc-resin)-AMC	500 mg
04-12-3915	Fmoc-Asp(Wang resin)-AMC	500 m
		1 g

# **NEW** Bifunctional amino-PEG-acid spacer

Boc-NH-(PEG)<sub>27</sub>-COOH (88 atoms)

#### Features & Benefits

- 88 atom amino-PEG-acid spacer
- Introduced using standard activation methods
- Imparts solubility to end-product
- Yields homogeneous product

Boc-NH-(PEG) $_{27}$ -C00H is the latest addition to Novabiochem's range of PEG-based building blocks for solid phase peptide synthesis. It can be introduced using standard coupling methods, such as PyBOP® or TBTU.

This derivative is prepared from highly purified monodisperse PEG to ensure an homogeneous product free from contaminating oligomers. Unlike similar PEG linkers based on polydisperse PEGs, products prepared using this reagent will be a single chemical entity and can therefore be characterized and purified using standard techniques.

01-63-0151 NEW	Boc-NH-(PEG) <sub>27</sub> -COOH (88 atoms)	500 mg 1 g
01-63-0141	Fmoc-NH-(PEG) <sub>2</sub> -COOH (20 atoms)	500 mg 1 g 5 g
01-63-0103	N <sub>3</sub> -(PEG) <sub>7</sub> -COOH (33 atoms)	500 mg 1 g
01-63-0102	Boc-NH-(PEG) <sub>6</sub> -COOH (30 atoms)	500 mg 1 g
01-63-0109	Fmoc-NH-(PEG) <sub>11</sub> -COOH (40 atoms)	500 mg 1 g
01-63-0150	Fmoc-NH-(PEG) <sub>27</sub> -COOH (88 atoms)	500 mg

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