



**ALL-IN-ONE™
RANDOM PRIME DNA LABELING MIX (-dATP)**

Product No. **R7022/R9522**
Technical Bulletin No. MB-500
June 1998

Product Information

TECHNICAL BULLETIN

Product Description

Sigma's All-In-One™ Random Prime DNA Labeling Mix is based on the method developed by Feinberg and Vogelstein.^{1,2} The sample DNA is denatured and allowed to anneal with random 9-mers, which serve as primers for the Klenow fragment of DNA Polymerase I. Deoxyadenosine triphosphate (dATP) modified with ³²P-, digoxin, fluorescein, or biotin, is incorporated into the newly synthesized DNA strand. This method allows the labeling of very small quantities of DNA and of DNA fragments that have been separated in low melting point agarose to high specific activities. After labeling, the DNA may be used as a probe in various hybridization applications.

Each All-In-One random prime reaction tube is ready for use after reconstitution with denatured sample DNA and the label of choice. Labeled nucleotide is not included and must be obtained separately.

Reagents Provided

Each tube is sufficient for labeling 10 ng-1 µg of template DNA

- **Random Prime Reaction Tubes,**
Product No. R7022 (1.5 ml tubes) or
Product No. R9522 (0.2 ml tubes)
Each reaction tube contains dCTP, dGTP, TTP,
random 9-mers, and Klenow fragment of DNA
polymerase I (15 units) lyophilized with reaction
buffer
- **Control DNA,** Product No. D7684 1 vial
λ Hind III, 125 ng/vial

Reagents That May be Required But Not Provided (Sigma product numbers are given where appropriate)

- 0.5 M molecular biology grade EDTA, Product No. E7889
- 100% trichloroacetic acid (TCA), Product No. 490-10
- α-³²P, biotin, digoxin, or FITC labeled dATP
- Glass fiber filter, >2.3 µm retention, or equivalent, e.g. Product No. F5019
- Molecular biology water, Product No. W4502
- 100 X TE Buffer (1 M Tris-HCl, pH 8, 100 mM EDTA), Product No. T9285
- Disodium pyrophosphate, Product No. P8135
- tRNA, Product No. R8508
- Vacuum manifold, Product No. M2536

Precautions

Sigma's All-In-One Random Prime DNA Labeling Mix is for laboratory use only. Not for drug, household or other uses. If the sample DNA is radioactively labeled, standard procedures for safely handling radioactive materials should be followed.

Storage

Store all kit components desiccated at room temperature. After reconstitution, store the control DNA at 2-8°C.

2

Reagent Preparation

1. Dilute 100X TE buffer concentrate 1:100 with molecular biology grade water to prepare 1X TE buffer.
2. Add 225 μ l of 1X TE buffer to the control DNA vial. The solution will contain approx. 0.56 ng/ μ l DNA. After reconstitution, store the solution at 2-8°C.
3. For labeling using 32 P-dATP, prepare 10% (v/v) trichloroacetic acid (TCA) containing 2% (w/v) disodium pyrophosphate and 5% (v/v) trichloroacetic acid (TCA) containing 2% (w/v) disodium pyrophosphate. Chill the solution containing 10% TCA on ice until needed.

Procedures

I. Standard Labeling with 32 P-dATP

A. Labeling Reaction

1. Prepare 10 ng-1 μ g linearized template DNA in a total volume of 45 μ l of 1X TE (see Note 1).
2. Denature the template DNA by heating at 95°C for 3 minutes and then immediately placing on ice for 2 minutes.
3. Reconstitute one of the All-In-One random prime reaction tubes by adding the 45 μ l denatured template DNA.
4. Add 5 μ l 32 P-dATP (10 μ Ci/ μ l; 3,000 Ci/mmol) to the tube. Mix briefly.
5. Incubate at 37°C for 15 minutes.
6. Stop the reaction by adding 2 μ l of 0.2 M EDTA, pH 8.0. The prepared probe is ready for use at this point.

Note 1. The DNA may be prepared in a smaller volume. However, note that the final volume in the All-In-One random prime reaction tube should be 50 μ l.

B. Determine the Precipitable Radioactivity

1. Dilute a 5 μ l aliquot of the labeled DNA mixture with 95 μ l of 10 mM EDTA.
2. Place a 5 μ l aliquot of the diluted DNA mixture from Step 1 into a tube containing 25 μ g of carrier tRNA.
3. Add 1 ml of ice cold 10% (v/v) trichloroacetic acid containing 2% (w/v) disodium pyrophosphate.
4. Invert to mix and centrifuge briefly.
5. Incubate on ice for 15 minutes.
6. Filter reaction onto a glass fiber filter disk using a vacuum manifold.
7. Wash the filter disk five times with 2 ml of 5% (v/v) trichloroacetic acid containing 2% (w/v) disodium pyrophosphate.
8. Remove filter and dry by air or under a heat lamp.
9. Place filter in scintillation cocktail and count in a liquid scintillation counter to determine the precipitable radioactivity (P).

C. Determine the Total Radioactivity

Spot a second 5 μ l aliquot of the diluted DNA mixture from Step B1 onto another glass fiber filter disk and dry without the intervening TCA/disodium pyrophosphate washes. Count in a liquid scintillation counter to determine the total radioactivity (T).

D. Determine the Specific Activity

Calculation for Specific Activity (SA) in dpm/ μ g:

$$SA = \frac{(\mu\text{Ci})(2.2 \times 10^9)(F)}{D_i + (1.3 \times 10^3)(F)(\mu\text{Ci/S})}$$

μ Ci = μ Ci of dATP in reaction

F = Fraction of input label incorporated into DNA:
Precipitated Samples (P)/Total Counts(T)

D_i = Mass of input template DNA (ng)

S = Specific activity of dATP (Ci/mmol or μ Ci/nmol)

2.2×10^9 = Factor to convert to dpm from μ Ci

1.3×10^3 = Average molecular weight of the four dNMP's
(4×325)

Using this procedure, a specific activity of 1×10^9 dpm/ μ g control DNA template is obtained within 15 minutes at 37°C. The procedure labels 25 ng of control DNA with 50 μ Ci of ^{32}P -dATP (3,000 Ci/mmol).

II. Labeling with biotin, digoxin, or fluorescein modified nucleotides

The following protocol is designed to label 25 ng of denatured DNA using biotin-7-dATP.

1. Prepare 10 ng-1 μ g linearized template DNA in a total volume of 45 μ l of 1X TE.
2. Denature the template DNA in a microcentrifuge tube by heating for 2-3 minutes to 95°C, then immediately place on ice for 2 minutes.
3. Reconstitute one of the All-In-One random prime reaction tubes by adding the 45 μ l denatured template DNA.
4. Add 5 μ l of 1 mM biotin-7-dATP for a final concentration of 0.1 mM.

5. Mix gently and incubate at 37°C for 1 hour.

6. Stop the reaction by adding 2 μ l of 0.2 M EDTA, pH 8.0. The prepared probe is ready for use at this point.

III. Labeling DNA in low melting point agarose

1. The DNA fragment to be labeled is carefully excised from a 1% low-melting agarose gel and transferred to a 1.5 ml tube.
2. Add deionized water in the ratio of 3 ml per g of agarose gel and heat the tube for 7 minutes at 100°C to melt the gel and denature the DNA.
3. Cool the solution to 37°C. The DNA/agarose mixture may now be used for labeling as described in the previously described labeling procedures.

Results

Procedural notes

1. If plasmid DNA is to be labeled it must be linearized first.
2. This procedure was optimized for labeling 25-50 ng of DNA. For labeling larger quantities (e.g. 1 μ g), lower specific activities should be expected.
3. DNA probes labeled by this procedure can be used in blot hybridizations without removing the unincorporated nucleoside triphosphate. The labeled probe may be removed by ethanol/ammonium acetate precipitation (Add 1/2 volume of 7.5 M ammonium acetate and 2 volumes of 100% ethanol to the mixture. Centrifuge and repeat the precipitation step once).

References

1. Feinberg, A.P. and Vogelstein, B., *Anal. Biochem.*, **132**, 6 (1983)
2. Feinberg, A.P. and Vogelstein, B., *Anal. Biochem.*, **132**, 266 (1984)

Sigma brand products are sold through Sigma-Aldrich, Inc.

Sigma-Aldrich, Inc. warrants that its products conform to the information contained in this and other Sigma-Aldrich publications. Purchaser must determine the suitability of the product(s) for their particular use. Additional terms and conditions may apply.

Please see reverse side of the invoice or packing slip.