

Product Information

MediaBoost

Non-animal Sterile Microbial Growth Supplement

MBD0055

Product Description

Studying microbial species via cutting edge genomic techniques such as next generation sequencing (NGS) has become increasingly prevalent in recent years; however, genomic methods alone cannot fully replace experimental testing of microbial species.^{1,2} Therefore, the ability to culture microbes in a laboratory setting remains essential for comprehensive study of these communities.

Some microbial species can be challenging to culture in standard culture media.³ Often species of interest or a specific process require specialized media compositions, to provide an optimal environment for maximal proliferation and yield.⁴ A few examples include:

- Chemically defined minimal growth media are often used to grow bacteria for proteomics, as a base for stable isotope enrichment in NMR protein structure-function studies⁵, but growth can be limited without the addition of an effective supplement.⁶
- Use of chemically defined minimal growth media in the production of secreted microbial proteins to ensure reproducibility and a higher concentration of the microbial-produced protein of interest.^{7,8}

MediaBoost is a patented (US 10,696,942), ready-to-use, non-animal, protein free, microbial growth media supplement that amplifies the growth of gram-positive and gram-negative bacteria, and yeast species in minimal and/or standard microbial growth media*. It can also induce spore germination in some species.

* MediaBoost is compatible with some but not all strains and media combinations and should be tested on a case-by-case basis by the researcher for experiment optimization. The first table in Application Data shows some of the strains and media combinations that have been tested.

Features and Benefits

- Boosts growth of microorganisms
- Reduces germination time
- Ready-made sterile solution
- Protein-free

Precautions and Disclaimer

This product is for R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

Store this product refrigerated (2-8 °C).

Preparation Instructions

The product is provided as a convenient, ready-made sterile solution. No preparation or filtering is necessary before use. MediaBoost is sterile and should be added to the culture medium after autoclaving, at a 1:30-1:300 dilution. The ideal dilution of MediaBoost may vary for each specific strain and experimental optimization by the researcher may be required.

Application Data

Compatibility of MediaBoost with Relevant Species and Culture Media.

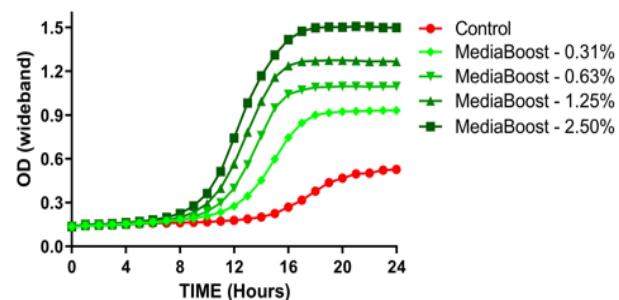
(✓) indicates that a relative increase in growth was observed when compared to media without MediaBoost, and (x) indicates that no growth improvement was observed with MediaBoost. Colored cells indicate media and species not tested together with MediaBoost.

Microbe/ Media Type	LB	1/3 LB	BM2	M9	IMDM	DMEM	YPD	1/3 YPD	MRS	1/3 MRS
<i>Bacillus subtilis</i>	✓				✓	✓				
<i>Lactobacillus rhamnosus</i>	✓	✓	x	x		✓			✓	✓
<i>Lactobacillus reuteri</i>					✓				✓	✓
<i>Lactobacillus salivarius</i>						✓			✓	✓
<i>Bacillus coagulans</i>					✓					
<i>Lactococcus lactis</i>					✓					
<i>Escherichia coli</i>	x				✓					
<i>Saccharomyces cerevisiae</i>	x		✓	✓	✓		✓	✓		
<i>Pseudomonas aeruginosa</i>	x			✓						

Growth of *Bacillus subtilis* Spores in LB

Bacillus subtilis, a gram-positive species, were cultured in LB media with increasing concentrations of MediaBoost. An untreated neat culture served as a negative control. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.

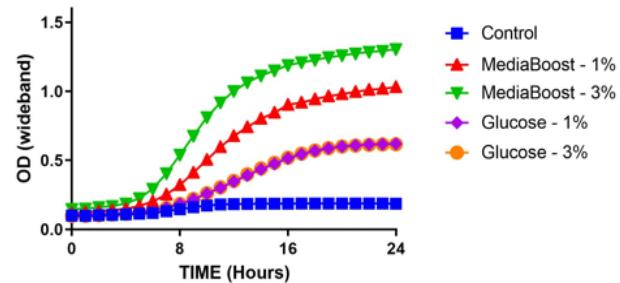
Growth of *Bacillus subtilis* spores in LB



Growth of *Lactobacillus rhamnosus* in 1/3rd LB

Lactobacillus rhamnosus, a gram-positive species, were cultured in 1/3 LB media with 1% or 3% MediaBoost. An untreated neat culture and cultures with an equivalent amount of added 50% glucose solution served as negative controls. MediaBoost increases growth more effectively than glucose, indicating that MediaBoost offers a more complex supplemental advantage than serving as a simple carbon source. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.

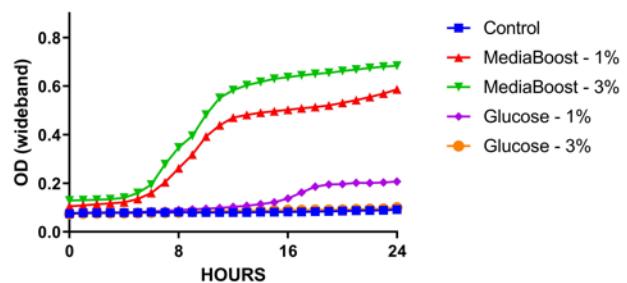
Growth of *Lactobacillus rhamnosus* in 1/3rd LB



Growth of *Pseudomonas aeruginosa* in M9

Pseudomonas aeruginosa, a gram-negative species, were cultured in M9 minimal culture media with 1% or 3% MediaBoost. An untreated neat culture and cultures with an equivalent amount of added 50% glucose solution served as negative controls. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.

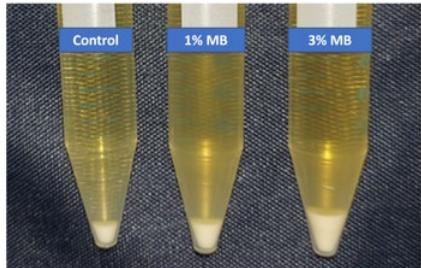
Growth of *Pseudomonas aeruginosa* in M9



Growth of *Saccharomyces cerevisiae* in YPD

Saccharomyces cerevisiae were cultured in YPD media for 24 hours at 32°C in a shaker flask, with 1% (middle tube) or 3% (right tube) MediaBoost (MB). An untreated neat culture served as a negative control (left tube). Aliquots were removed and tubes were centrifuged at high speed to obtain pellets.

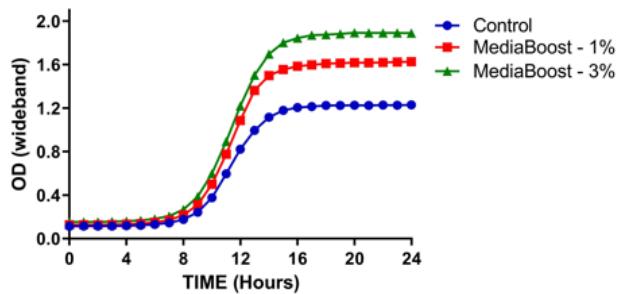
Growth of *Saccharomyces cerevisiae* in YPD



Growth of *Saccharomyces cerevisiae* in 1/3rd YPD

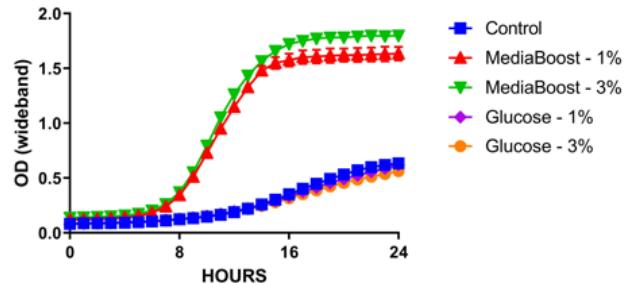
Saccharomyces cerevisiae were cultured in 1/3 YPD media with 1% or 3% MediaBoost. The addition of MediaBoost can substantially reduce the amount of basal medium required for growth such that one-half to one-third medium can be used. This facilitates not only cost reduction, but also more efficient downstream processing of microbial products.⁷ An untreated neat culture served as the negative control in this experiment. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.

Growth of *Saccharomyces cerevisiae* in 1/3rd YPD



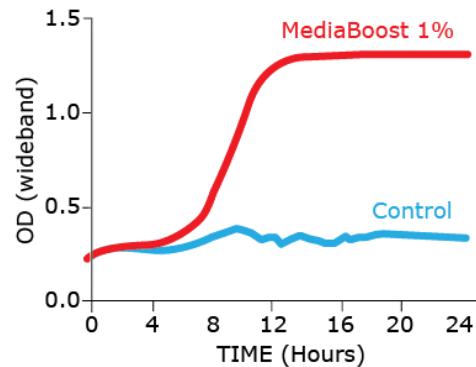
Growth of *Saccharomyces cerevisiae* in BM2

Saccharomyces cerevisiae were cultured in BM2 minimal media with 1% or 3% MediaBoost. An untreated neat culture and cultures with an equivalent amount of added 50% glucose solution served as negative controls. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.



Growth of *Saccharomyces cerevisiae* in IMDM

Saccharomyces cerevisiae was cultured in IMDM media with MediaBoost. MediaBoost can be added to protein-free media traditionally used for eukaryotic cell culture, such as DMEM and IMDM, to make microbial culture possible in these media as well. An untreated neat culture served as the negative control in this experiment. Data points represent means of triplicate cultures and S.D. did not exceed 10% of mean.



References

1. Tidjani Alou, M. et al. State of the Art in the Culture of the Human Microbiota: New Interests and Strategies. *Clin. Microbiol. Rev.* 34, (2020).
2. Lewis, W. H., Tahon, G., Geesink, P., Sousa, D. Z. & Ettema, T. J. G. Innovations to culturing the uncultured microbial majority. *Nat. Rev. Microbiol.* 19, 225–240 (2021).
3. Lagier, J.-C. et al. Current and Past Strategies for Bacterial Culture in Clinical Microbiology. *Clin. Microbiol. Rev.* 28, 208–236 (2015).
4. Bonnet, M., Lagier, J. C., Raoult, D. & Khelaifia, S. Bacterial culture through selective and non-selective conditions: the evolution of culture media in clinical microbiology. *New Microbes New Infect.* 34, 100622 (2020).
5. Thakur, C. S., Brown, M. E., Sama, J. N., Jackson, M. E. & Dayie, T. K. Growth of wildtype and mutant *E. coli* strains in minimal media for optimal production of nucleic acids for preparing labeled nucleotides. *Appl. Microbiol. Biotechnol.* 88, 771–9 (2010).
6. Azatian, S. B., Kaur, N. & Latham, M. P. Increasing the buffering capacity of minimal media leads to higher protein yield. *J. Biomol. NMR* 73, 11–17 (2019).
7. Tseng, C.-L. & Leng, C.-H. Influence of medium components on the expression of recombinant lipoproteins in *Escherichia coli*. *Appl. Microbiol. Biotechnol.* 93, 1539–1552 (2012).
8. Gąciarz, A. et al. Efficient soluble expression of disulfide bonded proteins in the cytoplasm of *Escherichia coli* in fed-batch fermentations on chemically defined minimal media. *Microb. Cell Fact.* 16, 108 (2017).
9. Reitermayer, D., Kafka, T. A., Lenz, C. A. & Vogel, R. F. Interrelation between Tween and the membrane properties and high pressure tolerance of *Lactobacillus plantarum*. *BMC Microbiol.* **18**, 72 (2018).

Notice

We provide information and advice to our customers on application technologies and regulatory matters to the best of our knowledge and ability, but without obligation or liability. Existing laws and regulations are to be observed in all cases by our customers. This also applies in respect to any rights of third parties. Our information and advice do not relieve our customers of their own responsibility for checking the suitability of our products for the envisaged purpose.

The information in this document is subject to change without notice and should not be construed as a commitment by the manufacturing or selling entity, or an affiliate. We assume no responsibility for any errors that may appear in this document.

Technical Assistance

Visit the tech service page at SigmaAldrich.com/techservice.

Terms and Conditions of Sale

Warranty, use restrictions, and other conditions of sale may be found at SigmaAldrich.com/terms.

Contact Information

For the location of the office nearest you, go to SigmaAldrich.com/offices.

Patent Information

This product is covered by several patents and patent applications including US 16/899949, US 10696942, CH 2773744, DE 602012054980.7, and related foreign patents DK/EP/ES/FR/GB/IE/IT/NL/SE 2773744.

The life science business of Merck KGaA, Darmstadt, Germany operates as MilliporeSigma in the U.S. and Canada.

MilliporeSigma, and Sigma-Aldrich are trademarks of Merck KGaA, Darmstadt, Germany or its affiliates. All other trademarks are the property of their respective owners. Detailed information on trademarks is available via publicly accessible resources.

© 2024 Merck KGaA, Darmstadt, Germany and/or its affiliates. All Rights Reserved.
MBD055pis Rev 04/24

Millipore
Sigma