



## Product Information

### SIGMA QUALITY CONTROL TEST PROCEDURE

#### Enzymatic Assay of ARYL ACYLAMIDASE (EC 3.5.1.13)

##### PRINCIPLE:

N-Acetyl-p-Aminophenol + H<sub>2</sub>O  $\xrightarrow{\text{Aryl Acylamidase}}$  p-Aminophenol + Acetate

**CONDITIONS:** T = 37°C, pH = 9.0, A<sub>615nm</sub>, Light path = 1 cm

**METHOD:** Colorimetric

##### REAGENTS:

- A. 100 mM Tris HCl Buffer, pH 9.0 at 37°C  
(Prepare 100 ml in deionized water using Trizma Base, Sigma Prod. No. T-1503. Adjust to pH 9.0 at 37°C with 1 M HCl.)
- B. 100 mM N-Acetyl-p-Aminophenol Solution (NAAP)  
(Prepare 50 ml in Reagent A using 4-Acetamidophenol, Sigma Prod. No. A-5000.)
- C. 5.0 mM p-Aminophenol Standard Solution (Std)  
(Prepare by dissolving 54.56 mg of p-Aminophenol, Sigma Prod. No. A-4076, in 80 ml of deionized water. Adjust the pH to 11.0 at 25°C with 0.1 M NaOH. The solution should be clear and purple at this point. Adjust the pH back to 9.0 using 0.1 M HCl. The color of the solution should turn brown. Quantitatively transfer to a 100 ml volumetric flask and dilute to 100 ml with deionized water. Protect the solution from light and use immediately after preparation.)
- D. 270 mM Ammonium Hydroxide Solution (NH<sub>4</sub>OH)  
(Prepare 100 ml in deionized water using Ammonium Hydroxide, Sigma Prod. No. A-6899.)
- E. 1.28 mM Cupric Sulfate Solution (CuSO<sub>4</sub>)  
(Prepare 100 ml in Reagent D using Cupric Sulfate, Pentahydrate, Sigma Prod. No. C-7631.)
- F. 0.41% (v/v) o-Cresol Solution  
(Prepare 48.2 ml by adding 0.2 ml of o-Cresol, Sigma Prod. No. C-7400 to 48 ml of deionized water. **PREPARE FRESH.**)

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**REAGENTS:** (continued)

- G. Color Reagent Solution (CRS)  
(Prepare 54.2 ml by adding 48.2 ml of Reagent F to 6 ml of Reagent E. Mix well. Prepare fresh and use within 30 minutes.)
- H. Aryl Acylamidase Enzyme Solution  
(Immediately before use, prepare a solution containing approximately 0.13 - 0.25 unit/ml of Aryl Acylamidase in cold Reagent A.)

**PROCEDURE:**

Pipette (in milliliters) the following reagents into suitable containers:

	<u>Test</u>	<u>Blank</u>
Reagent A (Buffer)	-----	0.10
Reagent B (NAAP)	2.90	2.90

Mix by swirling and equilibrate to 37°C. Then add:

Reagent H (Enzyme Solution)	0.10	-----
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Immediately mix by inversion and incubate at 37°C for exactly 10 minutes.

Pipette (in milliliters) the following reagents into suitable containers:

Reagent G (CRS)	2.50	2.50
Test Mixture	1.00	-----
Blank Mixture	-----	1.00

Mix by swirling and incubate at room temperature for exactly 5 minutes. Transfer the solutions to suitable cuvettes and record the  $A_{615nm}$  for the Test and Blank using a suitable spectrophotometer.

**Standard Curve:**

Prepare a standard curve by pipetting (in milliliters) the following reagents into suitable containers:

	<u>Std 1</u>	<u>Std 2</u>	<u>Std 3</u>	<u>Std 4</u>	<u>Std 5</u>	<u>Std Blank</u>
Reagent C (Std)	0.02	0.04	0.06	0.08	0.10	---
Deionized Water	0.08	0.06	0.04	0.02	---	0.10
Reagent B (NAAP)	2.90	2.90	2.90	2.90	2.90	2.90

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**PROCEDURE:** (continued)

Mix by swirling and incubate at 37°C for exactly 10 minutes.

Pipette (in milliliters) the following reagents into suitable containers:

	<u>Std 1</u>	<u>Std 2</u>	<u>Std 3</u>	<u>Std 4</u>	<u>Std 5</u>	<u>Std Blank</u>
Reagent G (CRS)	2.50	2.50	2.50	2.50	2.50	2.50
Std 1	1.00	---	---	---	---	---
Std 2	---	1.00	---	---	---	---
Std 3	---	---	1.00	---	---	---
Std 4	---	---	---	1.00	---	---
Std 5	---	---	---	---	1.00	---
Blank	---	---	---	---	---	1.00

Mix by swirling and incubate at room temperature for exactly 5 minutes. Transfer the solutions to suitable cuvettes and record the  $A_{615nm}$  for the Standards and Standard Blank using a suitable spectrophotometer.

**CALCULATIONS:**

Standard Curve:

$$\Delta A_{615nm} \text{ Standard} = A_{615nm} \text{ Standard} - A_{615nm} \text{ Standard Blank}$$

Plot the  $\Delta A_{615nm}$  Standard vs  $\mu\text{moles}$  of p-Aminophenol.

Sample Determination:

$$\Delta A_{615nm} \text{ Sample} = A_{615nm} \text{ Test} - A_{615nm} \text{ Test Blank.}$$

Determine the  $\mu\text{moles}$  of p-Aminophenol liberated using the Standard curve.

$$\text{Units/ml enzyme} = \frac{(\mu\text{moles of p-Aminophenol released})(df)}{(10)(0.1)}$$

df = Dilution factor

10 = Time (in minutes) of assay as per the Unit Definition

0.1 = Volume (in milliliter) of enzyme used in the assay

$$\text{Units/mg solid} = \frac{\text{units/ml enzyme}}{\text{mg solid/ml enzyme}}$$

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**CALCULATIONS:** (continued)

$$\text{Units/mg protein} = \frac{\text{units/ml enzyme}}{\text{mg protein/ml enzyme}}$$

**UNIT DEFINITION:**

One unit will convert 1.0  $\mu$ mole of N-acetyl-p-aminophenol (acetaminophen) to p-aminophenol per minute at pH 9.0 at 37°C.

**FINAL ASSAY CONCENTRATIONS:**

In a 3.00 ml reaction mix, the final concentrations are 100 mM Tris, 97 mM N-acetyl-p-aminophenol, and 0.0125 - 0.025 unit aryl acylamidase.

**REFERENCE:**

Price, C.P., Hammond, P.M., and Scawen, M.D. (1983) *Clinical Chemistry* **29**, 358-361

**NOTES:**

1. This assay is based on the cited reference.
2. Where Sigma Product or Stock numbers are specified, equivalent reagents may be substituted.

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