User Guide

mPAGE® TurboMix Bis-Tris Gel Casting Kit

TMKIT-10 TMKIT-60 TMRES-216ML TMSTK-120ML

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Introduction

The mPAGE® TurboMix Bis-Tris Gel Casting Kit is comprised of a Resolving Solution and a Stacking Solution. These solutions have been optimized to simplify gel preparation and minimize reagent waste. The mPAGE® TurboMix Bis-Tris Gel Casting Kit can be used with either Quick Cast or Traditional Casting methods.

The Quick Cast procedure polymerizes in one step by pouring the stacking gel immediately after the resolving gel. mPAGE® TurboMix Resolving Solution is provided at 20% acrylamide and formulated for dilution with deionized water, which allows the flexibility to cast different resolving gel percentages. The mPAGE® TurboMix Stacking Solution is provided at 4% acrylamide and requires only the addition of Ammonium persulfate (APS) and N,N',N'-tetramethylethane-1,2-diamine (TEMED). Review the entire user guide before first use.

Kit Components

- mPAGE® TurboMix Resolving Solution (20% acrylamide)
- mPAGE® TurboMix Stacking Solution (4% acrylamide)

Additional materials required (not provided)

- TEMED
- 10% APS
- Gel casting system such as mPAGE[®] Gel Caster or equivalent with compatible glass plates and combs
- 5 mL sterile serological pipettes
- MOPS-SDS running buffer or MES-SDS running buffer
- 4X LDS Sample Buffer
- mPAGE® MES-SDS or MOPS-SDS running buffer and mPAGE® 4X LDS Sample buffer can be used with mPAGE® TurboMix

Storage

Store bottles at 2-8 °C. Protect Resolving Solution and Stacking Solution from light. Storing solutions in kit box is recommended. See package label for expiration date.

Bring all solutions to room temperature before use.

Important:

- Bis-Tris gels are not compatible with Tris-Glycine-SDS running buffer. Only use MOPS-SDS or MES-SDS running buffers with this kit.
- Clean casting equipment of any acrylamide residue to ensure proper polymerization and inspect glass plates for nicks that could result in leakage.
- See page 3 to determine the volume of Resolving Solution and Stacking Solution for your gel size and thickness. Users may need to empirically determine the required volumes.



Volumes Required To Cast One Mini Gel

Below are the solution volumes needed to cast a single 7.4 x 8.2 cm mini gel in 1.0, 0.75, or 1.5 mm thicknesses. Volumes can be multiplied to cast several gels at once. When using different sized gel cassettes, the total volume required for casting can be determined empirically by filling cassette with water.

1 mm Thick Mini Gel

| Reso | ıvııı | G C I |
|------|-------|-------|

| Resulving dei | | | | |
|------------------------------------|--------|-------|--------|--------|
| Percentage | 8% | 10% | 12% | 15% |
| mPAGE® TurboMix Resolving solution | 2.4 mL | 3 mL | 3.6 mL | 4.5 mL |
| DI water | 3.6 mL | 3 mL | 2.4 mL | 1.5 mL |
| 10% APS | 30 uL | 30 uL | 30 uL | 30 uL |
| TEMED | 3 uL | 3 uL | 3 uL | 3 uL |
| Total | 6 mL | 6 mL | 6 mL | 6 mL |

| Stacking Gel Percentage | | |
|----------------------------|--|--|
| | | |

| Percentage | 4% |
|--------------------------------------|-------|
| mPAGE® TurboMix Stacking solution | 2 μL |
| DI water | |
| 10% APS | 20 μL |
| TEMED | 2 μL |
| Total | 2 mL |

0.75 mm Thick Mini Gel

Resolving Gel

| Percentage | 8% | 10% | 12% | 15% |
|---------------------------------------|---------|---------|---------|---------|
| mPAGE® TurboMix Resolving solution | 1.8 mL | 2.25 mL | 2.7 mL | 3.38 mL |
| DI water | 2.7 mL | 2.25 mL | 1.8 mL | 1.12 mL |
| 10% APS | 22.5 uL | 22.5 uL | 22.5 uL | 22.5 uL |
| TEMED | 2.25 uL | 2.25 uL | 2.25 uL | 2.25 uL |
| Total | 4.5 mL | 4.5 mL | 4.5 mL | 4.5 mL |

Stacking Gel

| Percentage | 4% |
|--------------------------------------|--------|
| mPAGE® TurboMix Stacking solution | 1.5 mL |
| DI water | |
| 10% APS | 15 uL |
| TEMED | 1.5 uL |
| Total | 1.5 mL |

1.5 mm Thick Mini Gel

Resolving Gel

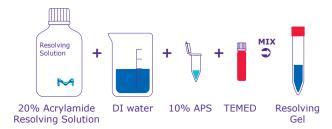
| itesorting ee | | | | |
|---------------------------------------|--------|--------|--------|---------|
| Percentage | 8% | 10% | 12% | 15% |
| mPAGE® TurboMix Resolving solution | 3.6mL | 4.5 mL | 5.4 mL | 6.75 mL |
| DI water | 5.4 mL | 4.5 mL | 3.6 mL | 2.25 mL |
| 10% APS | 45 uL | 45 uL | 45 uL | 45 uL |
| TEMED | 4.5 uL | 4.5 uL | 4.5 uL | 4.5 uL |
| Total | 9 mL | 9 mL | 9 mL | 9 mL |

Stacking Gel

| Percentage | 4% |
|--------------------------------------|-------|
| mPAGE® TurboMix Stacking solution | 3 mL |
| DI water | |
| 10% APS | 30 uL |
| TEMED | 3 uL |
| Total | 3 mL |

Quick Cast Instructions

- 1. Bring solutions to room temperature.
- 2. Clean and set up casting equipment.
- 3. Mark desired height of resolving gel, usually 0.5-1 cm below the bottom of the comb teeth.
- 4. Prepare 10% APS from powder. Note: 10% APS can be prepared fresh with every use or aliquoted and stored at -20 °C. Avoid repeated freeze-thaw cycles.
- 5. See <u>page 3</u> to calculate the volume of reagents required for the desired gel thickness.
 - Multiply volumes by the desired number of gels to cast multiple gels at once (up to 4 gels).
- 6. Prepare the desired resolving gel percentage by pipetting into a clean conical tube:

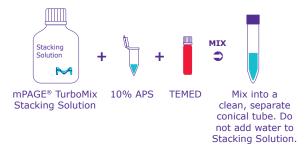


To avoid contamination, use a sterile serological pipette to draw mPAGE® TurboMix solutions from stock bottles.

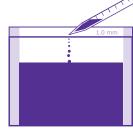
NOTE: Add 10% APS and TEMED immediately before casting. Gel will begin to polymerize after addition of APS and TEMED.

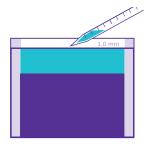
- 7. Gently mix reagents by inverting conical tube, avoiding introduction of air bubbles into the gel mixture.
- 8. Prepare the stacking gel by pipetting mPAGE®
 TurboMix Stacking Solution into a clean, separate
 conical tube and adding the required amount of
 TEMED and 10% APS.

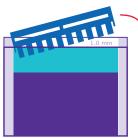
Note: Add 10% APS and TEMED immediately before casting. Gel will begin to polymerize after addition of APS and TEMED.



- Gently mix reagents by inverting conical tube, avoiding introduction of air bubbles into the gel mixture.
- Using a serological pipette, fill each cassette to marked height with resolving gel.
- 11. Position serological pipette at the middle of the cassette and gently add the stacking gel, filling to the top of the short plate. A dip may occur where pipetting takes place but will level out.
- Quickly and carefully insert the comb.
 Inserting at an angle may help to avoid trapping air bubbles below the teeth.
- 13. Allow gels to polymerize for 1 hour. Left over casting solution can be used to monitor polymerization.
- 14. Gels can be used immediately or wrapped in DI water-soaked paper towels and stored in an air-tight container at 4 °C for up to 4 weeks.



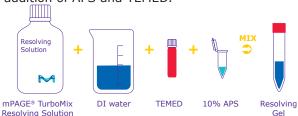




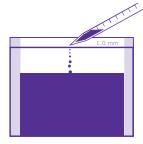
Traditional Cast Instructions

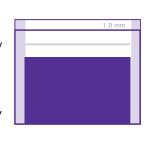
- 1. Bring solutions to room temperature.
- 2. Clean and set up casting equipment.
- 3. Mark desired height of resolving gel, usually 0.5-1 cm below the bottom of the comb teeth.
- Prepare 10% APS from powder. Note: 10% APS can be prepared fresh with every use or aliquoted and stored at -20 °C. Avoid repeated freeze-thaw cycles.
- 5. See page 3 to calculate the volume of reagents required for the desired gel thickness. Multiply volumes by the desired number of gels to cast multiple gels at once.
- Prepare the desired resolving gel percentage by pipetting mPAGE® TurboMix Resolving Solution, DI water, APS and TEMED into a clean conical tube. To avoid contamination, use a sterile serological pipette to draw mPAGE® TurboMix solutions from stock bottles.

NOTE: Add 10% APS and TEMED immediately before casting. Gel will begin to polymerize after addition of APS and TEMED.



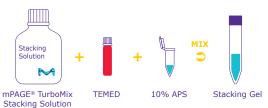
- Gently mix reagents by inverting conical tube, avoiding introduction of air bubbles into gel mixture.
- Using a serological pipette, fill each cassette to marked height with resolving gel.
- Carefully overlay
 with isopropanol.
 Allow resolving gel to
 polymerize, approximately
 30 minutes. A sharp line
 will appear below the
 isopropanol as acrylamide
 polymerizes. Alternatively,
 left over casting solution
 can be used to monitor
 polymerization.



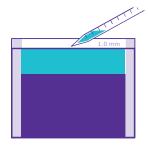


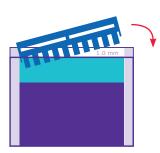
- 10. Remove the isopropanol and rinse with DI water. Blot with filter paper to remove residual water.
- 11. To prepare the stacking gel, pipette the required amount of mPAGE® TurboMix Stacking Solution, TEMED and 10% APS into a clean conical.

NOTE: Add 10% APS and TEMED immediately before casting. Gel will begin to polymerize after addition of APS and TEMED.



- 12. Gently mix reagents by inverting conical tube, avoiding introduction of air bubbles. Pipette stacking gel into cassettes and fill to the top of the short plate.
- 13. Quickly and carefully insert the comb. Inserting at an angle may help to avoid trapping air bubbles below the teeth.
- 14. Allow gels to polymerize for 1 hour.
- 15. Gels can be used immediately or wrapped in DI water-soaked paper towels and stored in an airtight container at 4 °C for up to 4 weeks.





Sample Preparation and Electrophoresis

1. Samples should be prepared just prior to electrophoresis, according to the table below.

Note: Do not store reduced samples for >2 hours as they may reoxidize.

| Reagent | Reduced Sample (µL) | Non-reduced sample (µL) |
|----------------------------|---------------------------|-------------------------------|
| Protein sample | X | X |
| 4X LDS Sample Buffer | 2.5 | 2.5 |
| 1M-DTT | 1 | N/A |
| Deionized water | 6.5-X | 7.5-X |
| Total Volume | 10 | 10 |

- 2. Heat samples for 10 minutes at 70 °C then briefly centrifuge. Do not boil samples.
- 3. Install gel into the electrophoresis tank and add MOPS or MES running buffer.
- 4. Remove the comb from the gel and gently rinse wells with running buffer.
- 5. Load samples. Close the tank with lid and connect leads to an appropriate power supply.
- 6. The gel can be run at a maximum of 200 V when using MOPS running buffer, and a maximum of 180 V when using MES running buffer.

Buffer Formulation

Use either MOPS or MES running buffer. Do not use Tris-Glycine running buffer with TurboMix gels.

4X LDS Sample Buffer

(MPSB-10ML or MPSB-250ML)

| Reagent | Amount |
|---|----------|
| Tris-HCl | 0.666 g |
| Tris-Base | 0.682 g |
| Lithium Dodecyl Sulfate (LDS) | 0.800 g |
| EDTA | 0.006 g |
| Glycerol | 4 g |
| Coomassie Brilliant Blue G250 (1% solution) | 0.75 mL |
| Phenol Red (1% solution) | 0.25 mL |
| Deionized water | to 10 mL |

1X mPAGE MOPS SDS Running Buffer

(MPMOPS)

| Reagent | Amount |
|-----------------|---------|
| MOPS | 10.46 g |
| Tris-Base | 6.06 g |
| SDS | 1.00 g |
| EDTA | 0.30 g |
| Deionized water | to 1 L |

1X mPAGE MES SDS Running Buffer

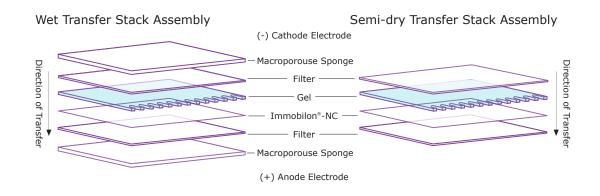
(MPMES)

| Reagent | Amount |
|-----------------|--------|
| MES | 9.76 g |
| Tris-base | 6.06 g |
| SDS | 1.00 g |
| EDTA | 0.30 g |
| Deionized water | to 1 L |

Protein Transfer Methods

Proteins can be transferred to PVDF (polyvinylidene difluoride) or nitrocellulose (NC) blotting membranes following protein separation. The membrane type used in the application may need to be determined experimentally based on the protein characteristics. It is recommended to use a low fluorescent blotting membrane such as the Immobilon FL when fluorescent detection is desired. Proteins can be transferred onto the blotting membrane by wet transfer, also known as tank transfer, semi-dry transfer, or fast semi-dry transfer.

Prepare fresh transfer buffer based on the type of transfer to be performed. For compatible buffer please refer to the table below. Depending on the protein of interest, choice of blotting membrane and transfer system used, optimization of the alcohol percent and transfer conditions may be required.



| | Wet transfer | Semi-Dry Transfer | Fast Semi-Dry Transfer |
|----------------------------------|---|---|---|
| Transfer system | mPAGE® Mini Wet Transfer System | Trans-Blot® SD Semi-Dry Transfer Cell (Bio-Rad) | Trans-Blot® Turbo™ Transfer System (Bio-Rad) |
| Transfer condition | 100V, 60 minutes | 25V, 30 minutes | 1.3 A, 25V limit, 7 minutes |
| Gel incubation prior to transfer | Optional | 5-10 minutes | No gel incubation |
| Filter paper | Immobilon® Blotting Filter Paper (IBFP0785C) | BioRad (1703966) | Part of transfer kit |
| Compatible Transfer buffer | 1X mPAGE® Transfer Buffer with 10% or 20% methanol | 2X mPAGE® Transfer Buffer with 10% methanol | Proprietary transfer buffer only available as part of a kit |
| | Tris-Glycine Buffer, pH 8.3 with 20% methanol | Tris-Glycine Buffer, pH 8.3 with 20% methanol | |
| | Bjerrum Schafer-Nielsen Buffer with 20% methanol | Tris-Glycine Buffer, pH 8.3 with 20% methanol, 0.025% SDS (best for high MW proteins) | _ |
| | | Bjerrum Schafer-Nielsen Buffer with 20% methanol | |

Product Ordering

Purchase online at <u>SigmaAldrich.com</u>.

| Description | Qty/Pk | Catalogue Number | | |
|--|-------------------------------------|-----------------------------|--|--|
| mPAGE TurboMix Bis-Tr | is Gel Cast | ting | | |
| mPAGE® TurboMix Bis-Tris Gel Casting Kit | ~10 mini gels | TMKIT-10 | | |
| mPAGE® TurboMix Bis-Tris Gel Casting Kit | ~60 mini gels | TMKIT-60 | | |
| mPAGE® TurboMix Resolving Solution | 216 mL | TMRES-216ML | | |
| mPAGE® TurboMix Stacking Solution | 120 mL | TMSTK-120ML | | |
| Gel Caster and Kits mPAGE® Gel Caster (2 pk) 1 GCR2 Contains: Gel Caster Base (2), Gel Caster Frame (2), Gel Caster Sealing Gaskets (2), Gel Releasers (5) | | | | |
| mPAGE® Gel Casting Kit, 0.75 mm Contains: Gel Caster (1), Co Combs 15 wells (5), Mini Sp Mini Short Plates (10) | 1 ombs 10 well pacer Plates (| MGCK-75M ls (5), (5), | | |
| mPAGE® Gel Casting Kit, 1.0 mm Contains: Gel Caster (1), Co Combs 15 wells (5), Mini Sp Mini Short Plates (10) | | | | |
| mPAGE® Gel Casting Kit, 1.5 mm Contains: Gel Caster (1), Co Combs 15 wells (5), Mini Sp Mini Short Plates (10) | | | | |
| mPAGE® Combs 0.75 mm, 10 wells | 5 | C75M10W | | |
| 0.75 mm, 15 wells | 5 | C75M15W | | |
| 1.0 mm, 10 wells | 5 | C1M10W | | |
| 1.0 mm, 15 wells | 5 | C1M15W | | |
| 1.5 mm, 10 wells | 5 | C15M10W | | |
| 1.5 mm, 15 wells | 5 | C15M15W | | |
| mPAGE® Mini Spacer Plates 0.75 mm | 5 | MSPA75 | | |
| 1.0 mm | 5 | MSPA10 | | |
| 1.5 mm | 5 | MSPA15 | | |
| mPAGE® Mini Short Plates | 10 | MSHRT | | |

| Description | Qty/Pk | Catalogue Number |
|--|------------|---------------------|
| mPAGE® Gel Caster Base | 1 | GCBASE |
| mPAGE® Gel Caster Frame | 1 | GCFRM |
| mPAGE® Gel Caster Sealing Gaskets | 5 | GCSEAL |
| mPAGE® Gel Releasers | 5 | GREL5 |
| mPAGE® Casting Plate Rack | 1 | CPS1 |
| Buffers | | |
| mPAGE™ 4X LDS | 10 mL | MPSB-10ML |
| Sample Buffer | 250 mL | MPSB-250ML |
| mPAGE™ MES SDS Running Buffer Powder (Each packet makes 1L) | 5 packets | MPMES |
| mPAGE™ MOPS SDS Running Buffer Powder (Each packet makes 1L) | 5 packets | MPM0PS |
| mPAGE™ Transfer Buffer Powder (Each packet makes 1L) | 10 packets | MPTRB |
| Protein Markers | | |
| mPAGE™ Color Protein Standard | 500 μL | MPSTD4 |
| mPAGE™ Unstained Protein Standard | 500 μL | MPSTD3 |
| mPAGE™ Western Protein Standard | 250 μL | MPSTD2 |
| Protein Gel Stains | | |
| EZBlue™ Gel Staining Reagent | 500 mL | G1041-500ML |
| ReadyBlue™ Protein Gel Stain | 1 L | RSB-1L |
| EZFluor™ 1-step Fluorescent Protein Gel Stain | 1 L | SCT145-1L |
| EZFluor™ UV 1-step Fluorescent Protein | 1 L | SCT147-1L |

Gel Stain

| Description | Qty/Pk | Catalogue Number |
|--|----------|---------------------|
| Reagents | | |
| Ammonium persulfate (APS) for molecular biology, for electrophoresis, ≥98% | 25 g | A3678-25G |
| N,N,N',N'- Tetramethylethylenediamine (TEMED) ReagentPlus®, 99% | 5 mL | T22500-5ML |
| DL-Dithiothreitol solution, 1 M | 10 mL | 43816-10ML |
| 2-Mercaptoethanol (BME) | 25 mL | 63689-25ML-F |
| Lithium dodecyl sulfate (LDS) | | L9781 |
| Sodium dodecyl sulfate (SDS) | | L3771 |
| Ethylenediaminetetraacetic acid (EDTA) | | E5134 |
| Ethylenediaminetetraacetic acid (MOPS) | | M1254 |
| 2-Morpholinoethanesulfonic acid monohydrate (MES) | | M3671 |
| SDS-PAGE and Trans | fer Syst | ems |

| SDS-PAGE | and | Transfer | Systems |
|-----------------|-----|-----------------|----------------|
|-----------------|-----|-----------------|----------------|

| mPAGE® Mini Gel Tank, makes 2 gels | 1 | MGT-2 |
|---------------------------------------|---------|-------|
| mPAGE® Mini Gel Tank, makes 4 gels | 1 | MGT-4 |
| mPAGE® Mini Wet Transfe System | er 1 | MWTS |

| Description | Qty/Pk | Catalogue Number | |
|---|------------|---------------------|--|
| Transfer Membranes | 5 | | |
| Immobilon®-E Membrane, PVDF, 0.45 µm, 8.5 cm x 10 m | 1 roll | IEVH85R | |
| Immobilon®-E PVDF Transfer Membranes, 7 cm x 8.4 cm sheet | 50 sheets | IEVH07850 | |
| Immobilon®-P Membrane, PVDF, 0.45 µm, 8.5 cm x 10 m | 1 roll | IPVH85R | |
| Immobilon®-P PVDF Transfer Membranes, 7 cm x 8.4 cm sheet | 50 sheets | IPVH07850 | |
| Immobilon®-FL Membrane PVDF, 0.45 µm, 8.5 cm x 10 m | 1 roll | IPFL85R | |
| Immobilon®-PSQ Membrane, PVDF, 0.2 μm, 8.5 cm x 10 m | 1 roll | ISEQ85R | |
| Immobilon®-PSQ PVDF Transfer Membranes, 7 cm x 8.4 cm sheet | 50 sheets | ISEQ07850 | |
| Immobilon®-NC Transfer Membrane, 0.45 µm, 8.5 cm x 10 m | 1 roll | HATF85R | |
| Immobilon®-NC Transfer Membrane, 0.45 µm, 7 cm x 8.4 cm sheet | 50 sheets | HATF07850 | |
| Power Supplies | | | |
| | US plug | MA400-US | |
| | Euro plug | MA400-EU | |
| mA400 Basic Power Supply | UK plug | MA400-UK | |
| | Japan plug | MA400-NI | |
| | China plug | MA400-ZH | |
| | US plug | MA700-US | |
| mA700 Essential Power Supply | Euro plug | MA700-EU | |
| | UK plug | MA700-UK | |
| | Japan plug | MA700-NI | |
| | China plug | MA700-ZH | |

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