Technical Data Sheet

# Pseudomonas Selective Broth – 2mL Liquid Media Ampoules
# Cat. No. MHA000P2P

This medium is recommended to enumerate Pseudomonas species in samples such as beer, bottled water, cider, soft drink, sport drink and wine.

**Mode of Action**
Pseudomonas Selective Broth is a selective medium used for the growth of Pseudomonas species and inhibits the growth of other bacteria by membrane filtration technique. Tryptone and gelatin peptone in the media supply nitrogenous and carbonaceous compounds, long chain amino acids, and other essential growth nutrients.

**Typical Composition (per liter of purified water)**

| | | | |
|---|---|---|---|
| Gelatin Peptone | 16.0 g | Glycerin | 10.0 ml |
| Casein Hydrolysate | 10.0 g | CFC Supplement, Oxoid SR 103E | |
| Potassium Sulfate | 10.0 g | Ethanol (95%, non-denatured) | 4.0ml |
| Magnesium Chloride | 1.4 g | | |

**Application**
1. Collect the liquid sample in a sterile container. Sodium thiosulfate is necessary when the liquid sample contains a residual disinfectant. The sample should be a 100 ml minimum. Dilution of the sample will be necessary if a high number of bacteria colonies are expected.
2. Invert one Pseudomonas Selective Broth ampoule 2 to 3 times. Open the ampoule. Remove the lid of a petri dish and carefully pour the contents equally onto the absorbent pad.
3. Set up the membrane filtration apparatus. Use sterile forceps to put the membrane filter in the assembly. The grid side is up.
4. Invert the sample / diluted sample for approximately 30 seconds to thoroughly mix the sample.
5. Pour the sample / diluted sample into the funnel. If the volume is less than 20ml, add 10 ml of sterile buffered dilution water to the funnel.
6. Apply the vacuum until the funnel is empty. Then stop the vacuum.
7. Rinse the funnel with 20ml to 30ml of sterile buffered dilution water. Apply the vacuum. Rinse the funnel two more times.
8. Stop the vacuum when the funnel is empty. Remove the funnel from the assembly. Use sterile forceps to lift the membrane filter.
9. Put the membrane filter on the absorbent pad. Let the membrane filter bend and fall equally across the absorbent pad to make sure that the air bubbles are not trapped below the filter.
10. Secure the lid on the petri dish and invert the dish.
11. Incubate the inverted petri dish for 24 - 72 hours at 25-35° C. Note: 35° C during 24 hours is favorable to Pseudomonas aeruginosa, 25° C for Pseudomonas species.
12. Remove the petri dish from the incubator. Use a microscope to count the number of bacteria colonies on the membrane filter.
13. Interpret and report the results.

**Results Reporting**
Report the colony density as the number of colonies in 100ml of sample. If there's more than 200 colonies, dilute the sample and use the diluted sample in the test procedure.

Colonies in 100ml = Colonies counted / ml of sample x 100**.**

**Storage and Shelf Life**
The product can be used until the expiry date if the unopened ampoules are stored sealed in the aluminum foil bag at 2 – 10°C.

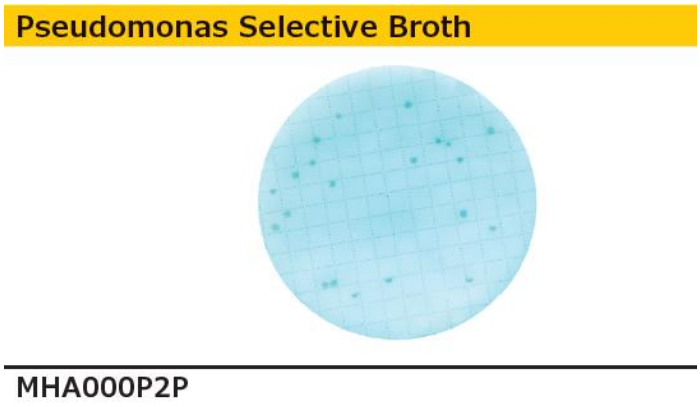
**Disposal**
Please dispose of used culture medium in accordance with local regulations (e.g. autoclave for 20 min at 121 °C, disinfect, incinerate etc.).

## Quality Control

| Function | Control Strains | Incubation | Reference Medium | Method of Control | Expected Results |
|---|---|---|---|---|---|
| Productivity | Pseudomonas aeruginosa ATCC® 27853 WDCM 00025 | 24 +/- 2 hours at 35 +/- 0.5° C | Previously validated batch of Pseudomonas Selective Broth | Quantitative | Recovery 85-115% Characteristic colonies |

Please refer to the actual batch specific certificate of analysis.

All growth on this medium indicate the presence of Pseudomonas species.
Colonies that are blue-green, brown or show fluorescence are presumptive of P. aeruginosa.




## Ordering Information

| Product | Cat. No. | Pack size |
|---|---|---|
| Pseudomonas Selective Broth | MHA000P2P | 50 x 2 mL plastic ampoules |

## Literature

American Public Health Association and American Water Works Association and Water Pollution Control Federation and Water Environment Federation (1915): Standard methods for the examination of water and wastewater.



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