

## Product Information

### 51723 LUCY® Molecular Weight Standard Kit

#### Kit components

- 43772 LUCY 565 solution (2 vials)
- S8445 SigmaMarker™, Wide Range (1 vial)
- Technical information sheet

#### Description

The "LUCY 565 Molecular Weight Standard Kit" contains the fluorescent protein stain LUCY 565 and a Molecular Weight Marker, consisting of 12 proteins. The MW-Marker allows the precise size determination of proteins on electrophoresis gels, and LUCY 565 allows a highly sensitive, easy, fast and robust staining procedure on various kinds of SDS gels.

LUCY 565 is suitable for neutral staining, i.e. a Western blot can be performed from a gel that has previously been stained with LUCY 565.

Protein staining by LUCY 565 does not interfere with subsequent MALDI-MS.

#### Storage

Protect from light; store kit at 4 °C; freezing is not recommended.

#### Molecular Weight Marker

The Sigma MW-Marker contains 12 proteins in the range of 6.5 – 200 kDa.

The vial contains the protein mixture in lyophilized form.

Preparation of the marker:

1. Add 100 µl water and reconstitute the marker by vortexing
2. Do not heat!
3. Immediately aliquot and store the unused portions at -20 °C. Repeated freezing and thawing of reconstituted SigmaMarker is not recommended.

#### LUCY 565

Spectral data:  $\lambda_{ex}$ =565 nm /  $\lambda_{em}$ =588 nm

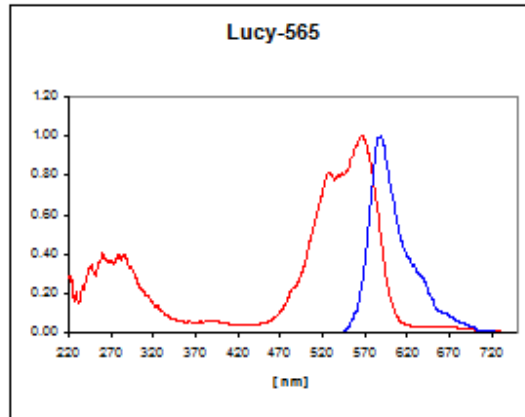
Contents: LUCY 565 is provided as a 5000 x stock-solution in DMSO (5 mg/ml)

Sensitivity: LOD: 5-10 ng/band

Linearity: Linear between 5 and 4000 ng/band

Handling: Warm to room temperature before opening. Do not expose to light unnecessarily.

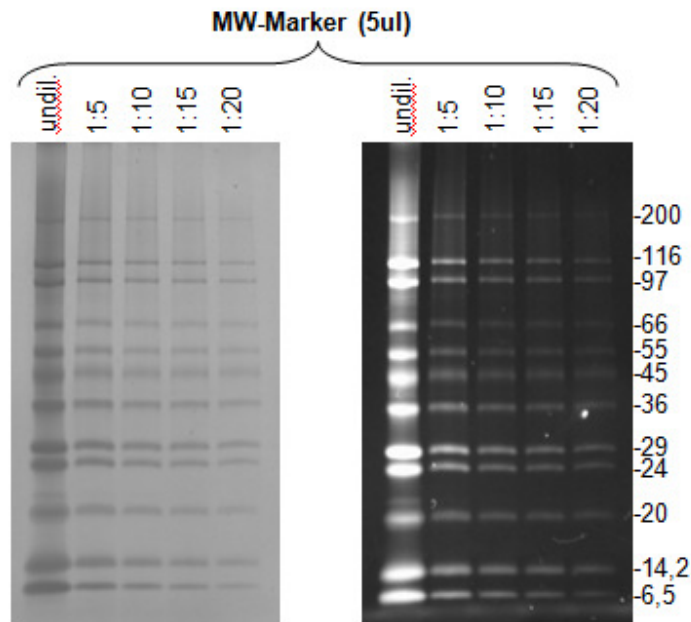
Reuse: Reuse of the dye will result in reduced sensitivity and is not recommended



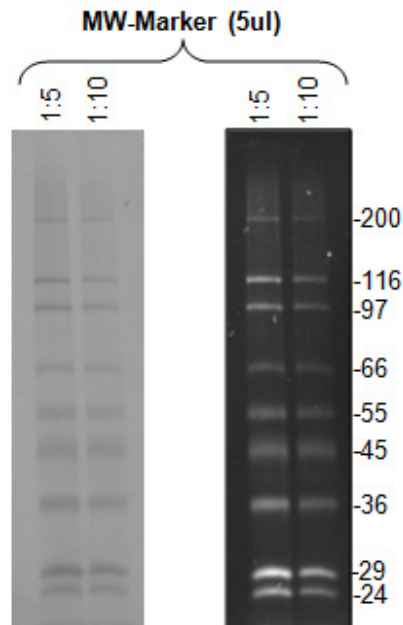
**Fig.1** Normalized fluorescence excitation (red) and emission (blue) spectra of LUCY 565 in the presence of BSA (0.2 mg/ml) and SDS (0.05 %)

**Standard protocol (staining of 1D Mini-Gels, or second dimension of 2D; 1 mm thickness)**

1. Add 5  $\mu$ l MW-Marker to a mini-gel (recommended dilution 1:5, see Fig.2)
2. Electrophoresis is performed under standard conditions (0.05 or 0.1 % SDS in the running-buffer)
3. After the run, the gel is immersed in 50 ml 1 x staining solution (10  $\mu$ l LUCY 565 in 50 ml water) for 60 min in the dark on a rocking table. Higher dye-concentrations will result in increased background staining
4. Rinse the gel with water for 30 s
5. Imaging



**Fig.2** 4-20 % Tris-Glycine gel. 5  $\mu$ l MW-Marker per lane in different dilutions, stained with LUCY 565. Left: imaged on FLA-3000 (ex 532 nm / em 580 nm), right: UV-screen, CCD-camera (Gel Logic 100), 590 nm filter.



**Fig. 3** 10 % Tris-Glycine gel. 5  $\mu$ l MW-Marker per lane in different dilutions, stained with LUCY 565. Left: imaged on FLA-3000 (ex 532 nm / em 580 nm), right: UV-screen, CCD-camera (Gel Logic 100), 590 nm filter.

### Additional staining protocols

#### Gel stains + Western Blotting:

It is possible to stain a gel and perform a Western blot afterwards:

- 1) Stain the gel with 10  $\mu$ l LUCY 565 in 50 ml water (not in acetic acid and without fixation before!)
- 2) The gel is immersed for 60 min in the dark on a rocking table
- 3) Short water rinse before imaging
- 4) Continue with Western blot transfer

It is not possible to use LUCY 565 for staining proteins on Western blot membranes.

#### Staining of large 2D-Gels:

- 1) The 2D-gel is stained for 2 h in the dark (80  $\mu$ l LUCY 565 in 400 ml 7.5 % acetic acid)
- 2) Destain for 30 s in 7.5 % acetic acid
- 3) Short water rinse before imaging

#### Staining of gels with a plastic backing:

Phast-gels or Dalt 12.5 gels may be stained using the standard protocol, however they show reduced sensitivity due to autofluorescence of the backing.

### Detection

Detection is performed by illuminating the gel on a UV-screen, and imaging the gel using a CCD-camera, e.g. Gel-Logic-100 (Kodak, 1-3 s, f-stop 3-5) with a 590 nm band-pass filter. Alternatively, a laser-scanner can be used (e.g. FLA-3000, Fuji), with 532 nm excitation and 580 nm emission-filter, or a Polaroid Camera. Other imaging systems are possible with the corresponding excitation sources and emission filter settings. Try to minimize the exposure to light, work quickly!

**Problems / interfering substances**

Do not use organic solvents during destaining or fixing (MeOH, EtOH), as it will strip off dye and SDS.

**Tested gel-systems**

- Tris-Glycine (Laemmli)
- Nupage<sup>®</sup> Bis-Tris (with MOPS-buffer)
- Dalt<sup>™</sup> 12.5 (GE)
- PhastGel<sup>™</sup> (GE)

**Related products**

Description	Cat. No.	Package size
LUCY 506	68721	500 µl
LUCY 565	43772	500 µl
LUCY 569	41629	500 µl
LUCY Starter Kit	51153	1 Kit
Laemmli Lysis-buffer, non-smelling	38733	5 x 2 ml

Nupage<sup>®</sup> is a registered trademark of Invitrogen.

Ettan<sup>™</sup> Dalt and PhastGel<sup>™</sup> are registered trademarks of GE Healthcare.

**Precautions and Disclaimer:**

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

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