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Product Information

PermaRed/AP Kit

Catalog Number **R3783**
Storage Temperature 2-8 °C

Product Description

PermaRed/AP is a chromogenic substrate which produces a strong red color and is used for immunohistochemical (IHC) or *in situ* hybridization (ISH) detection of secondary antibodies conjugated to alkaline phosphatase (AP). After the enzymatic reaction is completed, the resulting dye is insoluble in organic solvents including alcohol and xylene, which allows tissue sections stained with PermaRed/AP to be dehydrated, cleared and mounted after staining.

Components

PermaRed/AP Chromogen Catalog No. R3658	1 ml
PermaRed/AP Buffer Catalog No. B9813	30 ml
Dropper bottle Catalog No. R3908	1 bottle

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

Store at 2-8 °C. Store PermaRed/AP Chromogen protected from light.

Preparation Instructions

PermaRed/AP Working Solution: Add 3 ml of PermaRed/AP Buffer to the dropper bottle. Add one drop (~20 µL) of PermaRed/AP Chromogen solution to the dropper bottle. Replace the dropper tip and mix. Allow solution to reach room temperature before using. Use the PermaRed/AP Working Solution within 15 minutes of preparation. Discard any remaining solution after use.

Procedure

Please refer to primary antibody protocol. The secondary antibody must be conjugated to alkaline phosphatase (AP).

1. Wash tissue sections with 0.1-0.5 M Tris buffer, pH 7.5, containing 0.5% TWEEN® 20 or other appropriate wash buffer after conjugation with the AP-conjugated secondary antibody.

2. Remove excess buffer. Using the dropper bottle, add sufficient PermaRed/AP Working Solution to cover the tissue section.

3. Incubate at room temperature for 5-10 minutes. The time required for optimum staining may vary; for optimal results monitor the staining reaction through a microscope. When the desired staining result and signal-to-noise ratio are achieved, stop the enzymatic reaction by rinsing the slides in wash buffer.

4. Counterstain the sections using hematoxylin.

5. After staining, dehydrate the sections in alcohol. Clear the sections using xylene or xylene substitute and mount using permanent mounting medium.

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