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Product Information

NEK9 (1-979) (deleted fragment 346-731), active, GST tagged, human PRECISIO® Kinase recombinant, expressed in *Sf*9 cells

Catalog Number **SRP5053** Storage Temperature –70 °C

Synonyms: NEK8, NERCC, NERCC1, MGC16714, MGC138306, DKFZp434D0935

Product Description

NEK9 is a member of the NEK family and has high homology to NEK1, NEK3, and NEK4. NEK9 is activated during mitosis and binds specifically to RAN GTPase and is a substrate for CDC2 phosphorylation. NEK9 plays a role in the control of mitotic progression and is regulated by CDC2 and RAN GTPase. Overexpression of both active and inactive variants of NEK9 is toxic to cells and inhibits cell division causing abnormal nuclear morphologies. NEK9 can catalyze the phosphorylation of recombinant NEK6 and NEK7 *in vitro* leading to its activation. This suggests that NEK9 may be responsible for activation of NEK6 and NEK7 during mitosis.²

Recombinant human NEK9 is the full length protein with fragment (346-731) deleted. It was expressed by baculovirus in *Sf9* insect cells using an N-terminal GST tag. The gene accession number is NM_033116. Recombinant protein stored in 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM glutathione, 0.1 mM EDTA, 0.25 mM DTT, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~115 kDa

Purity: 70–95% (SDS-PAGE, see Figure 1)

Specific Activity: 102–138 nmole/min/mg (see Figure 2)

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at $-70~^{\circ}$ C is recommended. After opening, aliquot into smaller quantities and store at $-70~^{\circ}$ C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Typical Lot 70–95% (densitometry)

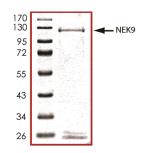
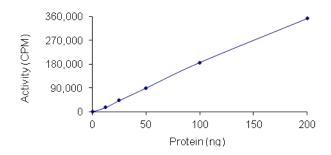


Figure 2.
Specific Activity of Typical Lot 102–138 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl₂, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/ul BSA.

Kinase Solution – Dilute the active NEK9 (0.1 μ g/ μ l) with Kinase Dilution Buffer to the desired concentration. Note: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active NEK9 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μ l aliquots at –20 °C.

 γ -³³P-ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of γ -³³P-ATP (1 mCi/100 μl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Dissolve the protein substrate in distilled water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ³³P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active NEK9, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ -33P-ATP Assay Cocktail may be thawed at room temperature.
- In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl:

10 μl of Kinase Solution 5 μl of Substrate Solution 5 μl of cold water (4 °C)

- 3. Set up a blank control as outlined in step 2, substituting 5 μl of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ - 33 P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- After the 15 minute incubation, stop the reaction by spotting 20 μl of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ - 33 P-ATP counts introduced into the reaction. Spot 5 μ l of the γ - 33 P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR =
$$\frac{\text{cpm of 5} \mu \text{l of } \gamma^{-33}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

cpm – value from control (step 7) nmole – 1.25 nmole (5 μl of 250 μM ATP Assav Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\Delta$$
cpm × (25/20)
SR × E × T

SR = specific radioactivity of the ATP (cpm/nmole ATP) Δ cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

20 = spot volume

T = reaction time (minutes)

E = amount of enzyme (mg)

References

- Roig, J. et al., Nercc1, a mammalian NIMA-family kinase, binds the Ran GTPase and regulates mitotic progression. Genes Dev., 16, 1640-1658 (2002).
- Belham, C. et al., A mitotic cascade of NIMA family kinases: Nercc1/Nek9 activates the Nek6 and Nek7 kinases. J. Biol. Chem., 278, 34897-34909 (2003).

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