

### **ProductInformation**

# SIGMA QUALITY CONTROL TEST PROCEDURE

## Enzymatic Assay of L-TYROSINE DECARBOXYLASE (EC 4.1.1.25)

### PRINCIPLE:

L-Tyrosine L-Tyrosine Decarboxylase Tyramine + CO<sub>2</sub>

Abbreviation used:

PRP = Pyridoxal 5-Phosphate

**CONDITIONS:**  $T = 37^{\circ}C$ , pH 5.5

**METHOD:** Radiolabelled Stop Reaction

#### **REAGENTS:**

 A. 200 mM Sodium Phosphate Solution (Prepare 50 ml in deionized water using Sodium Phosphate, Dibasic, Anhydrous, Sigma Prod. No. S-0876.)

- B. 100 mM Citrate Phosphate Buffer, pH 5.5 at 37°C (Buffer) (Prepare 100 ml in deionized water using Citric Acid, Free Acid, Anhydrous, Sigma Prod. No. C-0759. Adjust to pH 5.5 at 37°C with Reagent A.)
- C. McIlvaine's Buffer, pH 5.5 at 37°C (Buffer)
  (Prepare using 50 ml of Reagent A and adjust to pH 5.5 at 37°C with Reagent B.)
- D. 2.65 mM L-Tyrosine Solution (Prepare 10 ml in Reagent C using L-Tyrosine, Free Base, Sigma Prod. No. T-3754. Heat (70°C) may be required in order to solubilize.)
- E. 0.3 mM Pyridoxal 5-Phosphate Solution (PRP) (Prepare 2 ml in Reagent C using Pyridoxal 5-Phosphate, Sigma Prod. No. P-9255.)
- F. <sup>14</sup>C[COOH] L-Tyrosine (Use <sup>14</sup>C[COOH] L-Tyrosine, 50 60 mCi/mmol, 50 μCi/ml.)
- G. L-Tyrosine Reaction Cocktail (L-Tyr)
  (Prepare by adding 0.05 ml of Reagent E to 2 ml of Reagent C.)

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### **REAGENTS:**

- H. 1 M Methylbenzethonium Hydroxide Solution (MBH) (Use Benzethonium Hydroxide, approximately 1.0 M solution in Methanol, Sigma Prod. No. B2156.)
- Filter Paper
   (1 cm x 2 cm strips of filter paper are prepared using Whatman 3 MM paper. Add 0.050 ml of Reagent G to each strip and allow to air dry. This step is to be carried out in a fume hood.)
- J. Vacutainer Tubes (Use Vacutainer Blood Tubes, 13 x 75 mm, Becton Dickinson.)
- K. 25% (w/v) Trichloroacetic Acid Solution (TCA) (Prepare 10 ml in deionized water using Trichloroacetic Acid, 6.1 N Solution, approximately 100% (w/v), Sigma Stock No. 490-10.)
- L. L-Tyrosine Decarboxylase Enzyme Solution (Immediately before use, prepare a solution containing approximately 0.005-0.02 units/ml of L-Tyrosine Decarboxylase in cold Reagent C.)
- M. Scintillation Cocktail (Scint Fluid)
   (Use Sigma-Fluor Universal LSC Cocktail for Aqueous Samples, Sigma Prod. No. S-4273.)

### PROCEDURE:

Radiolabelled CO<sub>2</sub> is trapped by adsorbing it on an organic solvent which is present on filter paper located above the reaction mixture in a vacutainer tube.

Pipette (in milliliters) the following reagents into suitable vacutainer tubes.

	<u>Test</u>	<u>Blank</u>	PRP <sup>1</sup>
Reagent G (L-Tyr) Reagent E (PRP) Reagent C (Buffer)	0.10 0.10 	0.10 0.10 	0.10  0.10
Equilibrate to 37°C. Then add:			
Reagent L (Enz Soln) Reagent C (Buffer)	0.10	0.10	0.10

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**PROCEDURE:** (continued)

Immediately place Reagent H (Filter Paper) in the top of the vacutainer tube and seal the tubes by replacing the stopper. Incubate for exactly 10 or 20 minutes. Then add by injecting with a syringe and needle through the stopper. (**Do not remove stopper to add TCA.**)

	<u>Test</u>	<u>Blank</u>	PRP <sup>1</sup>
Reagent K (TCA)	0.40	0.40	0.40

Mix by swirling. Allow to stand for 12-16 hours at room temperature to allow for all evolved  $CO_2$  to be absorbed onto the filter paper.

Remove filter papers from vacutainer tubes and place into scintillation vials containing 7 ml of Reagent L (Scint). This should be performed in a hood.

Potential DPM (disintegration per minute) are prepared by pipetting 0.05 ml of Reagent F (L-Tyr) onto Reagent H (Filter Paper). Place into scintillation vial containing 7 ml of Reagent L (Scint Fluid).

### **CALCULATIONS:**

Potential DPM/
$$\mu$$
mole = 
$$\frac{\text{DPM of potential}}{\text{Total }\mu\text{moles of L-Tyrosine}}$$
 Units/ml enzyme = 
$$\frac{(\text{DPM Test - DPM Blank})(\text{df})}{(T)(0.1)(\text{potential DPM/}\mu\text{mole})}$$
 DPM = Disintegration per minute df = Dilution factor T = Time (in minutes) of assay as per the Unit Definition 0.1 = Volume (in milliliter) of enzyme used 
$$\frac{\text{units/ml enzyme}}{\text{mg solid/ml enzyme}}$$

#### **UNIT DEFINITION:**

One unit will liberate 1.0 µmole of CO<sub>2</sub> for L-tyrosine per minute at pH 5.5 at 37°C.

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#### FINAL ASSAY CONCENTRATION:

In a 0.30 ml reaction mix, the final concentrations are 100 mM citric acid<sup>3</sup>, 0.86 mM L- tyrosine, 0.1 mM pyridoxal 5-phosphate, and 0.0005-0.002 units L-tyrosine decarboxylase.

### **REFERENCE:**

Sundaresan, P.R. and Coursin, D.B. (1970) Methods in Enzymology, XVIII, Part A, 509-512

### NOTES:

- 1. The enzyme activity is also measured without any Reagent D(PRP) being present.
- 2. This assay is based on the cited reference.
- 3. This concentration is only an estimate since it is not known what exact volume of sodium phosphate is required to adjust the pH to 5.5 at 37°C.
- Where Sigma Product or Stock numbers are specified, equivalent reagents may be substituted.

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