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Product Information

COT (30-397), active, GST-tagged, human
PRECISIO® Kinase
recombinant, expressed in *Sf*9 cells

Catalog Number **C1370**
Lot Number SLBB4605V
Storage Temperature –70 °C

Synonyms: MAP3K8, EST, ESTF, TPL2, Tpl-2, c-COT, FLJ10486

Product Description

COT is an oncogene that can activate both the MAP kinase and JNK kinase pathways. COT activates IκB kinases and induces the nuclear production of NF-κB. C-terminal catalytic domain of KSR2 associates with COT and KSR2 can negatively regulate the kinase activity of COT *in vitro*. Co-transfection of KSR2 with COT in cells leads to reduced COT-mediated ERK activation and COT-induced IL-8 production in a dose-dependent manner.[1] COT is one of the MAP kinase kinase kinases that regulates the ERK1/ERK2 pathway in response to IL-1. Blockage of expression of COT results in failure of IL-1 to induce an increase in IL-8 and MIP-1β mRNA levels.[2]

This recombinant product was expressed by baculovirus in *Sf*9 insect cells using an N-terminal GST-tag. The gene accession number is NM 005204. It is supplied in 50 mM Tris-HCl, pH 7.5, with 150 mM NaCl, 0.25 mM DTT, 0.1 mM EGTA, 0.1 mM EDTA, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~70 kDa

Purity: ≥70% (SDS-PAGE, see Figure 1)

Specific Activity: 860–1,164 nmole/min/mg
(see Figure 2)

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at –70 °C is recommended. After opening, aliquot into smaller quantities and store at –70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.
SDS-PAGE Gel of Lot Number SLBB4605V:
>75% (densitometry)

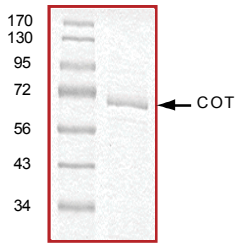
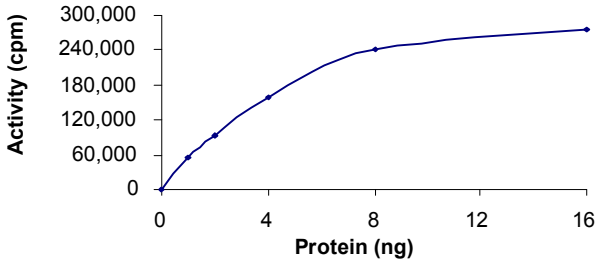


Figure 2.
Specific Activity of Lot Number SLBB4605V:
1,012 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM $MgCl_2$, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/µl BSA solution.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 µl aliquots at –20 °C.

Kinase Solution – Dilute the active COT (0.1 μg/μl) with Kinase Dilution Buffer to the desired concentration. Note: The lot-specific specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active COT kinase for optimal results.

γ-^{32}P-ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of γ-^{32}P-ATP (1 mCi/100 μl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Inactive MEK1 (0.2 μg/μl), Inactive ERK1 (0.2 μg/μl), Myelin Basic Protein (MBP) diluted in water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ^{32}P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

1. Thaw the active COT, Kinase Assay Buffer, Inactive MEK1, Inactive ERK1, and Kinase Dilution Buffer on ice. The γ-^{32}P-ATP Assay Cocktail may be thawed at room temperature.
2. In a pre-cooled microcentrifuge tube, prepare an activation mixture with a final volume of 20 μl:
   - 10 μl of Kinase Solution
   - 2 μl of inactive MEK1 (0.2 μg/μl)
   - 3 μl of inactive ERK1 (0.2 μg/μl)
   - 5 μl of Kinase Dilution Buffer
3. Start the activation reaction by adding 5 μl of 250 μM ATP and incubate in a water bath at 30 °C for 25 minutes.
4. In a microcentrifuge tube, add the following solutions to a volume of 20 μl on ice:
   - 5 μl of the activated mixture (step 3)
   - 5 μl of MBP Substrate Solution (4 °C)
   - 10 μl of cold water (4 °C)
5. Set up a blank control as outlined in step 4, substituting 5 μl of cold water (4 °C) for the Substrate Solution.
6. Initiate each reaction with the addition of 5 μl of the γ-^{32}P-ATP Assay Cocktail, bringing the final reaction volume to 25 μl. Incubate the mixture in a water bath at 30 °C for 15 minutes.
7. After the 15 minute incubation, stop the reaction by spotting 20 μl of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.
8. Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
9. Set up a radioactive control to measure the total γ-^{32}P-ATP counts introduced into the reaction. Spot 5 μl of the γ-^{32}P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
10. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
11. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

$$SR = \frac{\text{cpm of 5 μl of γ-}^{32}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

cpm – value from control (step 7)
nmole – 1.25 nmole (5 μl of 250 μM ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

$$\text{nmole/min/mg} = \frac{\Delta\text{cpm} \times (25/20)}{SR \times E \times T}$$

SR = specific radioactivity of the ATP (cpm/nmole ATP)
Δcpm = cpm of the sample – cpm of the blank (step 3)
25 = total reaction volume
20 = spot volume
T = reaction time (minutes)
E = amount of enzyme (mg)

References

1. Channavajhala, P.L. et al., J. Biol. Chem., **278**, 47089-47097 (2003).
2. Rodríguez, C. et al., Cell Signal., **18**, 1376-1385 (2006).

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