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# Peroxidase (POD), activated from horseradish

**Usi Version: 09** 

Content Version: December 2020

Reagent for the labeling of water-soluble substances carrying primary amino groups with peroxidase.

Cat. No. 11 428 861 001 8 mg

approx. 40 mg lyophilizate 5 labeling reactions

Store the product at +2 to +8°C.

1.	General Information	3
1.1.	Contents	3
1.2.	Storage and Stability	3
	Storage Conditions (Product)	
	Reconstitution	
1.3.	Additional Equipment and Reagent required	
1.4.	Application	4
2.	How to Use this Product	5
2.1.	Before you Begin	5
	General Considerations	
	Purification/fractionation of the conjugate	
	Rebuffering of antibody and conjugate	
	Reaction ratio IgG to PODImmunoglobulin	
	Influence of NaCl and potassium phosphate concentration	
	HPLC chromatography	
	Safety Information	
	Laboratory procedures	
	Waste handling	
	Working Solution	
2.2.	Protocols	
	Conjugation	
2.3.	ParametersInhibition	
	Molecular Weight	
	pH Stability	
	Purity	
	Specific Activity	
	Temperature Stability	9
3.	Supplementary Information	10
3.1.	Conventions	10
3.2.	Changes to previous version	10
3.3.	Trademarks	11
3.4.	License Disclaimer	11
3.5.	Regulatory Disclaimer	11
3.6.	Safety Data Sheet	11
3.7.	Contact and Support	11

# 1. General Information

### 1.1. Contents

Vial / bottle	Label	Function / description	Content
1	Peroxidase (POD), activated	Lyophilized from 0.5 ml.	1 vial,
			8 mg

# 1.2. Storage and Stability

### **Storage Conditions (Product)**

When stored at +2 to +8°C, the product is stable through the expiry date printed on the label.

Vial / bottle	Label	Storage
1	Peroxidase (POD), activated	Store dry at +2 to +8°C.

### Reconstitution

- 1 Reconstitute the lyophilizate in 0.5 ml double-distilled water.
- 2 Store 3 months at +2 to +8°C, or in aliquots shock frozen at −60°C or below and then stored at −15 to −25°C.

A 10 to 20% loss of activity is observed.

Peroxidase concentration = 16 mg/ml.

# 1.3. Additional Equipment and Reagent required

#### For preparation of solutions

- See section, Working Solution for additional information on preparing solutions.
- Double-distilled water
- Na<sub>2</sub>CO<sub>3</sub>, analytical grade
- NaHCO<sub>3</sub>, analytical grade
- NaBH<sub>a</sub>, analytical grade
- Triethanolamine, analytical grade
- HCl, analytical grade
- Glycine, analytical grade
- NaOH, analytical grade
- K<sub>2</sub>HP0<sub>4</sub>, analytical grade
- NaCl, analytical grade
- KH<sub>2</sub>PO<sub>4</sub>, analytical grade

#### 1. General Information

### For conjugation

- Bovine serum albumin
- · Kathon, CG
- Sephadex G25 or PD columns
- Water bath
- Dialysis tube (boiled water treated)
- HPLC TSK-3000 columns
- Chromatography equipment

# 1.4. Application

Peroxidase (POD), activated can be used for:

- Labeling water-soluble substances with reactive and accessible primary amino groups, for example, peptides or proteins with peroxidase for use in analytical methods.
- It is particularly suitable for the coupling of antibodies with peroxidase, as the resulting conjugate is used in immunochemical detection systems, such as ELISA, immunohistochemistry, and immunoblotting procedures.
- 1 The capacity is sufficient for approximately 6 mg IgG. Aliquot the total quantity to be conjugated into 5 portions; each portion will provide 0.5 ml to 1 ml of conjugate from approximately 1.2 mg IgG, a quantity that can be diluted 1:4,000 to 1:10,000 for ELISA applications.

### 2. How to Use this Product

# 2.1. Before you Begin

### **General Considerations**

### Purification/fractionation of the conjugate

When the conjugate is to be used for special applications, such as highly sensitive ELISA procedures or measurements in complex matrices, it can be further purified subsequent to dialysis and prior to stabilization with bovine serum albumin, by gel permeation chromatography, and the fractions tested for their suitability for the planned application.

### Rebuffering of antibody and conjugate

See section, Protocols for additional information about Steps 4 and 5.

The antibody can be rebuffered into Solution 2 either using Sephadex G25 or PD-columns or other suitable material. The conjugate can then be rebuffered subsequent to Step 4 using column chromatography, instead of the described dialysis (Step 5).

### Reaction ratio IgG to POD

*i* See section, **Protocols** for additional information about Step 1.

The conjugation of IgG and fab-fragments is optimized for the Step 1 ratio. Other stoichiometric proportions can be considered for special applications, but the protein concentrations indicated in the test procedure should be held constant and a molecular sieve fractionation should be carried out to separate residual IgG or POD.

### **Immunoglobulin**

The procedure is optimized for coupling immunoglobulin from rabbit; it can also be conjugated with IgG from sheep and goat. When Fab- or  $F(ab')_2$ -fragments of these species or immunoglobulin G from mouse are to be used, a reaction time of 3 hours at +15 to +25°C should be used.

### Influence of NaCl and potassium phosphate concentration

(Coupling in PBS with subsequent pH adjustment)

NaCl concentrations of 50 to 400 mM and simultaneous phosphate concentrations of 10 to 20 mM have a marginal effect on the reaction.

- For potassium phosphate, use a concentration in the range of 30 to 100 mM.
- The NaCl concentration should not exceed 150 mM.

### **HPLC** chromatography

For high reproducibility, reactions should be carried out under HPLC TSK-3000 control. Figures 1 to 3 show the TSK-3000 profiles of the starting material, immunoglobulin G from rabbit (Figure 1), POD (Figure 2), and the final product (Figure 3). The IgG is completely bound in the conjugate, corresponding to the TSK-3000 separation profile of Figure 3.

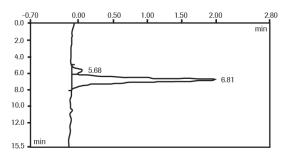


Fig. 1: HPLC, TSK-3000; IgG from rabbit.

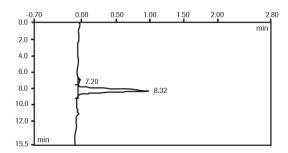


Fig. 2: HPLC, TSK-3000; Peroxidase.

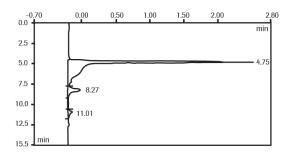


Fig. 3: HPLC, TSK-3000; Conjugate of IgG and POD.

# **Safety Information**

### **Laboratory procedures**

- Handle all samples as if potentially infectious, using safe laboratory procedures. As the sensitivity and titer of
  potential pathogens in the sample material varies, the operator must optimize pathogen inactivation by the Lysis /
  Binding Buffer or take appropriate measures, according to local safety regulations.
- Do not eat, drink or smoke in the laboratory work area.
- Do not pipette by mouth.
- Wear protective disposable gloves, laboratory coats and eye protection, when handling samples and kit reagents.
- Wash hands thoroughly after handling samples and reagents.

#### **Waste handling**

- Discard unused reagents and waste in accordance with country, federal, state, and local regulations.
- Safety Data Sheets (SDS) are available online on dialog.roche.com, or upon request from the local Roche office.

# **Working Solution**

*1* To avoid confusion, mark each solution with the appropriate number.

Solution No.	Solution	Preparation/Composition [+15 to +25°C]	Storage and Stability
1	1 M Sodium carbonate/ hydrogencarbonate, pH 9.4	<ul> <li>1 M Na<sub>2</sub>CO<sub>3</sub>: Dissolve 10.6 g Na<sub>2</sub>CO<sub>3</sub> in 80 ml double-distilled water and dilute up to 100 ml.</li> <li>1 M NaHCO<sub>3</sub>: Dissolve 8.4 g NaHCO<sub>3</sub> in 80 ml double-distilled water and dilute up to 100 ml.</li> <li>Adjust the pH of the NaHCO<sub>3</sub> solution to 9.4 by adding Na<sub>2</sub>CO<sub>3</sub> solution.</li> </ul>	Store 1 week at +2 to +8°C.
2	100 mM Sodium carbonate/- hydrogencarbonate, pH 9.8	Dilute 10 ml Solution 1 to 100 ml with double-distilled water.	Store 1 week at +2 to +8°C.
3	200 mM Sodium borohydride	Dissolve 8 mg NaBH <sub>4</sub> in 1 ml cold double-distilled water.	Prepare immediately before use. Keep cold on ice.
4	2 M Triethanolamine, pH 8.0	Dilute 2.66 ml triethanolamine with 3 ml double-distilled water, adjust the pH to 8.0 with 25% HCl, and dilute up to 10 ml with double-distilled water.	Store 1 week at +2 to +8°C.
5	1 M Glycine, pH 7.0	Dissolve 0.75 g glycine in approximately 6 ml double-distilled water, adjust to pH 7.0 with 0.1 M NaOH, and dilute up to 10 ml with double-distilled water.	Store 1 week at +2 to +8°C.
6	PBS (phosphate buffered saline); glycine; pH 7.4	<ul> <li>10 mM Potassium phosphate, 200 mM NaCl,</li> <li>10 mM glycine, pH 7.5</li> <li>Solution A (K<sub>2</sub>HPO<sub>4</sub>): Dissolve 4.56 g K<sub>2</sub>HPO<sub>4</sub> × 3 H<sub>2</sub>O, 23.4 g NaCl, and 1.5 g glycine in approximately 1,500 ml double-distilled water and dilute up to 2,000 ml with double-distilled water.</li> <li>Solution B (KH<sub>2</sub>PO<sub>4</sub>): Dissolve 2.72 g KH<sub>2</sub>PO<sub>4</sub>, 23.4 g NaCl, and 1.5 g glycine in approximately 1,500 ml double-distilled water and dilute up to 2,000 ml with double-distilled water.</li> <li>PBS: While controlling pH, add sufficient Solution B to Solution A until the pH is 7.4.</li> </ul>	Store 1 week at +2 to +8°C.
7	Antibody	<ul> <li>0.3 ml is required for each labeling reaction.</li> <li>The IgG concentration of the solution to be used is c = 4 mg/ml (3.8 to 4.2 mg/ml).</li> <li>This value is critical for the coupling and should be checked photometrically for every test and adjusted if necessary: A<sub>280nm</sub>, 1 cm, 1 mg/ml = 1.40.</li> <li>Do not use preservatives, such as sodium azide, and stabilizers, such as albumin.</li> <li>Immunoglobulin, salt-free, lyophilized:</li> <li>Weigh 1.6 mg into a suitable vessel and dissolve in 0.4 ml Solution 2.</li> <li>Check the concentration and pH and correct if necessary.</li> <li>Immunoglobulin in buffer:</li> <li>PBS buffer without additional proteins or preservatives: adjust the pH to 9.8 with Solution 1 and if necessary, dilute with Solution 2 to obtain an IgG concentration of 4 mg/ml.</li> </ul>	Prepare immediately before use.
		<ul> <li>Buffer with organic salts: Dialyze immunoglobulin into Solution 2 and adjust the concentration to 4 mg/ml with Solution 2.</li> </ul>	

### 2.2. Protocols

### Conjugation

The Conjugation protocol has been specially developed for the coupling of peroxidase to immunoglobulin G (lgG). It can be equally successfully used for lgG Fab- and F(ab')2-fragments from rabbit, mouse, sheep and goat, see section, **General Considerations**. If other proteins are to be conjugated, start with this procedure and check the results with gel chromatography on HPLC, TSK-3000. If necessary, the procedure can then be adapted to individual requirements by altering the stoichiometry and the concentration of reactants used for incubation. The test procedure describes the conjugation using 1/5 of the total quantity of reagent, sufficient to label 1.2 mg of lgG.

- 3 See section, Working Solution for information on preparing solutions.
- 1 Pipette exactly 0.3 ml Antibody solution into a suitable 1 to 2 ml vessel, add 0.1 ml activated Peroxidase, and mix well.
  - Reaction ratio: 1 M IgG:5 M POD.
  - Molecular weight of POD is 44,000 Da.
- 2 Incubate for 2 hours at +15 to +25°C in a water bath or for 16 hours overnight at +2 to +8°C.
- 3 Stop the reaction by adding 40 µl Solution 4 (Triethanolamine solution) to the incubation solution and mix.
  - Pipette 50 μl Solution 3 (Sodium borohydride solution) to the mixture, mix again, and incubate for 30 minutes at +2 to +8°C.
  - Add another 25 µl of Solution 4 (Triethanolamine solution) and incubate again for 2 hours at +2 to +8°C.
- A Stabilize the conjugate by pipetting 10 μl Solution 5 (1 M Glycine solution) into the incubation mixture; briefly mix.
- 5 Transfer conjugate to storage buffer by placing the incubation solution in a dialysis tube (boiled-water treated) and dialyze extensively, for example, overnight with 4 changes of 500 ml Solution 6 (PBS, glycine).
- 6 Stabilize the product for storage by placing the conjugate from the dialysis tube into a suitable vessel, add 10 mg/ml bovine serum albumin, and 1 mg/ml Kathon CG; mix gently to dissolve.
- 7 The conjugate can be stored for at least 2 months at +2 to +8°C.
  - For long-term storage without loss of activity, aliquot and shock freeze in liquid nitrogen and store at −60°C or below.
  - Never add sodium azide as a preservative to the Sodium borohydride solution, as it inhibits the peroxidase activity.
- The reaction proceeds in such a way that practically no (<5%) immunoglobulin is left in the dialysate, see section, **General Considerations, Figure 3**.
- The conjugate does not need to be purified for normal immunoassay procedures, as residual amounts of POD do not normally interfere with such tests.

### 2.3. Parameters

### Inhibition

Never add sodium azide as a preservative as it inhibits peroxidase activity.

## **Molecular Weight**

44,000 Da

### pH Stability

The pH should not be allowed to fall below 9.8. To ensure reproducible results, keep pH constant.

10.8. The maximum usable pH is 10.8.

### **Purity**

Purity number (A $_{\rm 405~nm}/A_{\rm 275~nm}$ ): 3.0 to 3.5 Isoenzyme distribution: >90% homogeneous isoenzyme C.

# **Specific Activity**

≥550 U/mg protein at +25°C with ABTS Substrate,  $H_2O_2$ , pH 5.0.

# **Temperature Stability**

### **Reaction temperature and duration**

The procedure is developed for a reaction temperature of +25°C, but is reaction time tolerant.

- At +15 to +25°C, the reaction time should be 2 hours, but this can be extended to 3 hours without influencing the results.
- Reactions can also be carried out at +2 to +8°C for 18 to 24 hours.

# 3. Supplementary Information

# 3.1. Conventions

To make information consistent and easier to read, the following text conventions and symbols are used in this document to highlight important information:

Text convention and symbols			
1 Information Note: Additional information about the current topic or procedure.			
⚠ Important Note: Information critical to the success of the current procedure or use of the product.			
1 2 3 etc.	Stages in a process that usually occur in the order listed.		
1 2 3 etc.	Steps in a procedure that must be performed in the order listed.		
* (Asterisk)	The Asterisk denotes a product available from Roche Diagnostics.		

# 3.2. Changes to previous version

Layout changes.

Editorial changes.

Update to include new safety Information to ensure handling according controlled conditions.

### 3.3. Trademarks

ABTS is a trademark of Roche.

All other product names and trademarks are the property of their respective owners.

### 3.4. License Disclaimer

For patent license limitations for individual products please refer to: **List of biochemical reagent products**.

# 3.5. Regulatory Disclaimer

For life science research only. Not for use in diagnostic procedures.

# 3.6. Safety Data Sheet

Please follow the instructions in the Safety Data Sheet (SDS).

# 3.7. Contact and Support

To ask questions, solve problems, suggest enhancements or report new applications, please visit our **Online Technical Support Site**.

To call, write, fax, or email us, visit **sigma-aldrich.com**, and select your home country. Country-specific contact information will be displayed.