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Product Information

EPHA6, active, GST tagged, human PRECISIO® Kinase recombinant, expressed in *Sf*9 cells

Catalog Number **SRP5028** Storage Temperature –70 °C

Synonyms: EPA6, FLJ35246, PRO57066, DKFZp434C1418

Product Description

EPHA6 is a member of the ephrin receptor subfamily of protein-tyrosine kinases, which have been implicated in axon guidance, neuron-target interactions, regional compartmentalization, and synaptic functions in nervous systems. EPHA6 is highly expressed in the brain and testis. Reduction in EPHA6 has been detected in hypospadias, a common defect affecting the growth and closure of the external genitalia. Genetic inhibition of EPHA6 in mice produces behavioral deficits specifically in tests of learning and memory. Mice deficient in EPHA6 show reduced memory of the consequences of the training context.

Recombinant human EPHB4 (561-end) was expressed by baculovirus in *Sf*9 insect cells using an N-terminal GST tag. The gene accession number is NM_001080448. Recombinant protein stored in 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM glutathione, 0.1 mM EDTA, 0.25 mM DTT, 0.1 mM PMSF, and 25% glycerol

Molecular mass: ~73 kDa

Purity: 70-95% (SDS-PAGE, see Figure 1)

Specific Activity: 7.6–10.4 nmole/min/mg (see Figure 2)

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at -70 °C is recommended. After opening, aliquot into smaller quantities and store at -70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Typical Lot 70–95% (densitometry)

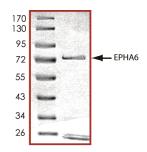
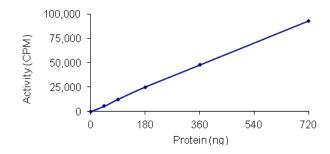


Figure 2.
Specific Activity of Typical Lot 7.6–10.4 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25mM MOPS, pH 7. 2, 12.5 mM glycerol 2-phosphate, 20 mM MgC1₂, 12.5mM MnC1₂, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 $ng/\mu l$ BSA.

Kinase Solution – Dilute the active EPHA6 (0.1 μ g/ μ l) with Kinase Dilution Buffer to the desired concentration. Note: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active EPHA6 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μ l aliquots at –20 °C.

 γ -³³P-ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of γ -³³P-ATP (1 mCi/100 μl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Dissolve the synthetic peptide substrate in distilled water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ³³P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active EPHA6, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ -33P-ATP Assay Cocktail may be thawed at room temperature.
- In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl:

10 μl of Kinase Solution

5 μ l of Substrate Solution

5 μ l of cold water (4 °C)

- 3. Set up a blank control as outlined in step 2, substituting 5 μl of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ - 33 P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- 5. After the 15 minute incubation, stop the reaction by spotting 20 μ l of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ - 33 P-ATP counts introduced into the reaction. Spot 5 μ l of the γ - 33 P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR =
$$\frac{\text{cpm of 5} \mu \text{l of } \gamma^{-33}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

cpm – value from control (step 7) nmole – 1.25 nmole (5 μl of 250 μM ATP Assav Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\Delta$$
cpm × (25/20)
SR × E × T

SR = specific radioactivity of the ATP (cpm/nmole ATP) Δ cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

20 = spot volume

T = reaction time (minutes)

E = amount of enzyme (mg)

References

- Shaut, C.A. et al., HOXA13 directly regulates EphA6 and EphA7 expression in the genital tubercle vascular endothelia. Dev. Dyn., 236(4), 951-960 (2007).
- Savelieva, K.V. et al., Learning and memory impairment in Eph receptor A6 knockout mice. Neurosci. Lett., 438(2), 205-209 (2008).

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