

Technical Bulletin

Fumarase Assay Kit

Catalogue number MAK521

Product Description

Fumarase (or Fumarate Hydratase) is an enzyme that catalyzes the reversible hydration/dehydration reaction of fumarate to malate. Fumarase exists in two isoforms: a cytosolic and mitochondrial form. In the citric acid cycle, it facilitates a transition step in the production of energy in the form of NADH.

This non-radioactive, colorimetric fumarase assay is based on the reduction of the tetrazolium salt MTT in a NADH-coupled enzymatic reaction to a reduced form of MTT which exhibits an absorption maximum at 565 nm. The increase in absorbance at 565 nm is proportional to the enzyme activity.

The linear detection range of the Fumarase Assay Kit is 0.4 to 70 U/L fumarase. The kit is suitable for determination of fumarase in biological samples such as plasma, serum, erythrocytes, tissue and culture media.

Components

The kit is sufficient for 100 colorimetric assays in 96-well plates.

• Assay Buffer	10 mL
Catalogue Number MAK521A	
• Enzyme A	120 μ L
Catalogue Number MAK521B	
• Enzyme B	120 μ L
Catalogue Number MAK521C	
• NAD/MTT	1 mL
Catalogue Number MAK521D	
• Substrate	600 μ L
Catalogue Number MAK521E	
• Calibrator	1.5 mL
Catalogue Number MAK521F	

Equipment Required but Not Provided

- Pipetting devices and accessories (for example, multichannel pipettor)
- Spectrophotometric multiwell plate reader.
- Clear flat-bottom 96-well plates (for example, Corning Costar). Cell culture or tissue culture treated plates are not recommended.
- 1.5 mL centrifuge tubes
- Refrigerated microcentrifuge capable of RCF \geq 14,000 \times g.

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The kit is shipped on wet ice. Store all components at -20 °C.

Preparation Instructions

Briefly centrifuge small vials prior to opening. Equilibrate reagents to desired reaction temperature.

Enzyme A and B: Keep enzymes on ice during experiment.

Note: This assay is based on a kinetic reaction. To ensure identical incubation time, addition of Working Reagent to samples should be quick and mixing should be brief but thorough. Use of a multi-channel pipettor is recommended. Assays can be executed at any desired temperature (for example, 25 °C or 37 °C).

Procedure

All samples and standards should be run in duplicates.

Sample Preparation

Serum and plasma samples can be assayed directly.

Tissue Samples:

1. Rinse tissue in phosphate buffered saline (pH 7.4) to remove blood, prior to dissection.
2. Homogenize 50 mg of tissue in ~200 μ L cold 50 mM potassium phosphate buffer, pH 7.5.
3. Centrifuge at 14,000 \times g for 10 minutes at 4 °C. Remove supernatant for assay.

Cell Samples:

1. Collect cells by centrifugation at 2,000 \times g for 5 minutes at 4 °C.
2. For adherent cells, do not harvest cells using proteolytic enzymes; rather use a rubber policeman.
3. Homogenize or sonicate cells in an appropriate volume of cold buffer containing 50 mM potassium phosphate (pH 7.5).
4. Centrifuge at 14,000 \times g for 10 minutes at 4 °C. Remove supernatant for assay.

Note: All samples can be stored at -20 to -80 °C for at least one month.

Assay

1. Transfer 100 μ L H₂O (OD_{H2O}) and 100 μ L Calibrator (OD_{CAL}) solution into separate wells of a clear flat bottom 96-well plate.
2. Transfer 20 μ L H₂O into one well, this will be the blank. Transfer 20 μ L of each sample into separate wells.

Working Reagents

1. Mix enough reagents for the number of assays to be performed. Prepare Working Reagents according to Table 1. For each well, prepare 89 μ L of Working Reagent.

Table 1.

Preparation of Working Reagents

Reagent	Working Reagent
Assay Buffer	74 μ L
NAD/MTT	8 μ L
Substrate	5 μ L
Enzyme A	1 μ L
Enzyme B	1 μ L

2. Add 80 μ L of Working Reagent to blank and sample wells and tap gently to mix.

Measurement

1. Read the optical density at 565 nm at 10 minutes and at 40 minutes on a plate reader.

Results

1. Subtract the Optical Density values recorded at 10 mins from 40 min for each sample to compute the ΔOD_S values.
2. Repeat the same for the blank to compute ΔOD_B values.
3. Fumarase activity can be calculated using the below equation:

Fumarase Activity (U/L) =

$$\frac{\Delta OD_S - \Delta OD_B}{\epsilon_{mtt} \cdot l} \times \frac{\text{Reaction Vol } (\mu\text{L})}{t(\text{min}) \cdot \text{Sample Vol } (\mu\text{L})} \times DF$$
$$= \frac{273}{t \text{ (min)}} \times \frac{\Delta OD_S - \Delta OD_B}{OD_{Cal} - \Delta OD_{H_2O}} \times DF$$

Where:

ϵ_{mtt} = Molar absorption coefficient of reduced MTT.

l = Path length which is calculated from the calibrator.

OD_{CAL} = Optical Density of Calibrator.

OD_B = Optical Density of Blank.

OD_S = Optical Density of Samples.

t = Difference in time between readings.

DF = Dilution factor if the sample needed to be diluted.

Reaction Vol = 100 μL .

Sample Vol = 20 μL .

Unit definition: 1 Unit (U) of Fumarase will catalyze the conversion of 1 μmole of L-fumarate to L-malate per minute at pH 7.8.

Note: If sample fumarase activity exceeds 70 U/L, dilute samples in water and repeat the assay. For samples with fumarase activity < 1 U/L, the incubation time can be extended to 2 hours.

Figure 1.

Typical raw kinetic data for fumarase.

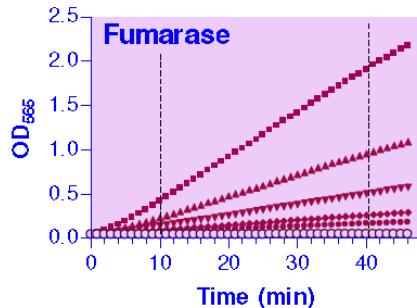
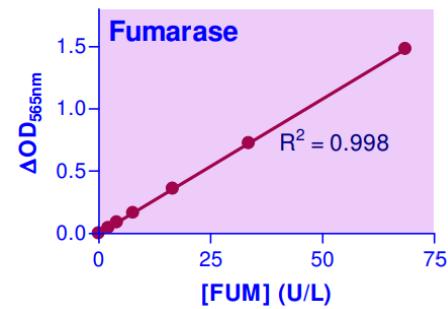


Figure 2.

Representative data of fumarase activity (30 minutes, 37 °C).



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