

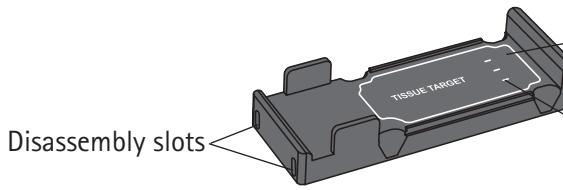
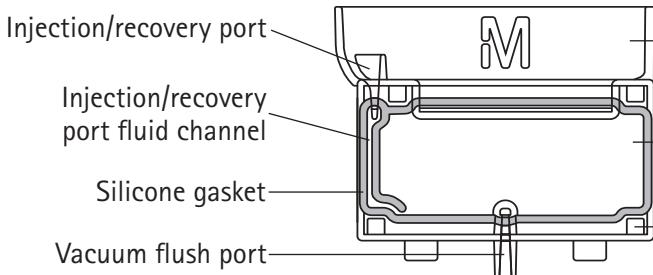
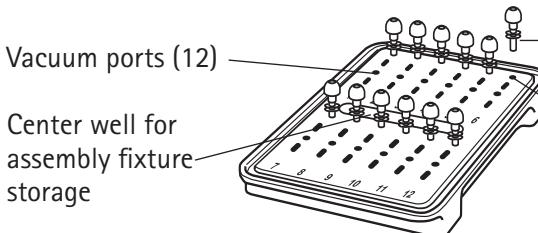
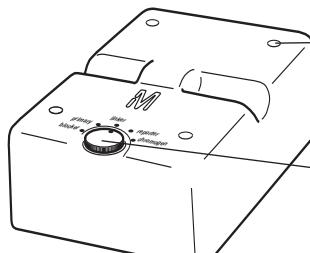
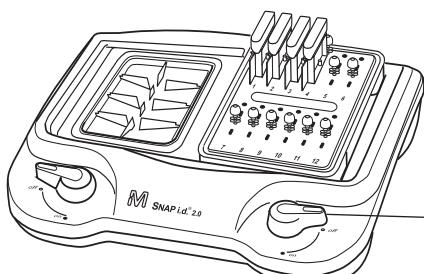


Quick Start Guide

SNAP i.d.[®] 2.0 IHC (Immunohistochemistry) System

For research use only

Before using the SNAP i.d.[®] 2.0 IHC System, please read the full-length user guide available at www.millipore.com/ihc.

Component	Features
Assembly Fixture Accepts 25 x 75 x 1 mm slide (slides not provided)	 <p>Tissue section target (guides tissue section size and placement) Disassembly slots Volume markings (for estimating solution volume required; each line represents ~150 µL)</p>
Slide Holder Injection/recovery port accepts 1 mL pipette tip	 <p>Injection/recovery port Injection/recovery port fluid channel Silicone gasket Vacuum flush port Wash reservoir (6 mL maximum volume) Tissue section chamber (600 µL maximum volume) Slide compression clips (4)</p>
IHC Frame Holds up to 12 slide holders	 <p>Vacuum ports (12) Center well for assembly fixture storage Port plugs (to maintain vacuum when port is not being used) Port plug storage holes</p>
Incubation Cover	 <p>Stacking recesses (to enable IHC frame stacking off-line) Task tracker (for keeping track of steps)</p>
SNAP i.d.[®] 2.0 IHC System Accommodates one or two IHC frames Includes tubing assembly kit	 <p>Vacuum control knob</p>

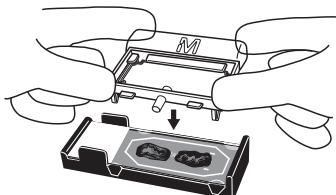
Setup

General Guidelines

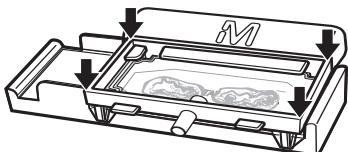
- Do not use the SNAP i.d.® 2.0 IHC system for deparaffinization, rehydration, dehydration, or mounting steps.
- Proper slide preparation is critical to successful use. Before placing tissue on slide, make sure that it will not extend beyond the target area of the assembly fixture.
- **IMPORTANT!** Do not touch the gasket on the slide holder during assembly.

Slide Holder Setup

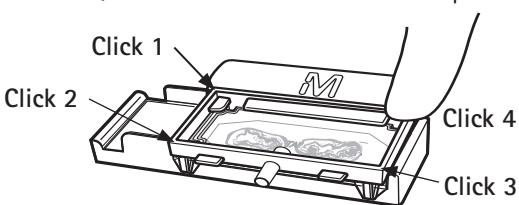
1. Place assembly fixture on a hard, flat surface.
2. Position slide in assembly fixture, with the tissue section facing up. Verify that the tissue section does not extend beyond the target area.
3. Hold the slide holder by the sides, clip side down, and align it with the assembly fixture containing the slide. **Do not touch the gasket.**



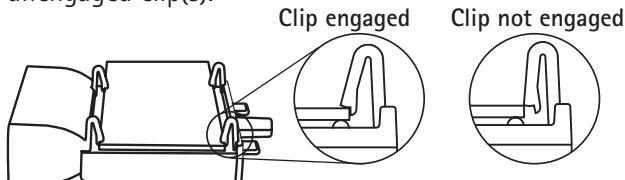
4. Press the slide holder down firmly over the slide until the four compression clips slip over the slide edges with an audible click.



5. To ensure that all clips are fully engaged, firmly press down on the top edge of each slide holder corner, one at a time, and listen for a click on each press.



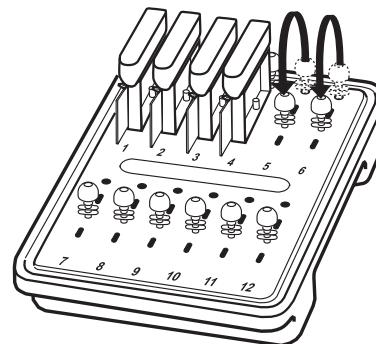
6. Remove the slide holder assembly from the fixture and closely inspect each clip to confirm that it is in the proper position over the slide. If any clip is not engaging the slide as shown below, place the assembly back into the fixture and repeat step 5 on the unengaged clip(s).



IHC Frame Setup

The IHC frame can be set up either on or off the SNAP i.d.® 2.0 system.

1. Place assembled slide holders into any vacuum ports on the frame, with slide labels oriented towards the port numbers.
2. Press blue port plugs **lightly** into all unused ports (excessive force is not required). Unused plugs can be left in the port plug storage holes.

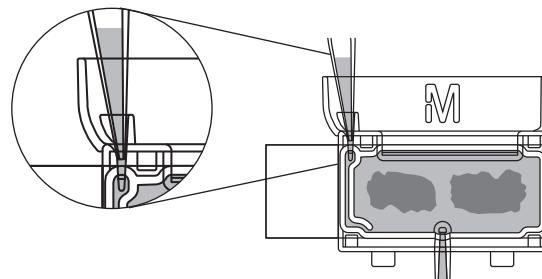


3. If the frame has been loaded off the SNAP i.d.® 2.0 system, place it in either position on the system before continuing.

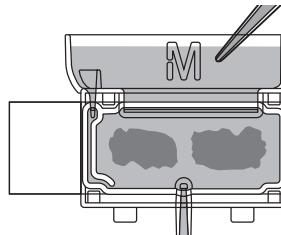
Staining Protocol

Tips for adding solutions to slide holder

- Small volumes (up to 600 μ L) should be dispensed with a 1 mL pipette through the injection/recovery port. Gently push the pipette tip into the port to create a seal. Dispense slowly to avoid bubble formation in the tissue section chamber.



- Larger volumes (up to 6 mL) can be loaded into the wash reservoir.



- Make sure that the fluid completely covers the tissue sections.

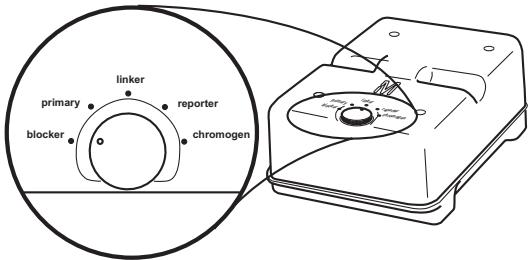
Washing (optional)

1. If washing is required before blocking, fill the wash reservoir of the slide holder with up to 6 mL of desired wash solution.
2. To flush the wash solution out of the tissue chamber, turn on the SNAP i.d.® 2.0 system vacuum.

NOTE: Residual fluid at the bottom of the tissue section chamber will not impact subsequent steps.

Blocking

1. Load blocking reagent through the injection/recovery port or the wash reservoir.
2. If incubation is required, place the cover on the IHC frame and set the task tracker to "blocker".



3. To flush the blocking solution from the slide holder, remove the cover and activate the vacuum.
4. If wash steps are needed, fill the wash reservoir with wash solution, then flush by applying vacuum. If necessary, repeat wash step two more times.

Primary Antibody

1. Inject up to 600 µL of primary antibody through the injection/recovery port, dispensing slowly to avoid bubbles, and ensuring complete coverage of the tissue section.
2. Cover the IHC frame and set the task tracker to "primary". When incubation is complete, remove the cover.
3. Recover primary antibody with a pipette via the injection/recovery port or flush to waste by applying vacuum.
4. Fill the wash reservoir with wash solution, then flush by applying vacuum. Repeat wash step two more times if required.

Linker (secondary antibody)

1. Inject up to 600 µL of linker through the injection/recovery port, dispensing slowly to avoid bubbles, and ensuring complete coverage of the tissue section.
2. Cover the IHC frame and set the task tracker to "linker". When incubation is complete, remove the cover.
3. Recover linker with a pipette via the injection/recovery port or flush to waste by applying vacuum.
4. Fill the wash reservoir with wash solution, then flush by applying vacuum. Repeat wash step two more times if required.

Reporter (tertiary, if required)

1. Inject up to 600 µL of reporter through the injection/recovery port, dispensing slowly to avoid bubbles, and ensuring complete coverage of the tissue section.
2. Cover the IHC frame and set the task tracker to "reporter". When incubation is complete, remove the cover.
3. Recover reporter with a pipette via the injection/recovery port or flush to waste by applying vacuum.
4. Fill the wash reservoir with wash solution, then flush by applying vacuum. Repeat wash step two more times if required.

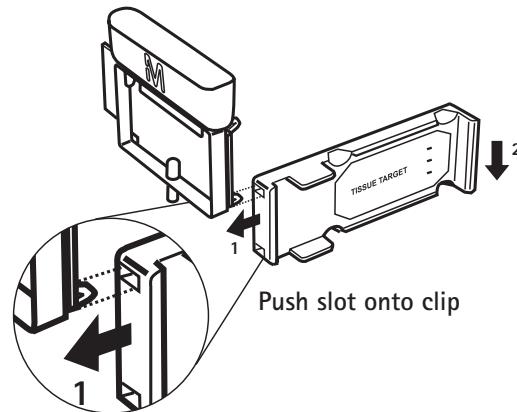
Chromogen

NOTE: Some stains such as hematoxylin and eosin may permanently stain the SNAP i.d.® 2.0 system.

1. Inject up to 600 µL of desired chromogen through the injection/recovery port, dispensing slowly to avoid bubbles, and ensuring complete coverage of the tissue section.
2. Cover the IHC frame and set the task tracker to "chromogen". When incubation is complete, remove the cover.
3. Recover chromogen with a pipette via the injection/recovery port or flush to waste by applying vacuum.
4. Fill the wash reservoir with wash solution, then flush by applying vacuum. Repeat wash two more times if required.

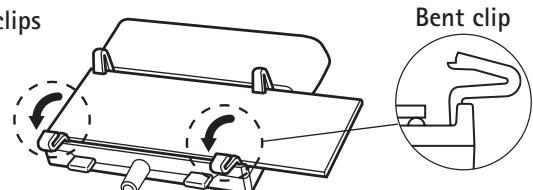
Slide Removal

1. After staining/counter-staining is complete, a final wash is recommended before removing the slide from the slide holder.
2. To remove the slide, push the assembly fixture disassembly slot over one of the bottom clips (1).



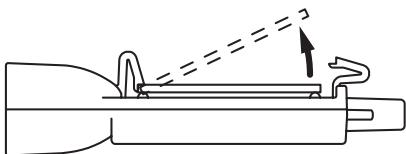
3. Bend the clip 90 degrees away from the slide (2). Repeat with the other bottom clip.

Bend bottom clips with assembly fixture



Slide Removal, continued

4. Carefully lift the slide up and out of the holder.



5. Discard the used slide holder. If required, proceed with dehydration and cover slip mounting protocols.

NOTE: Do not use the SNAP i.d.® 2.0 IHC system for dehydration or mounting steps.



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Product Ordering Information

This section lists the catalogue numbers for SNAP i.d.® 2.0 IHC System components. See the Technical Assistance section for contact information. You can purchase these products on-line at www.millipore.com/products.

Description	Cat. No.	Qty/Pk
SNAP i.d.® 2.0 Base	SNAP2BASE	1
SNAP i.d.® 2.0 IHC Frame	SNAP2FRIHC	1
SNAP i.d.® 2.0 IHC Slide Holder	SNAP2SH	24

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Warranty

The applicable warranty for the products listed in this publication may be found at www.millipore.com/terms ("Conditions of Sale").