

7-FLUOROBENZO-2-OXA-1,3-DIAZOLE-4-SULFONIC ACID AMMONIUM

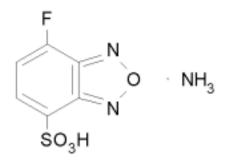
Product Number **F4383** Storage Temperature RT

CAS #: 84806-27-9

Synonyms: SBD-F¹, 4-Fluoro-7-Sulfobenzofurazan,

Ammonium salt²,

Product Description



Appearance: White to light yellow powder³ Molecular Formula: C₆H₃FN₂O₄S·NH₃

Molecular Weight: 235.2

Method of preparation of SBD-F was reported.1

SBD-F is a sensitive and specific fluorescent labeling reagent for low molecular weight compounds and for macromolecules containing thiol groups. 1,4-10 SBD-F reacts with sulfhydryl groups (disulfides do not react with SBD-F and must first be reduced to thiols, for example using tributylphosphine) to produce highly fluorescent compounds. SBD-F does not exhibit any fluorescence of its own and no fluorescent by-products are formed in the reaction with SBD-F. 1,8 Suggested optimal reaction conditions for formation of fluorescent derivatives are SBD-F (0.5 mM), 60°C, and pH 9.5 (0.1 M borate buffer containing 1 mM EDTA disodium) for one hour. Additional procedures have been reported. At pH 9.5, the inclusion of 1 mM EDTA in the buffer is important to prevent metal-catalyzed oxidation of thiols. 10 The rate of reaction of thiols with SBD-F gradually increases with increasing pH. 10 High fluorescence intensities are observed at pH 2-12 except for SBD-cysteine and SBD- (thiol containing amino acids) which require an acidic (pH 2) medium. 10

For the SBD-F derivative:

Excitation wavelength: 380-385nm^{1,4,5} Emission wavelength: 515nm^{1,4,5}, 510nm⁶

ProductInformation

SBD-F is suitable for the HPLC determination of biological thiols at the picomole level. The detection limit of thiol compounds, glutathione, coenzyme A, cysteamine, homocysteine, N-acetylcysteine, cysteine and D-3-mercaptomethylpropanoyl-L-proline (captopril) were 100, 120, 160, 330, 390, 3600, 150 pmol/ml, respectively¹. The amino acids, proline and alanine (contain no -SH groups), did not give any fluorescence with SBD-F. Some thiols (cysteine, glutathione and captopril) have also been determined by reaction with SBD-F then separated and quantified by HPLC. 7,10 The determination of SBD-thiols (both reduced and oxidized) in plasma by HPLC has been reported.4,7 SBD-F has been used in HPLC methods for measuring total plasma and serum homocysteine levels. 4,5,8-10 SBD-F has been used in the determination of metallothionein in a tandem column HPLC method with an isocratic solvent system.⁶ SBD-F has been used for determining the position of disulfide linkages of cysteine residues in proteins¹⁰⁻¹² and for the detection of cystine containing peptides.¹³ Positions of disulfide bonds in Yam acidic class IL (class IV) chitinase were determined by digestion of the chitinase. The resulting disulfide bonds containing peptides were separated by reversed-phase HPLC and detected using SBD-F¹¹: SBD-F was employed in the determination of disulfide bridges in Factor Va heavy chain. 12

Storage/Stability

The product is stable for 3 years when stored at room temperature and protected from light.³

Preparation Instructions

SBD-F has been solubilized in 1 M ammonium hydroxide at 50 mg/ml yielding a clear yellow to yellow-green solution.³ The product is also freely soluble in deionized water. A 10 mg/ml solution has been prepared.¹ SBD-F solutions in water were reported to be stable for more than one week at 2-8°C.¹ SBD-F derivatives were stable at pH 9.5¹ for at least one week at room temperature when stored in the dark ^{1,5,6,9} and 0°¹⁰ (derivatized samples may turn yellow because of the light sensitivity of SBD-F. Use of the derivatives did not seem to affect the assay for homocysteine determination in plasma).⁹ SBD-F homocysteine complexes are more stable than o-phthaldialdehyde (OPA)-homocysteine complexes when both were stored in the dark .⁵

References

- 1. Imai, K. et al., Anal. Biochem., 128, 471, (1983).
- 2. Supplier information
- 3. Sigma quality control
- Araki, A. and Sako, Y., J. Chromatogr., 422, 43, (1987).
- 5. Fermo, I. et al., J. Chromotagr. B, 719, 31, (1998).
- 6. Miyairi, S. et al., Anal. Biochem., 258, 168, (1998).
- 7. Toyo'oka, T. and Imai, K., J. Chromatogr., 282, 495, (1983).

- 8. Ubbink, J.B. et al., J. Chromatogr., 565, 441, (1991).
- 9. Vester, B. and Rasmussen, K., Eur. J. Clin. Chem. Clin. Biochem., 29, 549, (1991).
- 10. Imai, K. and Toyo'oka, T., Methods Enzymol., 143, 67, (1987).
- 11. Araki, T. et al., Arch. Biochem. Biophys., 335, 118, (1996).
- 12. Xue, J. et al., Biochem., 33, 13109, (1994).
- 13. Sueoyoshi, T. et al., J. Biochem., 97, 1811, (1985).

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