

Product Information

Neuropeptide Y EIA Kit

for serum, plasma, culture supernatant, and cell lysates

Catalog Number **RAB0387**

Storage Temperature –20 °C

TECHNICAL BULLETIN

Product Description

The Neuropeptide Y (NPY) Enzyme Immunoassay (EIA) Kit is an *in vitro* quantitative assay for detecting Neuropeptide Y peptide based on the principle of competitive enzyme immunoassay. The microplate in the kit is pre-coated with anti-rabbit secondary antibody. After a blocking step and incubation of the plate with anti-Neuropeptide Y antibody, both biotinylated Neuropeptide Y peptide, and peptide standard or targeted peptide in samples interacts competitively with the Neuropeptide Y antibody. Uncompeted (bound) biotinylated Neuropeptide Y peptide then interacts with Streptavidin-horseradish peroxidase (SA-HRP), which catalyzes a color development reaction. The intensity of colorimetric signal is directly proportional to the amount of biotinylated peptide-SA-HRP complex and inversely proportional to the amount of Neuropeptide Y peptide in the standard or samples. This is due to the competitive binding to Neuropeptide Y antibody between biotinylated Neuropeptide Y peptide and peptides in standard or samples. A standard curve of known concentration of Neuropeptide Y peptide can be established and the concentration of Neuropeptide Y peptide in the samples can be calculated accordingly.

Components

1. 96-well plate coated with secondary antibody (Item A) - RAB0387A: 96 wells (12 strips × 8 wells) coated with secondary antibody.
2. 20x Wash Buffer (Item B) - RABWASH3: 25 mL.
3. EIA Neuropeptide-Y Peptide standard, Lyophilized (Item C) - RAB0387C: 2 vials.
4. Anti-Neuropeptide Y Detection Antibody, Lyophilized (Item N) - RAB0387F: 2 vials.
5. EIA Neuropeptide-Y 1x Assay Diluent E (Item R) – RAB0385R: 2 vials, 25 mL/vial. Diluent for both standards and samples including serum or plasma, cell culture media, or other sample types.

6. Biotinylated Neuropeptide Y, Lyophilized (Item F) - RAB0387G: 2 vials.
7. HRP-streptavidin (Item G) - RABHRP3: 600 µL of 200x concentrated HRP-conjugated Streptavidin.
8. Neuropeptide Y Positive Control Sample, Lyophilized (Item M) - RAB0387K: 1 vial.
9. TMB Substrate solution (Item H) - RABTMB2: 12 mL of 3,3',5,5'- tetramethylbenzidine (TMB) in buffered solution.
10. Stop Solution (Item I) – RABSTOP3: 8 mL of 0.2 M sulfuric acid.

Reagents and Equipment Required but Not Provided.

1. Microplate reader capable of measuring absorbance at 450nm.
2. Precision pipettes to deliver 2 µL to 1 mL volumes.
3. Adjustable 1-25 mL pipettes for reagent preparation.
4. 100 mL and 1 liter graduated cylinders.
5. Absorbent paper.
6. Distilled or deionized water.
7. SigmaPlot software (or other software which can perform four-parameter logistic regression models).
8. Tubes to prepare standard or sample dilutions.
9. Orbital shaker.
10. Aluminum foil.

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices

Preparation Instructions

For sample and positive control dilutions, refer to steps 5, 6, 7, and 9 of Preparation Instructions.

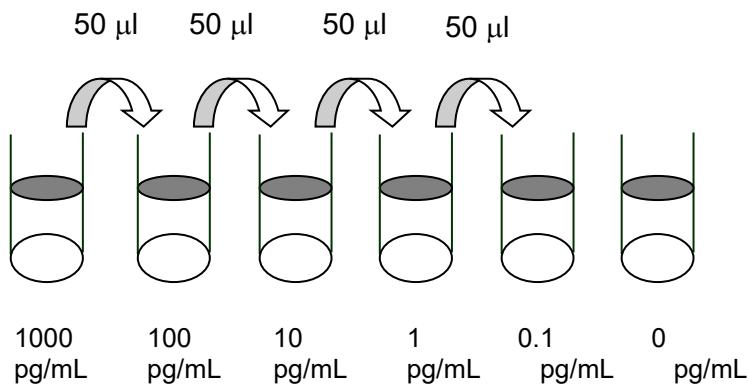
1. Keep kit reagents on ice during reagent preparation steps. Equilibrate plate to room temperature before opening the sealed pouch.

2. Briefly centrifuge the Neuropeptide Y Antibody vial (Item N) and reconstitute with 5 μ L of water before use. Add 50 μ L of 1x Assay Diluent E into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently.
3. The antibody concentrate should then be diluted 100-fold with 1x Assay Diluent E. This is the anti-Neuropeptide Y antibody working solution, which will be used in Procedure, step 2.

Note: The following steps may be done during the antibody incubation procedure (Procedure, step 2).

4. Briefly centrifuge the vial of Biotinylated Neuropeptide Y (Item F) and reconstitute with 20 μ L of water before use. Add 10 μ L of Item F to 5 mL of 1x Assay Diluent E. Pipette up and down to mix gently. The final concentration of biotinylated Neuropeptide Y will be 20 pg/mL. This solution will only be used as the diluent in Preparation, step 5.
5. Preparation of Standards: Label 6 microtubes with the following concentrations: 1,000, 100, 10, 1, 0.1, and 0 pg/mL. Pipette 450 μ L of biotinylated Neuropeptide Y solution into each tube, except for the 1,000 pg/mL (leave this one empty).

Figure 1.
Dilution Series for Standards



Note: It is very important to make sure the concentration of biotinylated Neuropeptide Y is 20 pg/mL in all standards.

- a. Briefly centrifuge the vial of Neuropeptide Y (Item C) and reconstitute with 10 μ L of water. In the tube labeled 1,000 pg/mL, pipette 8 μ L of Item C and 792 μ L of 20 pg/mL biotinylated Neuropeptide Y solution (Preparation, step 4). This is the Neuropeptide Y stock solution (1,000 pg/mL Neuropeptide Y and 20 pg/mL biotinylated Neuropeptide Y). Mix thoroughly. This solution serves as the first standard.
- b. To make the 100 pg/mL standard, pipette 50 μ L of Neuropeptide Y stock solution into the tube labeled 100 pg/mL. Mix thoroughly.
- c. Repeat this step with each successive concentration, preparing a dilution series (see Figure 1). Each time, use 450 μ L of biotinylated Neuropeptide Y and 50 μ L of the prior concentration until 0.1 pg/mL is reached. Mix each tube thoroughly before the next transfer.
- d. The final tube (0 pg/mL Neuropeptide Y, 20 pg/mL biotinylated Neuropeptide Y) serves as the zero standard (or total binding).

6. Prepare a 10-fold dilution of Item F. To do this, add 2 μ L of Item F to 18 μ L of the appropriate Assay Diluent. This solution will be used in Preparation, steps 7 and 9.
7. **Positive Control Preparation:** briefly centrifuge the positive control vial (Item M) and reconstitute with 100 μ L of water. To the tube of Item M, add 101 μ L of 1x Assay Diluent E. Also add 4 μ L of 10-fold diluted Item F (Preparation, step 6) to the tube. This is a 2-fold dilution of the positive control. Mix thoroughly. The positive control is a cell culture medium sample with an expected signal between 10–30% of total binding (70–90% of competition) if diluted as described. It may be diluted further if desired, but be sure the final concentration of Neuropeptide Y biotinylated is 20 pg/mL.
8. If Item B (20x Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 mL of Wash Buffer Concentrate into deionized or distilled water to yield 400 mL of 1x Wash Buffer.
9. **Sample Preparation:** Use 1x Assay Diluent E plus biotinylated Neuropeptide Y to dilute serum/plasma samples.

Note: It is very important to make sure the final concentration of the biotinylated Neuropeptide Y is 20 pg/mL in every sample.

For example, to make a 4-fold dilution of sample, mix together 5 μ L of 10-fold diluted Item F (Preparation, step 6), 182.5 μ L of 1x Assay Diluent E, and 62.5 μ L of the sample; mix gently. The total volume is 250 μ L, enough for duplicate wells on the microplate.

Do not use Item F diluent from Preparation, step 5 for sample preparation.

If undiluted samples are used, biotinylated Neuropeptide Y must be added to a final concentration of 20 pg/mL. For example, add 5 μ L of 10-fold diluted Item F to 245 μ L of sample.

10. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use. The HRP-Streptavidin concentrate should be diluted 200-fold with 1x Assay Diluent E.

Storage/Stability

Items C, F, and M should be stored at –20 °C. Avoid repeated freeze-thaw cycles.

The remaining kit components may be stored at 2–8 °C.

Opened microplate strips and Item N may be stored for up to 1 month at 2–8 °C. Return unused wells to the pouch containing desiccant pack and reseal along entire edge.

The kit remains active for up to 6 months.

Procedure

1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100 μ L of anti-Neuropeptide Y antibody (see Preparation, step 4) to each well. Incubate for 1.5 hours at room temperature with gentle shaking (1–2 cycles/sec) or incubate overnight at 4 °C.
3. Discard the solution and wash wells 4 times with 1x Wash Buffer (200–300 μ L each). Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 μ L of each standard (see Preparation, step 6), positive control (see Preparation, step 8), and sample (see Preparation, step 10) into appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1–2 cycles/sec) or overnight at 4 °C.
5. Discard the solution and wash 4 times as directed in step 3.
6. Add 100 μ L of prepared HRP-Streptavidin solution (see Preparation, step 11) to each well. Incubate for 45 minutes with gentle shaking at room temperature. It is recommended that incubation time should not be shorter or longer than 45 minutes.
7. Discard the solution and wash 4 times as directed in step 3.
8. Add 100 μ L of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1–2 cycles/sec).

9. Add 50 μ L of Stop Solution (Item I) to each well. Read absorbances at 450 nm immediately.

Results

Calculations

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit straight line through the standard points.

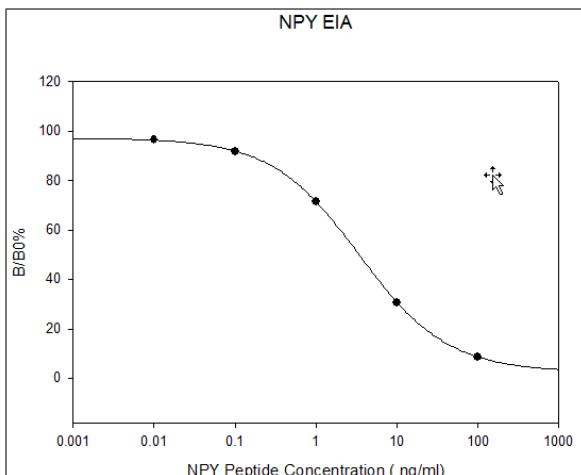
Percentage absorbance = $(B - \text{blank OD})/(B_0 - \text{blank OD})$ where

B = OD of sample or standard and

B_0 = OD of zero standard (total binding)

Typical Data

Standard curve(s) is for demonstration only. Standard curve(s) must be run with each assay.



Product Profile

Sensitivity: The minimum detectable concentration of Neuropeptide Y is 180 pg/mL or 16.59 pM.

Reproducibility:

Intra-Assay: CV <10%

Inter-Assay: CV <15%

Specificity

Cross Reactivity: This kit shows no cross-reactivity with any of the cytokines tested: Ghrelin, Nesfatin, Angiotensin II, and APC.

This kit was designed to recognize the C-terminus of NPY and, therefore, will detect full-length NPY.

However, it does not recognize active forms including NPY 1-36, 2-36, 3-36, or 3-35.

References

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Appendix
Troubleshooting Guide

Problem	Cause	Solution
Poor standard curve	Inaccurate pipetting	Check pipettes
	Improper standard dilution	Ensure a brief spin of Item C and dissolve the powder thoroughly with gentle mixing.
Low signal	Too brief incubation times	Ensure sufficient incubation time; Procedure, step 2 may change to overnight
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
Large CV	Inaccurate pipetting	Check pipettes
High background	Plate is insufficiently washed	Review the manual for proper wash. If using a plate washer, check that all ports are unobstructed.
	Contaminated wash buffer	Make fresh wash buffer
Low sensitivity	Improper storage of the ELISA kit	Store the standard at <-20 °C after reconstitution, others at 4 °C. Keep substrate solution protected from light
	Stop solution	Stop solution should be added to each well before measurement.

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