

# P 8092 **CYCLIN A PRIMER SET**

# **ProductInformation**

Cell division is a fundamental biological process, consisting of the splitting of the cell and its genetic material into two daughter cells. Regulation of cell cycle progression in eukaryotic cells depends on the expression of cyclin proteins. These proteins form complexes with the cell cycle dependent kinases (CDKs). Within these complexes, the cyclin subunit serves a regulatory role while the CDKs have a catalytic protein kinase activity. Complexes of cyclins and CDKs play a key role in the cell cycle by phosphorylating several cellular targets. Several cyclins have been identified, each having a specific activity in a distinct phase of the cell cycle. For example, members of the cyclin family when combined with p34 cdc2 kinase subunit, form an active cdc2 kinase which initiates M phase of mitosis and meiosis. Inactivation of these kinases is required for exit from mitosis. Besides involvement in the G2 to M transition, these complexes function as key regulators of each step of the cell cycle. 1 Cyclin A is one of the cyclins implicated to be active in M phase transition. It is a nuclear protein during S phase and disappears before metaphase. It binds both cdk2 and cdc2, giving two distinct cyclin A kinase activities, one appearing in G1/S phase, the other in G2/M.<sup>2</sup>

Cyclin A PCR Primer Set includes 2 synthetic oligonucleotides to be used in PCR<sup>H</sup> reaction for the detection of Cyclin A mRNA. The "sense" oligonucleotide represents the sequence between nucleotides 458-479 and the "antisense" oligonucleotide represents the sequence between nucleotides 737-758 on the human cyclin A mRNA coding region.\* Each primer is supplied in a separate tube and the complete set may be used for 50 amplification reactions (reaction volume of 50 µl). The primers are dried by Speed Vac and should be reconstituted in 50 µl of 1X PCR buffer to form a final working concentration of 20 µM. \* Gene bank accession No: X51688

## **REAGENTS PROVIDED**

Cyclin A sense primer, Product No. P 8217 5'-GTCACCACATACTATGGACATG-3' 1 vial

Cyclin A antisense primer, Product No. P 8342 5'-AAGTTTTCCTCTCAGCACTGAC-3'

1 vial

# MATERIALS AND REAGENTS REQUIRED BUT NOT PROVIDED

- Tag DNA Polymerase, Product No. D 4545 or D 1806
- Deoxynucleotide Mix, Product No. D 7295 10 mM dATP, 10 mM dCTP 10 mM dGTP, 10 mM TTP
  - Dilute to a final concentration of 2.5 mM deoxynucleotides with water
- Water, Product No. W 1754
- Mineral Oil, Product No. M 8662
- 10X PCR Buffer, Product No. P2192 or equivalent
- DNA to be amplified
- PCR pipet tips
- 0.5 ml thin wall PCR microcentrifuge tubes

# **STORAGE**

Store at -20 °C. Do not store in a "frost free freezer". Repeated freezing and thawing is **not** recommended.

## **PROCEDURE**

Use 1  $\mu$ I of each reconstituted primer in a 50  $\mu$ I PCR reaction mixture. Recommended annealing temperature: 53 °C.

Note: In order to obtain best results, determine optimal working conditions by titration test.

- 1. Spin down the dried primers in a microcentrifuge at maximal speed before reconstitution. Reconstitute the primers by adding 50 μl of 1X PCR buffer to each vial. Mix carefully to make sure all material is resuspended. Perform a short spin in a microcentrifuge at maximal speed.
- 2. Add the following reagents to a 500 μl microcentrifuge tube in the following order:
  - x μl Water (for a final volume of 50 μl)
  - 5 μl 10X PCR Buffer
  - 1 μl dNTP mix (2.5 mM)
  - 1 ul Sense Primer
  - 1 µl Antisense Primer
  - 0.5 μl Taq DNA Polymerase
  - y μl Template DNA (typically 10ng)
  - 50 μl Total volume
- 3. Mix gently by vortex and briefly centrifuge to collect all components to the bottom of the tube.
- 4. Add 100  $\mu$ l of mineral oil to the top of each tube to prevent evaporation.
- 5. The amplification parameters will vary depending on the primers and the thermocycler used. It may be necessary to optimize the system for individual primers, template, and thermocycler. Common cycling parameters are:
  - a. Denature the template at 95 °C for 1-2 minutes
  - b. Anneal primers at 53 °C for 1 minute
  - c. Extension at 72 °C for 1 minute

25-30 cycles of amplification are recommended.

- d. Final at 72 °C for 10 minutes
- e. Hold at 8 °C
- 6. The amplified DNA can be evaluated by agarose gel electrophoresis and subsequent ethidium bromide staining. Mineral oil overlay may be removed by a single chloroform extraction (1:1), recovering the aqueous phase.

The primers have been tested by RT-PCR. Their specificity has been tested on cloned plasmids encoding all the cyclins and no cross PCR amplification was observed. The Cyclin A PCR Primer Set amplifies a 300 bp fragment of the human and hamster cyclin A transcript from cDNA prepared from 80-90% confluent cell lines.

#### **REFERENCES**

- 1. Pines, J., and Hunter, T., J. Cell Biol., **115**, 1-17 (1991)
- 2. Pagano, M., et al., EMBO J. 11, 961-71, (1992)

†The PCR process is covered by patents owned by Hoffman-LaRoche, Inc.

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