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#### ChemiScreen<sup>™</sup> FLASH AEQUORIN CALCIUM-OPTIMIZED STABLE CELL LINE HUMAN RECOMBINANT EP₂ PROSTANOID RECEPTOR

CATALOG NUMBER:	HTS185AF	QUANTITY:	2 vials, 1 mL per vial
LOT NUMBER:	R0712E0023	CONCENTRATION:	2 x 10 <sup>6</sup> cells/mL
BACKGROUND:	Prostanoids bind to a family of 8 GPCRs to exert their biological effects (Narumiya a FitzGerald, 2001). The protanoid PGE <sub>2</sub> causes pain, vasodilation, immunosuppression of cells, bone resorption and promotion of carcinogenesis. Four related GPCRs, EP <sub>1</sub> , EP <sub>2</sub> , E and EP <sub>4</sub> , each bind to PGE <sub>2</sub> , but the different G protein coupling status of each receptor lea to distinct biological effects. EP <sub>2</sub> couples primarily to G <sub>s</sub> to increase intracellular cAMP leve Mice deficient in EP <sub>2</sub> receptor showed impaired ovulation and fertilization, and salt-sensiti hypertension (Kennedy <i>et al.</i> , 1999). It has been shown that EP <sub>2</sub> receptors are also involv in cancer-associated immunodeficiency. Thus, genetic knockout of the EP <sub>2</sub> receptor reduct tumor growth and prolonged survival in mice that had undergone isograft injection of MC26 Lewis lung carcinoma cells (Yang <i>et al.</i> , 2003). Millipore's cloned human EP <sub>2</sub> -expressing cline is made in the Chem-9 host which stably expresses a mitochondrially targeted flamutant form of aeguorin.		

ideal tool for screening for agonists and antagonists at EP<sub>2</sub>.

#### APPLICATIONS: Luminescent and fluorescent calcium flux assays, ligand binding assays



higher luminescent signal intensity than purfied wildtype aequorin. Thus, the cell line is an

**Figure 1.** Ligand-induced calcium flux in Flash Aequorin Chem-9 cell line stably transfected with EP<sub>2</sub>. Flash Aequorin Chem-9 stably co-expressing EP<sub>2</sub> were loaded with 5  $\mu$ M coelenterazine for 3 h at room temperature. Luminescence in response to PGE<sub>2</sub> was determined (A) in quadruplicate in a 384 well plate on a FLIPR<sup>TETRA</sup> with aequorin option from Molecular Devices, now part of MDS Analytical Technologies. Data were collected for area under curve for 70 sec. (B) PGE<sub>2</sub>-induced luminescence was determined in duplicate in a 96 well plate with a PerkinElmer Wallac Victor2. Data were collected for area under curve for 20 sec.

SPECIFICATIONS: EC50 for calcium mobilization by PGE<sub>2</sub>: ~ 123.8 nM (FLIPR<sup>TETRA</sup>) and ~ 95.4 nM (Wallac Victor2)

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HOST CELLS: Chem-9, an adherent cell line expressing the promiscuous G-protein.

TRANSFECTION: Full-length human PTGER2 cDNA encoding EP<sub>2</sub> (Accession Number: NM\_000956)

### **PRESENTATION:** Cells are frozen at $2 \times 10^6$ cells/mL in 90% fetal bovine serum/10% DMSO. Cell line tests negative for mycoplasma.

STORAGE/HANDLING

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- 1. Immediately upon receipt, thaw cells or place cells in liquid nitrogen. Maintain frozen in liquid nitrogen for up to 5 years.
- Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol. Transfer contents of the vial to a T75 flask containing growth media. Place the flask in a humidified incubator at 37°C with 5% CO<sub>2</sub>.
- 3. After 8-24 h, all live cells will be attached. Viability of the cells is expected to be 50-80%. At this time, replace media to remove residual DMSO, and return to incubator.
- 4. When cells are approximately 80% confluent, passage the cells as follows: Remove media and wash once with HBSS without Ca<sup>++</sup> and Mg<sup>++</sup> (10 mL/T75). Add 0.05% trypsin/0.2 g/L EDTA (1 mL/T75) and place in humidified incubator at 37°C with 5% CO<sub>2</sub> until cells begin to round up and detach (2-4 minutes). Gently rap the side of the flask to dislodge the cells. Neutralize trypsin by addition of 4 mL Chem-9 Aequorin Growth Media per 1 mL trypsin.
- 5. Cells are typically passaged 1:10 every 3-4 days. Passaging ratio may be varied according to requirements of the investigator.
- 6. Frozen stocks of cells should be prepared at the earliest passage possible after thawing, as follows: Count detached cells (prepared as in Step 4). Centrifuge cells at 200 x g for 5 min. Resuspend cells at 5 x 10<sup>6</sup> cells/mL in Freezing Media (cell densities of 2-10 x 10<sup>6</sup> are also acceptable if necessary). Dispense 1 mL aliquots into cryopreservation vials. Freeze the cells by a controlled rate process, such as in an isopropanol-jacketed container placed at –70°C overnight. Store the vials in liquid nitrogen.
- 7. Use of cells immediately after thawing is feasible for some cell lines and is being further validated. Some cell lines may need to be passaged at least once after thawing prior to use in calcium flux assays.

MEDIA: Chem-9 Aequorin Growth Media: DMEM with 4.5 g/L glucose and 4 mM glutamine (Millipore SLM-020-A) 10% heat-inactivated FBS 1x Nonessential amino acids (from 100x stock, Millipore TMS-001-C) 10mM HEPES (from 1 M HEPES, Millipore TMS-003-C) 100 U/mL Pen-Strep (from 100x stock, Millipore TMS-AB2-C) 250 μg/mL Genetecin/G-418 250 μg/mL Hygromycin 500 μg/mL Zeocin

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Chem-9 Aequorin Plating Media: DMEM with 4.5 g/L glucose and 4 mM glutamine 10% heat-inactivated FBS 1x NEAA 10mM HEPES 1x Pen-Strep

Freezing Media: 90% heat-inactivated FBS 10% DMSO (cell culture grade)

#### RECOMMENDED ASSAY CONDITIONS:

- Seed cells in 96-well white plate (top-read instruments) or opaque walled, clear bottom plate (bottom-read instruments) overnight at 50,000 cells/well in Chem-9 Aequorin Plating Media.
- Wash cells once (200 μl/well) with Wash Buffer (HBSS with Ca<sup>++</sup> and Mg<sup>++</sup> containing 10 mM HEPES) before loading with 5μM of coelenterazine (Millipore ES016) in wash buffer at room temperature for 3 hours.

**Note:** Luminescence activity has been determined to be optimal at room temperature. Incubation at 37 °C will result in substantially reduced signals.

3. After loading, wash cells once with Wash Buffer (200 µl/well) prior to addition of ligands.

**REFERENCE:** 

Kennedy CR *et al.* (1999) Salt-sensitivity hypertension and reduced fertility in mice lacking the prostaglandin EP<sub>2</sub> receptor. *Nat. Med.* 5:217-220.

Narumiya S and FitzGerald GA (2001) Genetic and pharmacological analysis of prostanoid receptor function. *J. Clin. Invest.* 108: 25-30.

Yang N *et al.* (2003) Cancer-associated immunodeficiency and dendritic cell abnormalities mediated by the prostaglandin EP<sub>2</sub> receptor. *J. Clin. Invest.* 111: 727–735.

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