

Technical Bulletin

EX-CELLTM Sp2/0: An Animal-Component Free, Protein-Free, Chemically Defined **Medium for Monoclonal Antibody Production**

Introduction

SAFC Biosciences has developed EX-CELL™ Sp2/0 Serum-Free Medium for Sp2/0 Cells, an animal-component free, protein-free, chemically defined, serum-free medium for growth and monoclonal antibody (MAb) production in Sp2/0-derived hybridoma cell lines. EX-CELL™ Sp2/0 is hydrolysate-free and contains no animal-or human-derived components.

The following study was undertaken to demonstrate the ability of EX-CELL™ Sp2/0 to support growth and MAb production in the mSxl 5 cell line (an Sp2/0-derived hybridoma) that secretes mouse MAb immunoglobulin G (lgG₁). The cell line was adapted to EX-CELL™ Sp2/0 by direct adaptation and growth and MAb production were assessed.

Materials

Cells

• mSxl 5, American Type Culture Collection, ATCC No. CRL-1951

Media and Supplements

- EX-CELL™ Sp2/0, SAFC Biosciences, Catalog No. 14660
- L-Glutamine Solution 200 mM, SAFC Biosciences, Catalog No. 59202
- RPMI 1640 Medium, SAFC Biosciences, Catalog No. 51502
- Fetal Bovine Serum Gamma Irradiated (FBS), SAFC Biosciences, Catalog No. 12107

Antibody Determination

• HPLC gel permeation column, ZORBAX GF-450, Agilent **Technologies**

Methods

Media/Supplement Preparation and Storage

Prior to use, EX-CELL™ Sp2/0 was supplemented with 8 mM L-glutamine on day zero. All media were stored at 2 to 8 C protected from light. Other supplements were stored at their recommended temperatures. Cultures were maintained using aseptic technique with no antibiotic or fungicide supplementation.

Culture Techniques

The mSxl 5 cells were cultured in 125 mL Erlenmeyer shaker flasks (30 mL media volume) at 165 rpm in a 37 C humidified incubator with 10% CO₂. Cultures were initiated in RPMI 1640 + 5% FBS and were subcultured every three days at a seeding density of 3 x 10^s cells/mL. The cultures were then directly adapted (without weaning) to EX-CELL™ Sp2/0 supplemented with L-glutamine, using the same culture conditions as described above. Cultures were monitored over multiple passages to assess growth kinetics and viability during and after the adaptation period. Cell counts and viability were determined daily by trypan blue exclusion and small aliquots of supernatant were also removed daily for IgG analysis. IgG concentration was determined by HPLC using ImmunoPure Mouse IgG as the standard.

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Results

Adaptation and Growth Studies

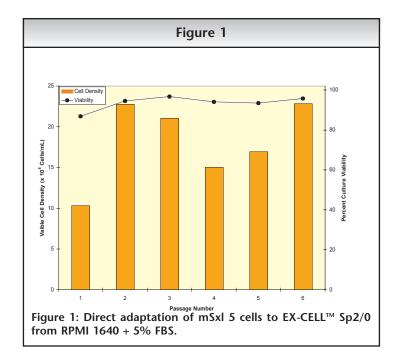
The mSxl 5 cells were adapted directly from RPMI 1640 Medium + 5% FBS into EX-CELLTM Sp2/0. Figure 1 illustrates direct adaptation. During the first passage in EX-CELLTM Sp2/0 cell counts were approximately 25 - 50% lower than average cell densities at each pass; however, they quickly recovered and displayed normal densities and viabilities thereafter. The mSxl 5 cells typically reached densities of 1.5-2.5 x 10^6 cells/mL and exhibited doubling times of 20 - 30 hours when subcultured every three days in EX-CELLTM Sp2/0 (Figure 1).

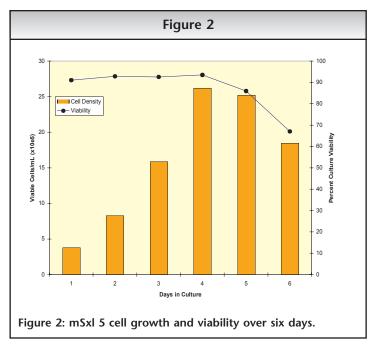
Figure 2 shows growth kinetics of mSxl 5 cells over a period of six days in EX-CELL™ Sp2/0 media. The cell densities per mL increase steadily over the first four days, reach a plateau by day five and decline after day five. Viabilities start to drop after day four and are below 70% on day six.

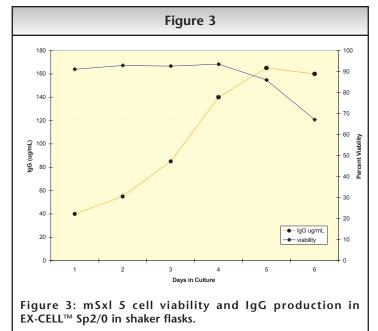
A typical viability and concurrent IgG production of adapted mSxl 5 cells in EX-CELLTM Sp2/0 is shown in Figure 3. Viabilities stay above 90% until day four and start to decline thereafter. IgG production steadily increases with peak productivity on days five and six.

Conclusions

SAFC Biosciences developed an animal-component free, protein-free, chemically defined, serum-free medium that supports the growth of mSxl 5 cells in suspension cultures, attaining densities of 2.5 x 10° cells/mL. Monoclonal antibody production remains in the 160 mg/L range at day five while viabilities have started to decline. EX-CELL™ Sp2/0 supports Sp2/0-derived hybridoma cell growth and MAb production.







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