3050 Spruce Street, St. Louis, MO 63103 USA
Tel: (800) 521-8956 (314) 771-5765 Fax: (800) 325-5052 (314) 771-5757
email: techservice@sial.com sigma-aldrich.com

Product Information

CryoSOfree™ DMSO-free Cryopreservation Medium

Catalog Number **C9249**
Storage Temperature 2–8 °C

Product Description

Cultured cells in a suspension can be cryopreserved with minimal damage from freezing and thawing steps. CryoSOfree™ DMSO-free Cryopreservation Medium preserves them with high viability and physiological activity. This medium is recommended for the preservation of stem cells, hepatocytes, tissue samples, and a wide variety of cell types and established cell lines.

CryoSOfree DMSO-free Cryopreservation Medium is a proprietary formulation, containing no DMSO nor animal proteins. It is ready-to-use and complete with no additives required.

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Preparation Instructions

The CryoSOfree DMSO-free Cryopreservation Medium Is ready-to-use and complete with no additives required. Wipe down the outside of the container with a 70% ethanol solution before opening. If the tamper evident seal has been broken, do not use.

Storage/Stability

Store the CryoSOfree medium at 2–8 °C.

Procedure

1. Freezing cell suspensions
   a. Centrifuge the cultured cells suspended in medium and then remove the supernatant from the cell pellet at 4–10 °C.
   b. Resuspend the cell pellet with CryoSOfree DMSO-free Cryopreservation Medium at a concentration of 5×10^5 to 5×10^6 cells/mL.
   c. Pipette the cell suspension into a cryotube at 1 mL/vial and freeze them at –80 °C.
   d. For long term preservation, transfer the cryotube into a liquid nitrogen chamber (–196 °C) on the following day.

2. Restoring preserved cells
   a. Warm the cryotube containing the preserved cells in a waterbath at 37 °C by gently shaking the tube and thaw the contents.
   b. Centrifuge the cells and wash in cold PBS before transferring the cells to an experimental medium. Whether the live cells are used for experiments or culturing, place them under proper physiological conditions as soon as possible (e.g., 37 °C).

Results

Viability assessment 24-hours post-thaw - Live/Dead fluorescent assays or metabolic assays (MTT or resazurin) are recommended for more accurate viability assessment. Visual inspection of adherent cells and cells “floating” in the medium are also recommended.
Notes: Sample assessment immediately post-thaw with membrane integrity indicators, such as trypan blue for comparative analysis of sample cell yield and viability, often results in significant overestimation of cell survival.

To obtain an accurate measure of cell viability following cryopreservation, assessment should be performed 24 hours post-thaw and compared to non-frozen controls.

CryoSOfree is a trademark of Sigma-Aldrich Co. LLC.

VA,DF,MAM 11/21-2