SIGMA

ProductInformation

CYCLIN B_1 PRIMER SET
Product Number **P 8467**

Product Description

Cell division is a fundamental biological process, consisting of the splitting of the cell and its genetic material into two daughter cells. Regulation of cell cycle progression in eukaryotic cells depends on the expression of cyclin proteins. These proteins form complexes with the cell cycle dependent kinases (CDKs). Within these complexes, the cyclin subunit serves a regulatory role while the CDKs have a catalytic protein kinase activity. Complexes of cyclins and CDKs play a key role in the cell cycle by phosphorylating several cellular targets. Several cyclins have been identified, each having a specific activity in a distinct phase of the cell cycle. The family of B cyclins when combined with p34 ^{cdc2} (CDK-1) form an enzyme complex which is active at the transition to the G2 phase, attaining maximum level at M phase. In prophase and metaphase, the complex associates with condensed chromosomes and is degraded at the metaphase-anaphase transition.

Cyclin B_1 PCR Primer Set includes 2 synthetic oligonucleotides to be used in a PCR† reaction for the detection of cyclin B_1 mRNA. The “sense” oligonucleotide represents the sequence between nucleotides 25-50 and the “antisense” oligonucleotide represents the sequence between nucleotides 482-504 on the hamster cyclin B_1 mRNA coding region.* Each primer is supplied in a separate tube and the complete set may be used for 50 amplification reactions (reaction volume of 50 µl). The primers are dried by Speed Vac and should be reconstituted in 50 µl of 1X PCR buffer to form a final working concentration of 20 µM.

* Gene bank accession No: D17293

Reagents

Reagents Provided

- **Cyclin B_1 sense primer**, 1 vial
  Product No. P 8592

  5′-CGAAACTTAACACAGAAAATAAGGCC-3′

- **Cyclin B_1 antisense primer**, 1 vial
  Product No. P 8717

  5′-CTTTCACATATTCGCTACAGAGG-3′

Materials and Reagents required but not provided

- *Taq* DNA Polymerase, Product No. D 4545 or D 1806
- Deoxynucleotide Mix, Product No. D 7295
  10 mM dATP, 10 mM dCTP
  10 mM dGTP, 10 mM TTP
  Dilute to a final concentration of 2.5 mM deoxynucleotides with water
- Water, Product No. W 1754
- Mineral Oil, Product No. M 8662
- 10X PCR Buffer, Product No. P 2192 or equivalent
- DNA to be amplified
- PCR pipet tips
- 0.5 ml thin wall PCR microcentrifuge tubes

Storage/Stability

Store at –20 °C. Do not store in a “frost free freezer”. Repeated freezing and thawing is **not** recommended.

Procedure

Use 1 µl of each reconstituted primer in a 50 µl PCR reaction mixture.
Recomended annealing temperature: 53 °C.

Note: In order to obtain best results, determine optimal working conditions by titration test.

1. Spin down the dried primers in a microcentrifuge at maximal speed before reconstitution. Reconstitute the primers by adding 50 µl of 1X PCR buffer to each vial. Mix carefully to make sure all material is resuspended. Perform a short spin in a microcentrifuge at maximal speed.

2. Add the following reagents to a 500 µl microcentrifuge tube in the following order:

x	µl Water (for a final volume of 50 µl)
5	µl 10X PCR Buffer
1	µl dNTP mix (2.5 mM)
1	µl Sense Primer
1	µl Antisense Primer
0.5	µl *Taq* DNA Polymerase
y	µl Template DNA (typically 10 ng)
50	µl Total volume

3. Mix gently by vortex and briefly centrifuge to collect all components to the bottom of the tube.

4. Add 100 µl of mineral oil to the top of each tube to prevent evaporation.

5. The amplification parameters will vary depending on the primers and the thermocycler used. It may be necessary to optimize the system for individual primers, template, and thermocycler. Common cycling parameters are:

   a. Denature the template at 95 °C for 1-2 minutes
   b. Anneal primers at 53 °C for 1 minute
   b. Extension at 72 °C for 1 minute

   25-30 cycles of amplification are recommended.

   d. Final at 72 °C for 10 minutes
   e. Hold at 8 °C

6. The amplified DNA can be evaluated by agarose gel electrophoresis and subsequent ethidium bromide staining. The mineral oil overlay may be removed by a single chloroform extraction (1:1) and recovering the aqueous phase.

The primers have been tested by RT-PCR. Their specificity has been tested on cloned plasmids encoding all the cyclins and no cross PCR amplification was observed. The Cyclin B_1 PCR Primer Set amplifies a 479 bp fragment of the rat and hamster cyclin B_1 transcript in 80-90% confluent cell lines.

References

1. Pines, J., and Hunter , T. , J. Cell Biol., **115,** 1-17 (1991).
2. Sherwood, S. W., *et al.*, Exp. Cell Res. **211**, 275-281 (1991).

†The PCR process is covered by patents owned by Hoffman-LaRoche, Inc.

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