ProductInformation

ANTI-MATRIX METALLOPROTEINASE-18 (MMP-18), C-TERMINAL
Developed in Rabbit, Affinity Isolated Antibody

Product Number **M 4934**

Product Description

Anti-Matrix Metalloproteinase-18 (MMP-18) is developed in rabbit using a synthetic peptide corresponding to the C-terminal of *Xenopus* MMP-18 (collagenase-4, *Xenopus* collagenase) as immunogen. Affinity isolated antigen specific antibody is obtained from rabbit anti-MMP-18 antiserum by immuno-specific purification which removes essentially all rabbit serum proteins, including immunoglobulins, which do not specifically bind to the peptide.

Rabbit Anti-MMP-18, C-terminal may be used for the detection and localization of *Xenopus* MMP-18 by various immunochemical techniques including immunoblotting, immunoprecipitation, immunohistochemistry, and ELISA.

Rabbit Anti-MMP-18, C-terminal specifically binds to MMP-18 and does not cross-react with the other MMP family members (MMP-1, MMP-2, MMP-3, MMP-9, etc). The antibody recognizes the pro-form and active forms of MMP-18, as well as further activation/breakdown products. By immunoblotting against the reduced protein, the antibody reacts with bands at 53 kDa and 51 kDa, as well as the smaller activated forms. Anti-MMP-18, C-terminal also recognizes non-reduced MMP-18.

The matrix metalloproteinases (MMPs) are a family of at least eighteen secreted and membrane-bound zinc-endopeptidases. Collectively, these enzymes can degrade all the components of the extracellular matrix, including fibrillar and non-fibrillar collagens, fibronectin, laminin and basement membrane glycoproteins. In general, a signal peptide, a propeptide, and a catalytic domain containing the highly conserved zinc-binding site characterizes the structure of the MMPs. In addition, fibronectin-like repeats, a hinge region, and a C-terminal hemopexin-like domain allow categorization of MMPs into the collagenase, gelatinase, stomelysin and membrane-type MMP subfamilies.[1-3] MMPs contain the motif His-Glu-X-X-His (X represents any amino acid) that binds zinc in the catalytic site, as well as another zinc molecule and two calcium molecules structurally. They fall within the matrixin subfamily and are EC designated 3.4.24.x. This group also contains astacin, reprolysin, and serralysin, as well as other more divergent metalloproteinases. All MMPs are synthesized as proenzymes, and most of them are secreted from the cells as proenzymes. Thus, the activation of these proenzymes is a critical step that leads to extracellular matrix breakdown.

MMPs are considered to play an important role in wound healing, apoptosis, bone elongation, embryo development, uterine involution, angiogenesis,[4] and tissue remodeling, and in diseases such as multiple sclerosis,[2, 5] Alzheimer's,[2] malignant gliomas,[2] lupus, arthritis, periodontis, glumerulonephritis, atherosclerosis, tissue ulceration, and in cancer cell invasion and metastasis.[6] Numerous studies have shown that there is a close association between expression of various members of the MMP family by tumors and their proliferative and invasive behavior and metastaic potential.

The tissue inhibitors of metalloproteinases (TIMPs) are naturally occurring proteins that specifically inhibit matrix metalloproteinases and regulate extracellular matrix turnover and tissue remodeling by forming tight-binding inhibitory complexes with the MMPs. Thus, TIMPs maintain the balance between matrix destruction and formation. An imbalance between MMPs and the associated TIMPs may play a significant role in the invasive phenotype of malignant tumors. MMPs and TIMPs can be divided into two groups with respect to gene expression: the majority exhibit inducible expression and a small number are produced constitutively or are expressed at very low levels and are not inducible. Among agents that induce MMP and TIMP production are the inflammatory cytokines TNF-α and IL-1β. A marked cell type specificity is a hallmark of both MMP and TIMP gene expression (i.e., a limited number of cell types can be induced to make these proteins).

Matrix Metalloproteinase-18 (MMP-18), also known as collagenase-4 and *Xenopus* collagenase, is an interstitial collagenase. This protein was originally described in *Xenopus* during early development.[7] *Xenopus* collagenase (MMP-18) has no sequence homology with any of the other known collagenases

and is distinct from MMP-1, which it most closely resembles. This protein digests intact type-I collagen at the same site as the classical collagenases (MMP-1, MMP-8, and MMP-13).

Xenopus collagenase may facilitate larval tissue degeneration and adult organogenesis during amphibian metamorphosis.[8] MMP-18 (*Xenopus* collagenase) is regulated in a tissue-dependent manner. During metamorphosis, MMP-18 is transiently produced in the gastrula stage, expressed in the neurula stage, and down-regulated in the pre-tail bud stage of embryo development.[7]

Reagent
Rabbit Anti-MMP-18, C-Terminal is supplied in 0.01 M phosphate buffered saline, pH 7.4, containing 50 % glycerol and 0.1 % sodium azide.
Protein concentration is approximately 1 mg/ml.

Precautions and Disclaimer
Due to the sodium azide content a material safety data sheet (MSDS) for this product has been sent to the attention of the safety officer of your institution. Consult the MSDS for information regarding hazards and safe handling practices.

Storage/Stability
For continuous use, store at 2 °C to 8 °C for up to six months. For extended storage, the solution may be stored 0 °C to –20 °C. The antibody is supplied with 50 % glycerol to prevent freezing. If slight turbidity occurs upon prolonged storage, clarify the solution by centrifugation before use.

Product Profile
A working dilution of 1:1,000 is determined by immunoblotting using an alkaline phosphatase conjugated secondary antibody and BCIP/NBT as the substrate.

Note: MMP-18 is produced at very low levels in normal, quiescent Xenopus cells and tissues. Collagenase production can be stimulated by mitogens such as TPA or TNF-α in receptive cell lines.

In order to obtain best results and assay sensitivity in different techniques and preparations we recommend determining optimum working dilutions by titration assay.

References

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