Product Information

REDExtract-N-Amp™ Blood PCR Kits

XNABS, XNAB, XNABE, XNABR, P8240

Product Description

The REDExtract-N-Amp™ Blood PCR Kits for direct PCR contain the reagents required to rapidly extract and amplify human genomic DNA from whole blood, whole blood dried on a blood card, and cultured mammalian cells without the need for purification, organic extraction, filtration or alcohol precipitation. Briefly, DNA is released by incubating the sample with the Lysis Solution at room temperature for 5 minutes for whole blood, at 55 °C for 15 minutes for blood cards, or at 75 °C for 5 to 10 minutes for cell monolayers. After adding the Neutralization Solution, the extract is ready for PCR.

An aliquot of the neutralized extract is then combined with the REDExtract-N-Amp™ Blood PCR ReadyMix and user-provided PCR primers to amplify the target DNA. The REDExtract-N-Amp™ Blood PCR ReadyMix is a 2x reaction mixture containing buffer, salts, dNTPs, and *Taq* polymerase. It also contains the JumpStart™ *Taq* antibody for hot start PCR to enhance specificity and the RED*Taq*® dye to allow direct loading of the PCR product onto an agarose gel.

Reagents Provided	Cat. No.	XNABS 10 preps, 10 PCRs	XNAB 100 preps, 100 PCRs	XNABE 100 preps, 500 PCRs	XNABR 1,000 preps, 1,000 PCRs
Lysis Solution for Blood	L3289	0.3 mL	2.5 mL	2.5 mL	25 mL
Neutralization Solution for Blood	N9784	2 x 1.5 mL	25 mL	25 mL	250 mL
REDExtract-N-Amp™ Blood PCR ReadyMix. This is a 2X PCR reaction mix containing buffer, salts, dNTPs, *Taq* polymerase, RED*Taq*® dye, and JumpStart™ *Taq* antibody.	P8240	0.15 mL	1.2 mL	5 x 1.2 mL	12 mL

Storage

The REDExtract-N-Amp™ Blood PCR Kits can be stored at 2-8 °C on a short-term basis up to 3 weeks. For long-term storage greater than 3 weeks, -20 °C is recommended. Do not store in a "frost-free" freezer.

Reagents and Equipment Required

(Not provided)

- Microcentrifuge tubes or multi-well plate for extractions (200 µL minimal volume)
- Punch and cards for dried blood
- Incubator or oven for blood cards (55 °C) or monolayer cells (75 °C)
- Tubes or plate for PCR
- Thermal cycler
- PCR primers, Cat. No. OLIGO
- Water, PCR reagent, Cat. No. W1754
- AlumaSeal® II Film, Cat. No. A2350

Precautions and Disclaimer

The REDExtract-N-Amp™ Blood PCR Kits are for R&D use only. Not for drug, household, or other uses. The Lysis Solution is caustic. Avoid contact with skin. Wear gloves, safety glasses, and suitable protective clothing when handling this or any other reagent provided with the kit. Consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Procedure

All steps are carried out at room temperature unless otherwise noted.

DNA extraction from whole blood

1. Collect blood into tubes containing EDTA, sodium citrate, or sodium heparin. The best results may be obtained with EDTA or sodium citrate. Mix thoroughly by inversion or rocking.

   Note: For non-human sources, collect blood into tripotassium EDTA, Cat. No. E0270, at a final concentration of 5 mM to prevent coagulation.
2. Place 20 µL of the Lysis Solution for Blood into a microcentrifuge tube or well of a multi-well plate for each extraction.
3. Add 10 µL of blood. Mix thoroughly by vortexing or pipetting.
4. Incubate at room temperature for 5 minutes.
5. Add 180 µL of the Neutralization Solution for Blood. Mix thoroughly by vortexing or pipetting.
6. Store the neutralized blood extract at 4 °C or use 2 µL immediately in PCR. Continue to PCR amplification, step 1.

   Note: DNA is stable in the extract for at least 6 months at 4 °C.

DNA extraction from blood cards

1. Collect the blood sample on to a collection card, such as Cat. No. WHAWB100014. Allow to dry completely.
2. Punch a disk (preferably 1/8 inch or 3 mm) from the blood card and place into a microcentrifuge tube. Make sure that the punch contains as much of the blood-stained area as possible.
3. Pipette 20 µL of the Lysis Solution for Blood onto the blood card punch. Samples can be spun in a microcentrifuge for a few seconds to force the solution into the punch.
4. Incubate at 55 °C for 15 minutes.
5. Add 180 µL of the Neutralization Solution for Blood. Mix thoroughly by vortexing or pipetting.
6. Store the neutralized blood extract at 4 °C or use 2 µL immediately in PCR. Continue to PCR amplification, step 1.

   Note: DNA is stable in the extract for at least 6 months at 4 °C.

DNA extraction from cultured mammalian cells

1. Grow monolayer cells in a multi-well plate until 90 to 95% confluent.
2. Aspirate the medium from the wells using a pipette tip connected to a vacuum system. The medium must be removed completely.
3. Add 20 µL of the Lysis Solution for Blood to each of the wells.

   Note: It is preferred at this point to seal the plate with AlumaSeal® II Film, Cat. No. A2350, to prevent loss by evaporation during incubation in step 4. The plate seal can be pierced with a pipette tip to add the Neutralization Solution for Blood in step 5. A new layer of plate seal can be placed over the original layer to reseal the plate for storage.
4. Incubate the plate at 75 °C for 5 to 10 minutes (for a 24-well plate, 5 minutes is recommended to avoid over drying the samples).
5. Add 180 µL of the Neutralization Solution for Blood to each of the wells. Mix the samples by pipetting up and down.
6. Store the neutralized cell extract at 4 °C or use 2 µL immediately in PCR. Continue to PCR amplification, step 1.

   Note: DNA is stable in the extract for at least 6 months at 4 °C.

PCR amplification

The REDExtract-N-Amp™ Blood PCR ReadyMix contains the JumpStart™ *Taq* antibody for specific hot start amplification. Therefore, PCR reactions can be assembled at room temperature without premature *Taq* DNA polymerase activity.

Typical final primer concentrations are approximately 0.4 µM each. The optimal primer concentration and cycling parameters will depend on the system used.

1. Add the following reagents to a thin-walled PCR microcentrifuge tube or plate:

Reagent	Volume
Water, PCR Reagent	Variable
REDExtract-N-Amp™ Blood PCR ReadyMix	10 µL
Forward primer	Variable
Reverse primer	Variable
Neutralized blood extract	2 µL
Total volume	20 µL

Note: The neutralized blood extract may inhibit PCR amplification of products larger than 2 kb. Neutralization Solution B, Cat. No. N3910, can be used to overcome this inhibition and allows successful amplification of longer PCR products. Add 1 µL of Neutralization Solution B to each reaction. Neutralization Solution B is not part of this kit and must be purchased separately.

2. Mix gently.
3. For thermal cyclers without a heated lid, add 20 µL of mineral oil on top of the mixture in each tube to prevent evaporation.
4. Perform thermal cycling. The amplification parameters should be optimized for individual primers, template, and thermal cycler (see References for guidance).

Common cycling parameters:

Step	Temp.	Time	Cycles
Initial Denaturation	94 °C	3 minutes	1
Denaturation	94 °C	30 seconds	
Annealing	45-68 °C	30 seconds	30-40
Extension	72 °C	1-2 minutes (~1 min/kb)	
Final Extension	72 °C	10 minutes	1
Hold	4 °C	Indefinitely	

5. The amplified DNA can be loaded directly onto an agarose gel after the PCR is completed. It is not necessary to add a separate loading buffer/tracking dye.

   Note: PCR products can be purified, if desired, for applications such as sequencing with the GenElute™ PCR Clean-Up Kit, Cat. No. NA1020.

Product Ordering

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Related Products	Cat. No.
PCR 96-well plates	Z374903
PCR 384-well plates	Z374911
Sealing mats and tape	Z374938
AlumaSeal® II	A2350
EDTA, tripotassium salt dihydrate	E0270
Collection Card	WHAWB100014
PCR microtubes	Z374873, Z374962, Z374881
Neutralization Solution B	N3910
Mineral Oil	M8662
PCR Marker	P9577
Precast Agarose Gels	P6097
TBE Buffer	T4415, T6400, T9525
GenElute™ PCR Clean-Up Kit	NA1020

Troubleshooting

Problem	Cause	Solution
Little or no PCR product is detected.	PCR reaction is inhibited due to contaminants in the blood extract.	Use less extract or dilute the extract with water and repeat PCR. To test for inhibition, include a DNA control and/or add a known amount of template (100-500 copies) into the PCR mixture along with the blood extract.
	A PCR component is missing or degraded.	Run a positive control to ensure components are functioning. A checklist is also recommended when assembling reactions.
	Too few cycles are performed.	Increase the number of cycles (5-10 additional cycles at a time).
	The annealing temperature is too high.	Decrease the annealing temperature in 2-4 °C increments.
	The primers are not designed optimally.	Confirm the accuracy of the sequence information. If the primers are less than 22 nucleotides long, try to lengthen the primer to 25-30 nucleotides. If the primer has a GC content of less than 45%, try to redesign the primer with a GC content of 45-60%.
	The extension time is too short.	Increase the extension time in 1-minute increments, especially for long templates.
	The target template is complex.	In most cases, inherently complex targets are due to unusually high GC content and/or secondary structure. Betaine has been reported to help amplification of high GC content templates at a concentration of 1.0-1.7 M.
Multiple products are seen.	JumpStart™ *Taq* antibody is not working correctly.	Do not use DMSO or formamide with REDExtract-N-Amp™ PCR ReadyMix. It can interfere with the enzyme-antibody complex. Other solvents, salts, and extremes in pH or other reaction conditions may reduce the affinity of the JumpStart™ *Taq* antibody for the *Taq* polymerase and thereby compromise its effectiveness.
	Touchdown PCR may be needed.	"Touchdown" PCR significantly improves the specificity of many PCR reactions in various applications. Touchdown PCR uses an annealing/extension temperature that is higher than the TM of the primers during the initial PCR cycles. The annealing/extension temperature is then reduced to the primer TM for the remaining PCR cycles. The change can be performed in a single step or in increments over several cycles.
Negative control shows a PCR product or "false positive" results are obtained.	Reagents are contaminated.	Include a reagent blank without DNA template be included as a control in every PCR run to determine if the reagents used in extraction or PCR are contaminated with a template from a previous reaction.

References

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