



Fluorescent Human ES/iPS Cell Characterization Kit

Catalog No. SCR078

FOR RESEARCH USE ONLY
Not for use in diagnostic procedures.

Introduction

Human embryonic stem (hES) cells are pluripotent cells derived from the inner cell mass of pre-implantation blastocysts¹. Human induced pluripotent (hiPS) cells are pluripotent cells generated by reprogramming human somatic cells using four transcription factors, Oct-4, Klf-4, Sox-2, and c-Myc, or their variants². Both hESC and hiPSC can self-renew and have the ability to generate all three germ layers: ectoderm, mesoderm, and endoderm. In vitro, hESC/iPSC are normally maintained and propagated on mouse fibroblast feeders for extended periods in media containing basic fibroblast growth factor (bFGF)³. However, spontaneous differentiation may occur in subpopulations of cells. Several pluripotent markers are commonly used to distinguish pluripotent hESC/iPSC from differentiated cells.

- **Alkaline phosphatase (AP)** is an enzyme that hydrolyzes the phosphate group from many types of molecules, including nucleotides, proteins and alkaloids. Although AP is primarily found in liver and bone, pluripotent stem cells have also been found to have elevated expression of AP⁵. Both human and mouse ESC/iPSC are characterized by high expression levels of AP.
- **Oct4, Sox-2, and Nanog** are three transcription factors that are highly expressed in pluripotent cells. They share a significant proportion of their target genes and form the core transcriptional regulatory circuitry that contributes to pluripotency and self-renewal of hESC/iPSC⁶. The successful reprogramming of somatic cells with Oct-4, Sox-2 together with Klf-4 and c-Myc genes further confirms the essential role of these transcription factors in maintaining pluripotency^{2,4}.
- **TRA-1-60 and TRA-1-81** are cell surface antigens that are expressed in pluripotent human ES/iPS cells and not on mouse ES/iPS cells. Both antibodies recognize different proteoglycan epitopes on the same protein, podocalyxin⁷.
- **Dapi** or 4',6-diamidino-2-phenylindole is a fluorescent dye that binds strongly to A-T rich regions in DNA and is thus frequently used to label the cell nucleus.

EMD Millipore's Fluorescent Human ES/iPS Cell Characterization Kit contains a range of sensitive tools for the phenotypic assessment of the pluripotent status of human ES/iPS cells. Included in the kit is an enzymatic assay to measure alkaline phosphatase activity in the cells along with validated directly conjugated antibodies to pluripotent transcription factors, Oct-4, Sox-2 and Nanog and cell surface epitopes TRA-1-60 and TRA-1-81 to enable rapid immunocytochemical marker analysis. The Dapi nuclear dye is conveniently included to aid in cell quantification. While the expression levels of pluripotent markers are expected to be diminished upon differentiation, each possess specific expression kinetics. For example, it has been noted that upon differentiation, Oct-4 and TRA-1-60 expressions are the first to be down-regulated while Nanog and alkaline phosphatase down-regulate at a much slower timeframe⁷.

For Research Use Only; Not for use in diagnostic procedures

Kit Components

1. Fast Red Violet solution (Part No. 90239): One 15 mL bottle.
2. Naphthol AS-BI phosphate solution (Part No. 90234). One 15 mL bottle.
3. Mouse anti-Oct-4 (POU5f1), clone 7F9.2, Alexa Fluor® 488 conjugate (Part No. MAB4419A4-50UL). One vial containing 50 µL of 0.5 mg/mL conjugated monoclonal antibody.
4. Mouse anti-Sox-2, clone 10H9.1, Cy3 conjugate (Part No. MAB4423C3-50UL). One vial containing 50 µL of 0.5 mg/mL conjugated monoclonal antibody.
5. Mouse anti-Nanog, clone 7F7.1, Alexa Fluor® 488 conjugate (Part No. MABD24A4-50UL). One vial containing 50 µL of 0.5 mg/mL conjugated monoclonal antibody.
6. Mouse anti-TRA-1-60, clone TRA-1-60, Cy3 conjugate (Part No. MAB4360C3-50UL). One vial containing 50 µL of 0.5 mg/mL conjugated monoclonal antibody.
7. Mouse anti-TRA-1-81, clone TRA-1-81, Cy3 conjugate (Part No. MAB4381C3-50UL). One vial containing 50 µL of 0.5 mg/mL conjugated monoclonal antibody.
8. DAPI, 100 µL (Part No. 90229). One vial containing 100 µL volume.

Related Products

The following related products are available from EMD Millipore as separate items.

1. ES Cell Characterization Kit (Cat. No. SCR001)
2. ES Cell Marker Sample Kit (Cat. No. SCR002)
3. Alkaline Phosphatase Detection Kit (Cat. No. SCR004).
4. Quantitative Alkaline Phosphatase ES Characterization Kit (Cat. No. SCR066)
5. Anti-OCT-4 [POU5F1], clone 7F9.2, Alexa Fluor® 488 conjugate, 100 µL (Cat. No. MAB4419A4)
6. Anti-OCT-4 [POU5F1], clone 7F9.2, Cy3 conjugate, 100 µL (Cat. No. MAB4419C3)
7. Anti-OCT-4, clone 10H11.2, Alexa Fluor® 488 conjugate, 100ul (Cat No. MAB4401A4)
8. Anti-OCT-4, clone 10H11.2, Cy3 conjugate, 100ul (Cat No. MAB4401C3)
9. Anti-NANOG, clone 7F7.1, Alexa Fluor® 488 conjugate, 100 µL (Cat. No. MABD24A4)
10. Anti-NANOG, clone 7F7.1, Cy3 conjugate, 100 µL (Cat. No. MABD24C3)
11. Anti-TRA-1-60, clone TRA-1-60, Cy3 conjugate, 100 µL (Cat. No. MAB4360C3)
12. Anti-TRA-1-60, clone TRA-1-60, Alexa Fluor® 488 conjugate, 100 µL (Cat. No. MAB4360A4)

13. Anti-TRA-1-81, clone TRA-1-81, Cy3 conjugate, 100 μ L (Cat. No. MAB4381C3)
 14. Anti-TRA-1-81, clone TRA-1-81, Alexa Fluor® 488 conjugate conjugate, 100 μ L (Cat. No. MAB4381A4)
 15. Anti-Sox-2, clone 10H9.1, Cy3 conjugate, 100 μ L (Cat. No. MAB4423C3)
 16. Anti-Sox-2, clone 10H9.1, Alexa Fluor® 488 conjugate, 100 μ L (Cat. No. MAB4423A4)
-

Materials Not Supplied

1. Tissue culture-wares and supplies
 2. Fixative (e.g. 4% Paraformaldehyde in 1X PBS)
 3. Millicell EZ SLIDE 8-well glass, sterile (Cat. No. PEZGS0896)
 4. Phosphate-Buffered Saline (1X PBS) (Cat. No. BSS-1005-B)
 5. 1X Rinse Buffer (e.g. TBST: 20 mM Tris-HCL, pH 7.4, 0.15M NaCl, 0.05% Tween 20)
 6. Blocking Solution (3% normal goat or donkey serum, 0.2% Triton X-100, 0.05% NaN_3 in 1X PBS)
 7. Non-Permeable Blocking Solution (3% normal goat or donkey serum in 1X PBS).
 8. Antifading mounting solution (DABCO/PVA)
 9. Microscope
-

Storage

The Fluorescent Human ES/iPS Cell Characterization Kit contains two components used for alkaline phosphatase activity determination as well as 5 ES cell-specific antibodies and a nuclear staining dye. When stored at 2° to 8°C, the kit components are good for 4 months from date of receipt. Do not freeze or expose to elevated temperatures.

Preparation of Reagents

1. Naphthol/Fast Red Violet Solution: Mix Fast Red Violet (FRV) with Naphthol AS-BI phosphate solution and water in a 2:1:1 ratio (FRV:Naphthol:water) fresh before each staining assay.

Staining Protocol

Alkaline Phosphatase Staining Procedure:

1. Culture human ES/iPS cells for three to five days prior to analyzing AP activity. (NOTE: This time-period is critical to be able to observe good levels of AP activity).
2. Aspirate the media and fix the human ES/iPS cells with a fixative (e.g. 4% paraformaldehyde in 1X PBS) for 1-2 minutes.

***Note:** Do not overfix. Fixing cells longer than 2 minutes will result in the inactivation of alkaline phosphatase.*
3. Aspirate the fixative and rinse with 1X Rinse Buffer. DO NOT allow the cells to dry.
4. Prepare reagents for Alkaline Phosphatase staining as described in “Preparation of Reagents” section.
5. Add enough stain solution to cover each well (e.g. 2 mL for a well of a 6-well plate). Incubate in the dark at room temperature for 15 minutes.
6. Aspirate the staining solution and rinse the wells with 1X Rinse Buffer. Cover the cells with 1X PBS to prevent drying and then count the number of colonies expressing AP (red stem cell colonies), versus the number of differentiated colonies (colorless).
7. AP staining criteria: Greater than 90% of colonies should remain undifferentiated and express alkaline phosphatase.

Immunofluorescent Staining Procedure:

For optimal results, cell staining should be performed on cell colonies that have been in culture for approximately 3-5 days after passaging. TRA-1-60 Cy3 and TRA-1-81 Cy3 conjugates can be used to stain live human ES/hiPS cells. In the case of live staining, skip steps 4 – 6 and go directly to step 7. **Note:** Do not add NaN_3 to the blocking solution for live cell staining.

1. Culture human ES or iPS cells in a 6-well plate in human ESC expansion media of choice. The staining protocol will work similarly using feeder or feeder-free media systems so please follow the manufacturer's instructions for specific media.
2. Remove the media from the wells. Be careful to not aspirate the cells.
3. Rinse once with 1X PBS then aspirate.
4. Add 4% Paraformaldehyde (PFA, diluted in 1X PBS) to each well. Incubate for 15-30 minutes at room temperature.
5. Carefully aspirate the PFA from the wells. Be careful to not aspirate the cells.

6. Wash three times with 1X PBS (~2-3 minutes per wash). At this point the fixed cells can be stored in 1X PBS at 4°C for a couple of weeks if necessary.
7. Aspirate the 1X PBS. Apply a blocking solution for 30 – 60 minutes at room temperature or overnight at 4°C . **IMPORTANT: Do not shake the cells.** For optimal results, use the Blocking Solution (3% Normal Goat or Donkey Serum, 0.2% Triton X-100, and 0.05% NaN₃ in 1X PBS) with antibodies directed against intracellular gene targets, Oct-4, Sox-2, and Nanog. Use the Non-Permeable Blocking Solution (3% Normal Goat or Donkey Serum in 1X PBS) with antibodies directed against cell surface epitopes TRA-1-60 and TRA-1-81.
8. Before the end of the incubation time, prepare 1:100 dilutions of the conjugated antibodies in the appropriate blocking buffer (protected from light).
9. Aspirate the blocking buffer. Be careful to not aspirate the cells.
10. Add the 1:100 diluted antibodies to the designated well(s). Incubate for 1-2 hours at room temperature. Cover the plate(s) with tin foil to protect from the light.
11. Aspirate to remove the antibodies. Be careful to not aspirate the cells.
12. Wash three times with 1X PBS (3-4 minutes per wash).
13. Prepare the DAPI dye. Dilute the DAPI in 1X PBS at 1:1000 dilution.
14. Remove the last wash, add DAPI staining solution and incubate at room temperature for 5-10 minutes.
15. Remove the DAPI solution; wash three times with 1X PBS (3-4 minutes per wash).
16. If cell staining is on plates, cells should be covered with 1X PBS for visualization. However, if using glass coverslips, mount the coverslip onto glass slides using anti-fading mounting solution (e.g. DABCO/PVA).
17. Visualize the cell staining using a fluorescence microscope.
Note: Be sure to use the correct filter when visualizing fluorescent-labeled cells.

Results

The following are representative results obtained by using the Fluorescent Human ES/iPS Cell Characterization Kit on pluripotent H9 Human ESCs and human iPSCs.

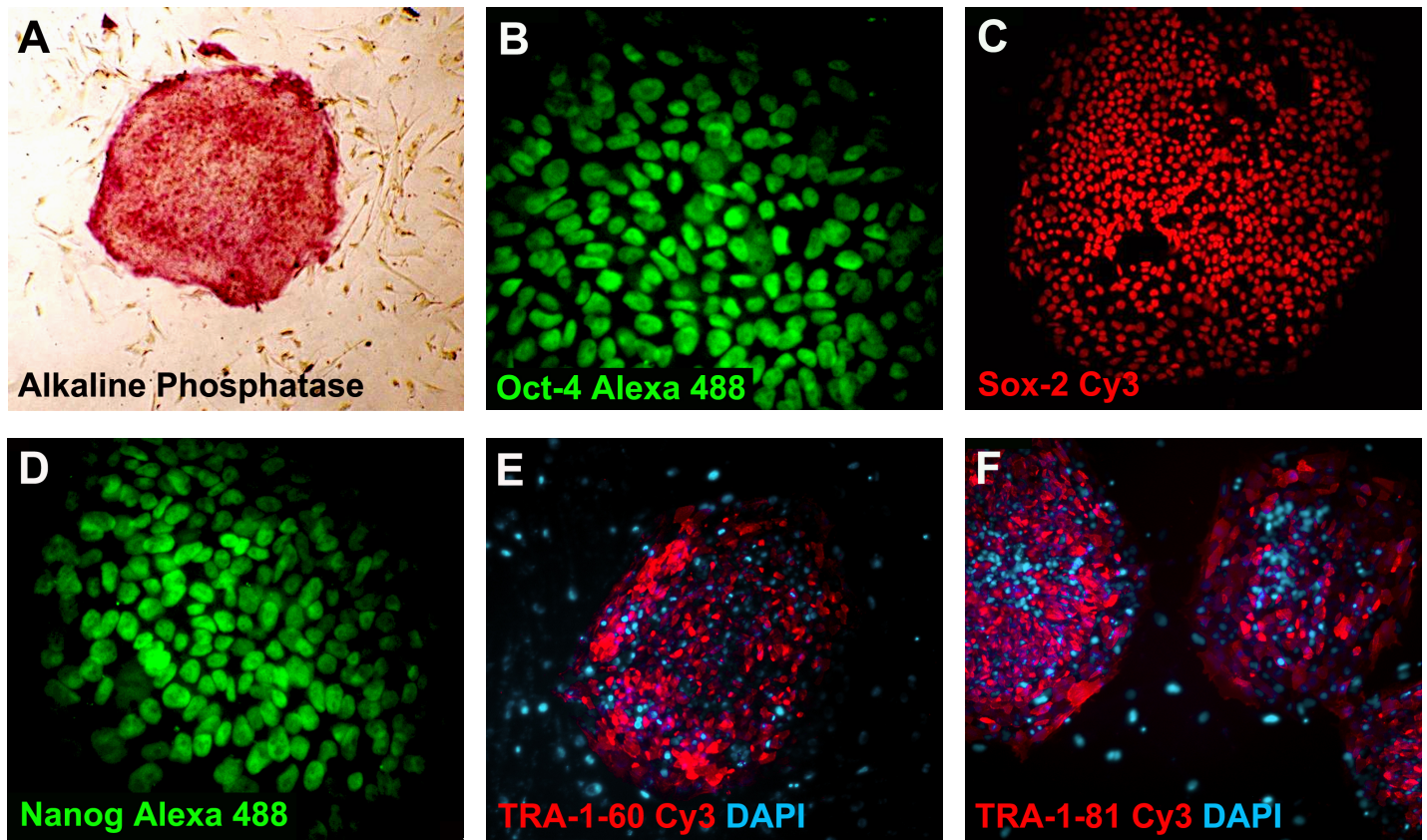


Figure 1. Pluripotent hES/iPS cells express pluripotent markers, alkaline phosphatase (40x) (A), Oct-4 Alexa 488 (400x) (B), Sox-2 Cy3 (100x) (C), Nanog Alexa 488 (400x) (D), TRA-1-60 Cy3 (100x) (E), and TRA-1-81-Cy3 (100x) (F). All conjugated antibodies were used at 1:100 dilutions. Nuclei were counterstained with DAPI (blue). Human foreskin fibroblasts were reprogrammed using the Human STEMCCA Cre-Excisable Constitutive Polycistronic (OKSM) Lentivirus Reprogramming Kit (Cat. No. SCR545).

References

1. Thomson, J. A., Kalishman, J., Golos, T. G., Durning, M., Harris, C. P., Becker, R. A., and Hearn, J. P. (1995) Isolation of a primate embryonic stem cell line. *Proc. Natl. Acad. Sci. USA* **92(17)**: 7844-8.
2. Yu, J., Vodyanik, M. A., Smuga-Otto, K., Antosiewicz-Bourget, J., Frane, J. L., Tian, S., Nie, J., Jonsdottir, G. A., Ruotti, V., Stewart, R., Slukvin, I. L., and Thomson, J. A. (2007) Induced pluripotent stem cell lines derived from human somatic cells. *Science* **318(5858)**: 1917-20.
3. Thomson, J. A., Itskovitz-Eldor, J., Shapiro, S. S., Waknitz, M. A., Swiergiel, J. J., Marshall, V. S., and Jones, J. M. (1998) Embryonic stem cell lines derived from human blastocytes. *Science* **282(5391)**: 1145-7.
4. Takahashi, K. and Yamanaka, S. (2006) Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* **126(4)**: 663-76.
5. Chen, L. and Daley, G. Q. (2008) Molecular basis of pluripotency. *Hum. Mol. Genet.* **17(R1)**: R23-7.
6. Natunen, S., Satomaa, T., Pitkanen, V., Salo, H., Mikkola, M., Natunen, J., Otonkoski, T., and Valmu, L. (2011) The binding specificity of the marker antibodies TRA-1-60 and TRA-1-81 reveals a novel pluripotency-associated type 1 lactosamine epitope. *Glycobiology* **21(9)**: 1125-30.
7. Ramirez, J. M., Gerbal-Chaloin, S., Milhavet, O., Qiang, B., Becker, F., Assou, S., Lemaitre, J. M., Hamamah, S., De Vos, J. (2011) Brief report: benchmarking human pluripotent stem cell markers during differentiation into the three germ layers unveils a striking heterogeneity: all markers are not equal. *Stem Cells* **29(9)**: 1469-74.

Warranty

EMD Millipore Corporation (“EMD Millipore”) warrants its products will meet their applicable published specifications when used in accordance with their applicable instructions for a period of one year from shipment of the products. **EMD MILLIPORE MAKES NO OTHER WARRANTY, EXPRESSED OR IMPLIED. THERE IS NO WARRANTY OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE.** The warranty provided herein and the data, specifications and descriptions of EMD Millipore products appearing in EMD Millipore’s published catalogues and product literature may not be altered except by express written agreement signed by an officer of EMD Millipore. Representations, oral or written, which are inconsistent with this warranty or such publications are not authorized and if given, should not be relied upon.

In the event of a breach of the foregoing warranty, EMD Millipore Corporation’s sole obligation shall be to repair or replace, at its option, the applicable product or part thereof, provided the customer notifies EMD Millipore Corporation promptly of any such breach. If after exercising reasonable efforts, EMD Millipore Corporation is unable to repair or replace the product or part, then EDM Millipore shall refund to the Company all monies paid for such applicable Product. **EMD MILLIPORE CORPORATION SHALL NOT BE LIABLE FOR CONSEQUENTIAL, INCIDENTAL, SPECIAL OR ANY OTHER DAMAGES RESULTING FROM ECONOMIC LOSS OR PROPERTY DAMAGE SUSTAINED BY ANY COMPANY CUSTOMER FROM THE USE OF ITS PRODUCTS.**

Unless otherwise stated in our catalog or other company documentation accompanying the product(s), our products are intended for research use only and are not to be used for any other purpose, which includes but is not limited to, unauthorized commercial uses, in vitro diagnostic uses, ex vivo or in vivo therapeutic uses or any type of consumption or application to humans or animals.

Alexa Fluor is a registered trademark of Life Technologies. Cy3 is a registered trademark of GE Healthcare.

(c) 2009 - 2012: Merck KGaA, Darmstadt. All rights reserved. No part of these works may be reproduced in any form without permission in writing

Cat No. SCR078

July 2012
Revision B