

(Catalog Number: CYL4039)

CATALOG NUMBER:

CYL4039 QUANTITY:

200 units (assayed with [³H]-astemizole)

Note: this quantity is equivalent to 400 units assayed with [¹²⁵I]-BeKm.

LOT NUMBER: VOLUME/CONCENTRATION: 1 mL, 2 mg/mL

BACKGROUND: The human ether-a-go-go related gene (hERG) is a potassium ion channel which is essential for normal cardiac repolarization. In drug screening models, the hERG K⁺ channel has been indicated to inhibit a wide variety of compounds, and its blockage can lead to cardiac QT interval prolongation and life threatening arrhythmias (Murphy *et al.* 2006). Cardiac safety relating to I_{Kr} K⁺ channels has been identified as the firmest link to QT prolongation (Chiu *et al.* 2004). Millipore's hERG membrane preparations are crude membrane preparations made from HEK293 stable recombinant cell lines (Millipore cat. # CYL3039), which is ideal HTS tools for screening antagonists against the hERG channel. The membrane preparations exhibit a K_d of 5.2 nM for [³H]-Astemizole. With 10 µg/well hERG Membrane Prep and 3.0 nM [³H]-Astemizole, a greater than 4-fold signal-to-background ratio was obtained.

APPLICATIONS:

Radioligand binding assay

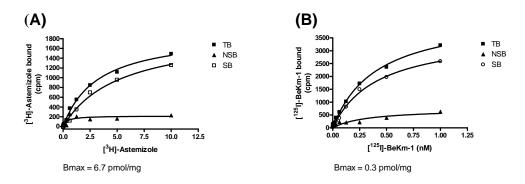
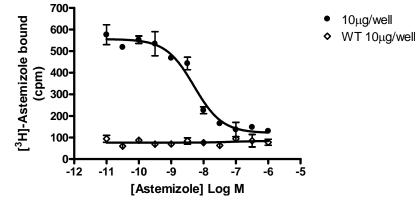


Figure 1. Saturation binding for hERG with [³**H]-astemizole and [**¹²⁵**I]-BeKm.** (A), one unit of hERG membrane preparation was incubated with increasing amount of [³H]-astemizole in the absence (total binding, TB) or presence (nonspecific binding, NSB) of 500-fold excess unlabeled astemizole. (B), half a unit of hERG Membrane Preparation was incubated with increasing amount of [¹²⁵I]-BeKm-1 in the absence or presence of 500-fold excess unlabeled BeKm-1. In both curves, specific binding (SB) was determined by subtracting NSB from TB.



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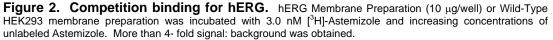


Table 1. Signal: background and specific binding values obtained in a competition binding assay for the hERG membrane prep.

	10µg/well
Signal: background	4.6
Specific binding (cpm)	435

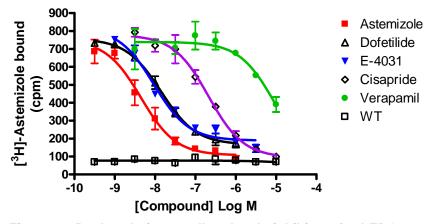


Figure 3. Rank ordering small molecule inhibitors for hERG. hERG Membrane Preparation (10μ g/well) was characterized by evaluating the activity for known hERG small molecule inhibitors in a competition binding assay. The membranes were incubated with 3.0 nM [³H]-Astemizole and increasing concentrations of unlabeled compounds to determine sample activity and rank order.

Table 2. Rank order comparison of various small molecule hERG inhibitors using [³H]-



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astemizole binding assay and E-Phys assays:

	Binding Ki (nM) with hERG membrane preps		Electrophysiology IC50 (nM) with hERG cell lines		
	Millipore CYL4039M value	Literature value*	Millipore CYL4039 by IonWorks PPC	Millipore CYL4039 by PatchExpress	Literature value by Patch Clamp
Astemizole	3.4 ± 0.7	3.3 ± 0.7	4.6 ± 0.003	ND	0.5
Dofetilide	5.8 ± 2.4	28 ± 6	ND	ND	15.3 ± 2.5
E-4031	6.1 ± 0.8	58 ± 5	115 ± 0.02	144 ± 44	14
Pimozide	18.8 ± 2.3	14 ± 4	ND	ND	18
Cisapride	104.9 ± 30.9	123 ± 25	75.6 ± 0.02	5 ± 1	44.5 ± 10.6
Haloperidol	241.4 ± 21.4	234 ± 20	ND	ND	63
Risperidone	> 4000	4302 ± 319	ND	ND	167
Verapimil	> 4000	3902 ± 529	1332 ± 120	3100 ± 400	830

* Data obtained from Chiu et al., 2004

(**A**)

(B)

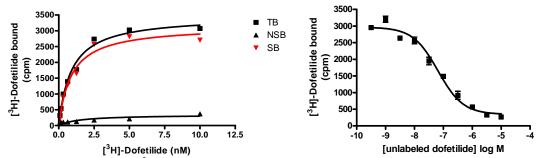


Figure 4. Binding of [³**H]-dofetilide to hERG membrane preparation.** (A), five units (50 µg) of hERG membrane preparation was incubated with increasing amount of [³H]-Dofetilide (American Radiolabeled Chemicals) in the absence (total binding, TB) or presence (nonspecific binding, NSB) of 1000-fold excess unlabeled Dofetilide. A Kd of 0.9 nM and Bmax of 0.84 pmol/mg were obtained. (B), five units (50 µg) of hERG membrane preparation was incubated with 5 nM [3H]-dofetilide and increasing concentrations of unlabeled dofetilide.

SPECIFICATIONS: 1 unit = 10 μ g membrane preparation with [³H]-astemizole B_{max} with [³H]-astemizole: 6.7 pmol/mg K_d for [³H]-astemizole: 5.2 nM

Species: Human ERG (Accession number U04270)

HOST CELLS: HEK293

RECOMMENDED ASSAY CONDITIONS: Membranes are mixed with radioactive ligand and unlabeled competitor (see Figures 1 and 2 for concentrations tested) in binding buffer in a nonbinding 96-well plate, and incubated for 1-2 h. Prior to filtration, an FB 96-well harvest



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plate (Millipore cat. # MAHF B1H) is coated with 0.33% polyethyleneimine for 30 min, then washed with 25 mM Tris, pH 7.4, 130 mM NaCl, 5 mM KCl, 0.8 mM CaCl₂, 0.1% BSA. Binding reaction is transferred to the filter plate, and washed 6 times (250μ L per well per wash) with Wash Buffer. The plate is dried and counted.

Binding buffer: 10 mM Hepes, pH 7.4, 130 mM NaCl, 5 mM KCl, 0.8 mM MgCl₂, 1 mM NaEGTA, 10 mM glucose, 0.1% BSA, filtered and stored at 4° C

Radioligand: [³H]-Astemizole (Perkin Elmer # NET-1140)

Wash Buffer: 25 mM Tris, pH 7.4, 130 mM NaCl, 5 mM KCl, 0.8 mM MgCl₂, 0.05 mM CaCl₂, 0.1% BSA, filtered and stored at 4°C.

One package contains enough membranes for at least 200 assays (units), where a unit is the amount of membrane that will yield greater than 4-fold signal: background with ³H-labeled Astemizole.

PRESENTATION: Liquid in packaging buffer: 50 mM Tris pH 7.4, 10% glycerol and 1% BSA with no preservatives. Packaging method: Membrane protein was adjusted to the indicated concentration in packaging buffer, rapidly frozen, and stored at -80°C.

STORAGE/HANDLING: Store at –70°C. Product is stable for at least 6 months from the date of receipt when stored as directed. Do not freeze and thaw.

REFERENCES: Ackerman MJ *et al.* (1998) The long QT syndrome: ion channel diseases of the heart. *Mayo Clin. Proc.* 73: 250-269.

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Chouabe C *et al.* (1998) HERG and KvLQT1/IsK, the cardiac K⁺ channels involved in long QT syndromes, are targets for calcium channel blockers. *Mol. Pharmacol.* 54: 695-703.

Murphy SM *et al.* (2006) Evaluation of functional and binding assays in cells expressing either recombinant or endogenous hERG channel. *J. Pharmacol. Toxicol. Methods* 54: 42-55.

Rampe D *et al.* (1997) A mechanism for the proarrhythmic effects of cisapride (Propulsid): high affinity blockade of the human cardiac potassium channel HERG. *FEBS Lett.* 417: 28-32.

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