

# **ProductInformation**

### SIGMA QUALITY CONTROL TEST PROCEDURE

# Enzymatic Assay of DEOXYRIBONUCLEASE II (EC 3.1.22.1)

### PRINCIPLE:

Deoxyribonuclease II > 3'-Oligodeoxyribonucleotides

Abbreviation used:

DNA = Deoxyribonucleic Acid

**CONDITIONS:** T = 25°C, pH 4.6,  $A_{260nm}$ , Light path = 1 cm

**METHOD:** Continuous Spectrophotometric Rate Determination

### **REAGENTS:**

A. 1 M Sodium Acetate Buffer
 (Prepare 100 ml in deionized water using Sodium Acetate, Trihydrate, Sigma Prod.
 No. S-8625. Adjust to pH 4.6 at 25°C with 6 N HCl.)

- B. 20 mM Magnesium Sulfate Solution (MgSO<sub>4</sub>)
   (Prepare 25 ml in deionized water using Magnesium Sulfate, Heptahydrate, Sigma Prod. No. M-1880.)
- C. 0.033% (w/v) Deoxyribonucleic Acid Solution (DNA)
   (Prepare 30 ml in deionized water using Deoxyribonucleic Acid, Sodium Salt, Sigma Prod.
   No. D-1501. Gently mix by stirring at 4°C for 15 24 hours.)
- D. 150 mM Sodium Chloride Solution (NaCl)
   (Prepare 25 ml in deionized water using Sodium Chloride, Sigma Prod. No. S-9625.)
- E. Standard DNase Solution (Std Soln)<sup>1</sup> (Immediately before use, reconstitute a vial of Deoxyribonuclease II, Standardized vial, containing approximately 1500 Kunitz units, Sigma Prod. No. D-9784, with 2 ml of cold deionized water.)
- F. Deoxyribonuclease II Enzyme Solution (DNase II) (Immediately before use, prepare a solution containing 500 1000 Kunitz units/ml in cold deionized water.)

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# PROCEDURE:

Prepare a reaction cocktail by pipetting (in milliliters) the following reagents into a suitable container:

Reagent A (Buffer)	7.50
Reagent B (MgSO <sub>4</sub> )	3.75
Reagent C (DNA)	9.00
Deionized water	54.75

Mix by swirling and equilibrate to 25°C.

Pipette (in milliliters) the following reagents into suitable cuvettes:

	<u>Test</u>	<u>Std</u>	Blank
Reaction Cocktail	2.50	2.50	2.50
Reagent D (NaCl)	0.40	0.40	0.40

Mix by inversion and equilibrate to 25°C. Monitor the A<sub>260nm</sub> until constant, using a suitably thermostatted spectrophotometer. Then add:

Reagent E (Std Soln)		0.10	
Reagent F (DNase II)	0.10		
Deionized Water			0.10

Immediately mix by inversion and record the increase in A<sub>260nm</sub> for approximately 10 minutes. Obtain the  $\Delta A_{260nm}$ /minute using the maximum linear rate for the Test, Standard, and Blank.

# **CALCULATIONS:**

Units/ml Test Solution = 
$$\frac{(\Delta A_{260nm}/min \text{ Test - } \Delta A_{260nm}/min \text{ Blank})(3)(df)}{(0.001)(0.1)}$$
Units/ml Std Solution = 
$$\frac{(\Delta A_{260nm}/min \text{ Std - } \Delta A_{260nm}/min \text{ Blank})(3)(df)}{(0.001)(0.1)}$$

3 = Volume (in milliliters) of assay

df = Dilution factor

 $0.001 = \Delta A_{260nm}$  per unit as per the Unit Definition

0.1 = Volume (in milliliter) of enzyme used

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**CALCULATIONS:** (continued)

CF = Correction factor

Experimental Units/ml Std Soln

750 = Theoretical Units/ml Std Soln

Corrected Units/ml enzyme = (CF)(Units/ml Test Solution)

# **UNIT DEFINITION:**

One Kunitz unit will produce a  $\Delta A_{260}$  of 0.001 per minute per ml at pH 4.6 at 25°C. [Mg<sup>++</sup>] = 0.83 mM

#### FINAL ASSAY CONCENTRATION:

In a 3.00 ml reaction mix, the final concentrations are 83 mM sodium acetate, 0.83 mM magnesium sulfate, 0.0033% (w/v) deoxyribonucleic acid, 20 mM sodium chloride, and 50 - 100 units deoxyribonuclease II.

#### **REFERENCES:**

Kunitz, M. (1950) Journal of General Physiology 33, 349-362

Kunitz, M. (1950) Journal of General Physiology 33, 363-377

Lindberg, U. (1964) Biochimica et Biophysica Acta 82, 237-248

### NOTES:

There is no absolute standard for the assay of DNase. When the procedure of Kunitz is used, the result is affected by the particular lot of substrate used. For DNase studies, we offer a standard DNase II vial, Sigma Prod. No. D-9784, which has been standardized to contain approximately 1500 Kunitz units (actual contents listed on the label) using Deoxyribonucleic Acid, Sodium Salt, Sigma Prod. No. D-1501 as substrate.

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### NOTES:

- 2. This assay is based on the cited references.
- 3. Where Sigma Product or Stock numbers are specified, equivalent reagents may be substituted.

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