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Product Information

AURORA A, active, GST-tagged, human PRECISIO[®] Kinase recombinant, expressed in *Sf*9 cells

Catalog Number **A1983** Lot Number 061M0745 Storage Temperature –70 °C

Synonyms: AURKA, AIK, ARK1, AURA, BTAK, STK6, STK7, STK15, AURORA2, MGC34538

Product Description

AURORA A belongs to a multigenic family of mitotic serine/threonine kinases, which are involved in the control of chromosome segregation. AURORA A is involved in centrosome separation, duplication, and maturation as well as in bipolar spindle assembly and stability.¹ AURORA A is expressed and active at the highest level during G₂-M phase of the cell cycle. Overexpression of AURORA A has been correlated with the grade of various human solid tumors. Ectopic AURORA A overexpression in any culture cell line leads to polyploidy and centrosome amplification.²

This recombinant product was expressed by baculovirus in *Sf*9 insect cells using an N-terminal GST-tag. The gene accession number is NM 003600. It is supplied in 50 mM Tris-HCI, pH 7.5, with 150 mM NaCI, 0.25 mM DTT, 0.1 mM EGTA, 0.1 mM EDTA, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~72 kDa

Purity: ≥70% (SDS-PAGE, see Figure 1)

Specific Activity: 62-84 nmole/min/mg (see Figure 2)

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at -70 °C is recommended. After opening, aliquot into smaller quantities and store at -70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Lot Number 061M0745: >75% (densitometry)

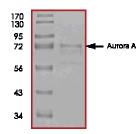
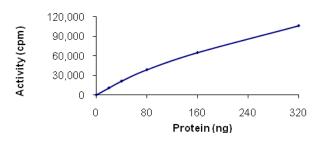


Figure 2.

Specific Activity of Lot Number 061M0745: 62 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl₂, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 $ng/\mu I$ BSA solution.

Kinase Solution – Dilute the active AURORA A $(0.1 \ \mu g/\mu)$ with Kinase Dilution Buffer to the desired concentration.

<u>Note</u>: The lot-specific specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active AURORA A kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μl aliquots at –20 °C.

 γ -³²P-ATP Assay Cocktail (250 μ M) – Combine 5.75 ml of Kinase Assay Buffer, 150 μ l of 10 mM ATP Stock Solution, 100 μ l of γ -³²P-ATP (1 mCi/100 μ l). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Dissolve the synthetic peptide substrate myelin basic protein (MBP) in water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

<u>Kinase Assay</u>

This assay involves the use of the ³²P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active AURORA A, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ -³²P-ATP Assay Cocktail may be thawed at room temperature.
- In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl: 10 μl of Kinase Solution
 - $5 \,\mu$ l of Substrate Solution
 - 5 μ l of cold water (4 °C)
- Set up a blank control as outlined in step 2, substituting 5 μl of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ -³²P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- 5. After the 15 minute incubation, stop the reaction by spotting 20 μ l of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- 6. Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ^{-32} P-ATP counts introduced into the reaction. Spot 5 µl of the γ^{-32} P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR = $cpm of 5 \mu l of \gamma^{-32}P-ATP Assay Cocktail nmole of ATP$

cpm – value from control (step 7) nmole – 1.25 nmole (5 μl of 250 μM ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\frac{\Delta \text{cpm} \times (25/20)}{\text{SR} \times \text{E} \times \text{T}}$$

SR = specific radioactivity of the ATP (cpm/nmole ATP) \triangle cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

- 20 = spot volume
- T = reaction time (minutes)
- E = amount of enzyme (mg)

References

- 1. Dutertre, S. et al., On the role of aurora-A in centrosome function. Oncogene, **21**, 6175-6183 (2002).
- Katayama, H. et al., The Aurora kinases: role in cell transformation and tumorigenesis. Cancer Metastasis Rev., 22, 451-464 (2003).

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