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Product Information

NUAK2, active, GST tagged, human PRECISIO® Kinase recombinant, expressed in *Sf*9 cells

Catalog Number **SRP5054** Storage Temperature –70 °C

Synonyms: SNARK, FLJ90349, DKFZP434J037, DKFZp686F01113

Product Description

NUAK2 or SNF1/AMP kinase-related kinase (SNARK) is a member of the NUAK family of SNF1-like kinase 2. NUAK2 is activated by muscle contraction and is a unique mediator of contraction-stimulated glucose transport in skeletal muscle. NUAK2 is involved in cellular stress responses linked to obesity and type 2 diabetes. NUAK2 shows kinase activity against a synthetic test peptide and the activity in keratinocytes is increased by AMP and 5-amino-4-imidazole-carboxamide riboside, implying AMPK kinase-dependent pathway can activate NAUK2. Glucose deprivation also increases NAUK2 activity in baby hamster kidney fibroblasts.

Recombinant full-length human NUAK2 was expressed by baculovirus in *Sf9* insect cells using an N-terminal GST tag. The NUAK2 gene accession number is NM_030952. Recombinant protein stored in 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM glutathione, 0.1 mM EDTA, 0.25 mM DTT, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~110 kDa

Purity: 70-95% (SDS-PAGE, see Figure 1)

Specific Activity: 68–92 nmole/min/mg (see Figure 2)

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at -70 °C is recommended. After opening, aliquot into smaller quantities and store at -70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1. SDS-PAGE Gel of Typical Lot 70–95% (densitometry)

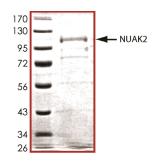
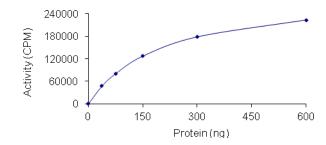


Figure 2.Specific Activity of Typical Lot 68–92 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl₂, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/μl BSA.

Kinase Solution – Dilute the active NUAK2 (0.1 μ g/ μ l) with Kinase Dilution Buffer to the desired concentration. Note: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active NUAK2 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μ l aliquots at –20 °C.

 γ -³³P-ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of γ -³³P-ATP (1 mCi/100 μl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Dissolve the synthetic peptide substrate in distilled water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ³³P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active NUAK2, Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ -³³P-ATP Assay Cocktail may be thawed at room temperature.
- 2. In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl:

10 μ l of Kinase Solution 5 μ l of Substrate Solution

5 μl of cold water (4 °C)

- 3. Set up a blank control as outlined in step 2, substituting 5 μ l of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ - 33 P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- 5. After the 15 minute incubation, stop the reaction by spotting 20 μ l of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ - 33 P-ATP counts introduced into the reaction. Spot 5 μ l of the γ - 33 P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR =
$$\frac{\text{cpm of 5} \, \mu \text{l of } \gamma^{-33}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

cpm – value from control (step 7) nmole – 1.25 nmole (5 μl of 250 μM ATP Assav Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\Delta$$
cpm × (25/20)
SR × E × T

SR = specific radioactivity of the ATP (cpm/nmole ATP) Δ cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

20 = spot volume

T = reaction time (minutes)

E = amount of enzyme (mg)

References

- Lefebvre, D.L. et al., Identification and characterization of a novel sucrose-non-fermenting protein kinase/AMP-activated protein kinase-related protein kinase, SNARK. Biochem. J., 355, 297-305 (2001).
- Rune, A. et al., Regulation of skeletal muscle sucrose, non-fermenting 1/AMP-activated protein kinase-related kinase (SNARK) by metabolic stress and diabetes. Diabetologia, 52, 2182-2189 (2009).

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